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(54) METHODS AND COMPOSITIONS FOR TREATING AND IDENTIFYING COMPOUNDS TO TREAT AGE-RELATED MACULAR DEGENERATION TREATMENT

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CPC A61K 31/198 (2013.01); A61K 31/00 (2013.01); A61K 31/07 (2013.01); A61K 31/195 (2013.01); A61K 31/355 (2013.01); A61K 31/375 (2013.01); A61K 33/30 (2013.01); A61K 33/34 (2013.01); A61K 45/06 (2013.01); A61K 49/0008 (2013.01); G01N 33/5023 (2013.01); G01N 33/5041 (2013.01); G01N 2333/4704 (2013.01); G01N 2333/726 (2013.01); G01N 2500/10 (2013.01)

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See application file for complete search history.

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ABSTRACT

The present invention provides methods for treating or limiting development of age-related macular degeneration, as well as methods for identifying compound suitable for such use.

20 Claims, 9 Drawing Sheets

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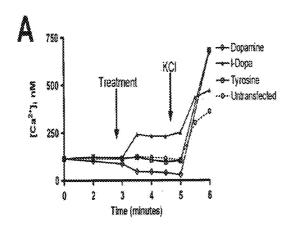
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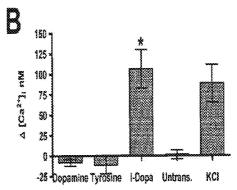
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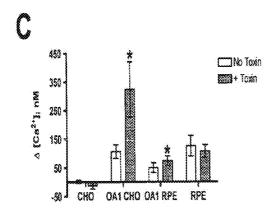
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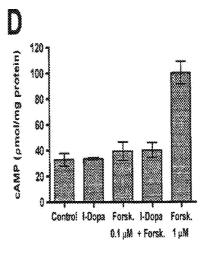
Figure 1 B OA1 OA1 OA1-GFE Actin Actio In Situ 500μM Tyrosine 1μM Tyrosine 500µM Tyrosine tµM Tyrosine D 10000 Standardized Total DA1 500 7600 400 300 5000 2500 100 L.T LT Control Control

Figure 2









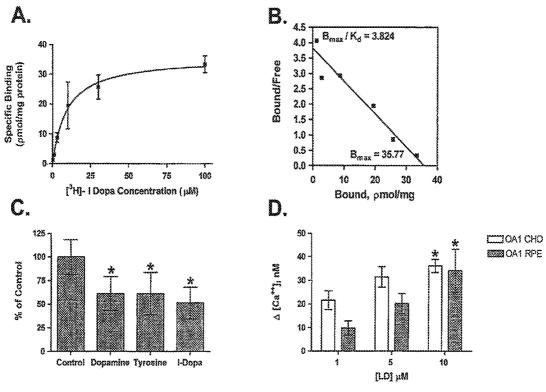


FIGURE 3

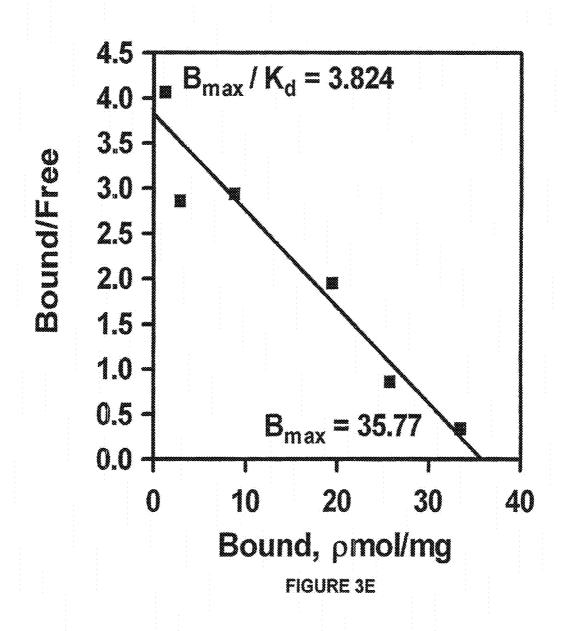


Figure 4

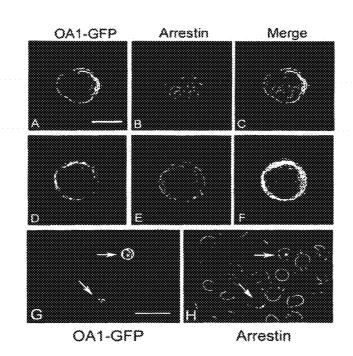
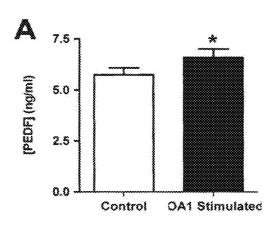
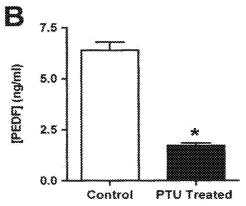


Figure 5





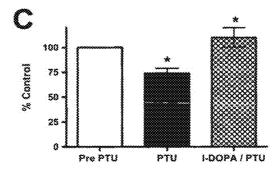
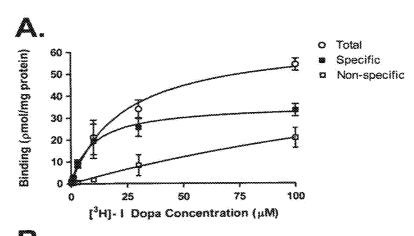


Figure 6



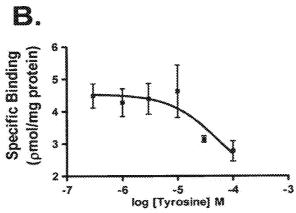
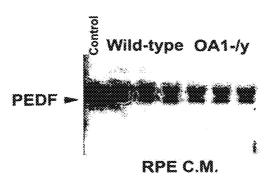


Figure 7



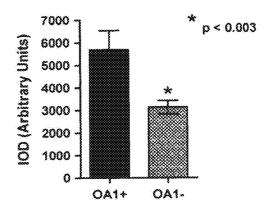
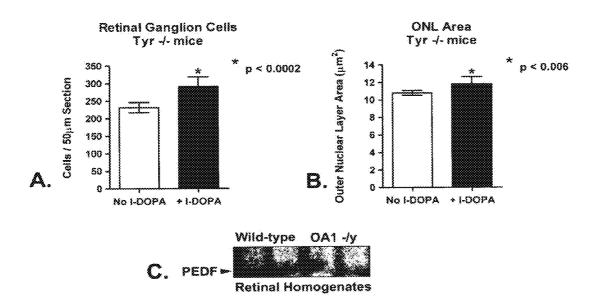


Figure 8



METHODS AND COMPOSITIONS FOR TREATING AND IDENTIFYING COMPOUNDS TO TREAT AGE-RELATED MACULAR DEGENERATION TREATMENT

RELATED APPLICATIONS

This application is a divisional of U.S. patent application Ser. No. 12/937,669 filed Nov. 5, 2010, which is a U.S. national phase filing of PCT/US09/041021 filed Apr. 17, 10 2009, which claims priority to U.S. Provisional Patent Application Ser. No. 61/124,624, filed Apr. 18, 2008, each of which is incorporated by reference herein in its entirety.

STATEMENT OF GOVERNMENT RIGHTS

This invention was made with government support under National Institutes of Health, Grant Number R03 EY014403. The government has certain rights in the invention

BACKGROUND

Age-related macular degeneration ("AMD") is an aging-associated disease resulting in the loss of vision in the 25 macula (the center of the visual field) because of damage to the retina. AMD is a prevalent disorder of the aged, with approximately 10% of patients 66 to 74 years and 30% of patients 75 to 85 years of age having some level of macular degeneration. Currently there is no effective treatment available for most patients with AMD, and no early stage intervention.

SUMMARY OF THE INVENTION

In one aspect, the present invention provides methods for treating age-related macular degeneration (AMD), comprising administering to a subject with AMD an amount effective for treating AMD of an agonist of the OA1 receptor. In a second aspect, the present invention provides methods for 40 limiting development of AMD, comprising administering to a subject at risk of developing AMD an amount effective for limiting development of AMD of an agonist of the OA1 receptor. In one preferred embodiment of either of these aspects of the invention, the agonist of the OA1 receptor is 45 selected from the group consisting of L-DOPA and L-DOPA analogues.

In another aspect, the present invention provides methods for identifying compounds to treat AMD, comprising contacting cells with a test compound, wherein the cells comprise:

- (a) a first cell population expressing OA1; and, optionally,
- (b) a second cell population not expressing OA1; and
- (c) identifying as positive test compounds those test compounds that increase one or both of
 - (i) pigment epithelium-derived factor (PEDF) expression in the first cell population relative to one or both (A) PEDF expression in the first population of cells not contacted with the test compound, and (B) the second cell population, and
 - (ii) intracellular calcium concentration in the first cell population relative to one or both (A) intracellular calcium concentration in the first population of cells not contacted with the test compound, and (B) the second cell population;

wherein the positive test compounds are candidate compounds for treating and/or limiting development of AMD.

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In a further aspect, the present invention provides methods for identifying compounds to treat AMD, comprising

- (a) administering a test compound to a tyrosinase deficient pregnant female non-human mammal, wherein the test compound is administered during embryonic photoreceptor and/ or retinal ganglion development; and
- (b) comparing an effect of the test compound on photoreceptor and/or retinal ganglion development in the embryo or post-natal non-human mammal, to photoreceptor and/or retinal ganglion development in an embryo or post-natal non-human mammal not administered the test compound, wherein those test compounds that increase photoreceptor and/or retinal ganglion development are candidate compounds for treating and/or limiting development of AMD.

In a still further aspect, the invention provides compositions comprising:

- (a) an amount effective of L-DOPA or an L-DOPA analogue for treating or limiting development of AMD; and
- (b) an amount effective for treating or limiting development of AMD of a composition comprising a source of vitamin C, a source of vitamin E, a source of vitamin A, a source of zinc, and a source of copper.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1(A-C) Western blot analysis of proteins bound (B) or unbound (U) to strepavidin-conjugated beads after biotinylation of RPE in situ, cultured RPE (b), or COS cells transfected to express OA1-GFP (c). Blots were probed to visualize OA1 and actin after cell surface biotinylation and fractionation using streptavidin-conjugated beads. For cultured cells (b, c) cells were either maintained in 500 µM (normal DMEM) or 1 µM tyrosine for 3 days prior to analysis. (D) Quantification of western blot analysis by densitometry. OA1 densitometry is shown as the % of the control for paired cell cultures, transfected then split into 2 equal groups, one of which was the control, maintained in normal DMEM (control). The other group was maintained in 1 μM tyrosine DMEM (LT) until harvest. Paired t-test analysis was used to test whether the difference was significant, and * denotes p<0.001. Actin, analyzed the same way showed no differences, and p=0.724. (E-F) Composite confocal microscopy of pigmenting RPE cells maintained in normal DMEM (e) or 1 µM tyrosine (f) then stained with anti-OA1 antibodies and imaged at 20x. Bar=25 μm.

FIG. 2(A) Representative traces of [Ca²⁺]i during the time course of the standard experimental protocol in transfected and untransfected CHO cells. After establishment of a stable baseline for 3 minutes, the test agent was added at 1 µM. At 5 minutes, KCl was added to serve as a control that the cells were Fura-2 loaded and patent. Identical protocols were performed for both transfected cells and paired untransfected cells. (B) Summary data for [Ca2+]i in response to tyrosine, dopamine, and L-DOPA in transfected and untransfected CHO cells. Untransfected cells are shown with L-DOPA treatment. The experimental control of membrane depolarization with KCl is also shown. Each bar represents data collected from at least 10 experiments and is presented as the mean change from baseline [Ca²⁺]i after test agent 60 addition. Error bars represent S.D., and t-test analyses were used to test for significant differences, * denotes p<0.01. Analysis of pertussis toxin sensitivity of [Ca²⁺]i increase in cells transfected to express OA1 or RPE that express the natural protein. Data represent mean of at least 6 experiments. (C) Analysis of pertussis toxin sensitivity of [Ca²⁺]i increase in cells transfected to express OA1 or RPE that express the natural protein. Data represent mean of at least

culture.

6 experiments for each group of transfected cells and 20 individual experiments for each the treated and untreated RPE with endogenous OA1 expression. T-tests analyses were used to test for significant differences, and * denotes p<0.01. (D) cAMP was measured in CHO transfected to 5 express OA1. The control group represents transfected but untreated CHO cells and the basal level of cAMP in those cells. Cells were treated with 1.0 µM L-DOPA, 0.1 µM forskolin, L-DOPA+0.1 µM forskolin, and as a positive control 1 µM forskolin. Results represent the mean cAMP levels observed in at least 6 experiments in which all experimental groups were analyzed in a paired fashion using replicate monolayers in the same culture plate. Error bars represent the S.D. of each group, and the only significant difference observed was the increase in cAMP levels after forskolin treatment.

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FIG. 3(A) Binding kinetics between OA1 and L-DOPA were determined using radiolabeled ligand binding assays. Results represent data collected from 5 such experiments and are presented as mean specific binding +/- SEM. The hyperbolic curve fit exhibited an R² value of 0.994, Kd was determined to be $9.34 \times 10^{-6} \text{M} + /1.14 \times 10^{-6} \text{M}$. (B) Comparative binding of 5 μM [H³] L-DOPA to OA1 transfected CHO cells was compared in the presence of 1.0 mM dopamine, tyrosine, or L-DOPA. The data represent mean total binding 25 +/-S.D. for each group. * denotes p<0.05 when comparing the results between the control group to the binding in the presence of the potential competitive ligands. (C) Competitive interaction between 5 μM [H³] L-DOPA and dopamine were assessed to determine whether dopamine functions as 30 an antagonist of OA1 activity. Results indicate that dopamine and L-DOPA compete for the same OA1 binding site, and the data fits the binding model with an r^2 value of 0.95. The Ki for dopamine was 2.388 +/-0.266 µM (mean+/-SEM), similar to the Kd for L-DOPA. (D) Dose-dependent 35 OA1 signaling through OA1. Data represent mean increase in [Ca²⁻], elicited by L-DOPA treatment of the cells at the concentrations given (n=6 for each dose). T-test analyzes were used to compare between the responses achieved at each dose, and * denotes p<0.01 for the comparison at 1 and $10 \mu M$.

FIG. 3(E) Scatchard plot illustrating the kinetics of a single site binding relationship based on FIG. 3(a).

FIG. **4**(A-H) All images represent 2 μm thick confocal sections of CHO cells transfected to express OA1-GFP. β -arrestin was visualized using immuno fluorescence methods. Prior to addition of L-DOPA (a-c) and after treatment with 1 μM L-DOPA (d-f), and the merged images (c, f) illustrate regions where the two proteins co-localize, at the resolution of white light imaging. (g,h) are low magnification of field of transfected CHO cells, with two transfected cells visible (arrows) (g). The remainder of the cell population is visualized using antibodies to β -arrestin (h) to illustrate that β -arrestin recruitment to the membrane only occurred in the OA1 expressing cells (arrows).

FIG. 5 (A) PEDF concentrations were determined by ELISA of cell conditioned medium. RPE cells were control cells, without L-DOPA treatment, or OA1 stimulated cells that were treated with 1 μM L-DOPA prior to being maintained for 3 days in normal DMEM. Data are presented as the mean of 3 experiments conducted in triplicate, error bars represent S.D, and * denotes P<0.01 using a paired t-test. (B) PEDF concentrations in conditioned medium from pigmenting RPE determined by ELISA. Cells were either control pigmenting RPE cultures or paired cultures treated with phenylthiourea (PTU) at 200 μM. Data are presented as the mean of 3 experiments conducted in triplicate, error bars represent S.D, and * denotes P<0.01 using a paired t-test. (C) PEDF concentrations in conditioned medium of pigmented

RPE cells treated with PTU then treated with L-DOPA to stimulate OA1 signaling. ELISA assays were conducted prior to PTU treatment, then after PTU treatment, and then from the same cultures after L-DOPA stimulation. Results are presented as mean+/-S.D. of the value achieved related to that culture of cells. * denotes p<0.01 when comparing PTU to the control (same culture tested prior to PTU), and L-DOPA/PTU compared to the PTU sample from that same

FIG. 6(A) Data represents mean+/-SEM bound [3H]-L-DOPA in all fractions, total, specific and non-specific. Nonspecific binding was determined by measuring radiolabeled-L-DOPA bound in the presence of excess unlabeled L-DOPA (1 mM). Specific binding at each given concentration is determined by subtracting the measured nonspecific binding from the measured total binding. (B) The figure illustrates competitive interaction between tyrosine and L-DOPA, measured using increasing concentrations of tyrosine and 5 μM [H³] L-DOPA. Each data point represents the mean data from 5 replicate wells, and the error bars are S.D. Data illustrate that tyrosine competes for binding with L-DOPA, but with a low affinity. The results suggest tyrosine has a Ki of 52.9 μM, and fits the single site binding model with an r² value of 0.85. Saturation could not be achieved because of the limited solubility of tyrosine.

FIG. 7 Western blot and graphical representation of PEDF secretion in wild-type vs OA deficient mice.

FIG. **8**(A) is a graphical representation of data demonstrating that L-DOPA supplementation increases retinal ganglion cell numbers compared to what is expected in a normal wild-type mouse. (B) is a graphical representation of data demonstrating that L-DOPA supplementation increases photoreceptor numbers compared to what is expected in a normal wild-type mouse. (C) is a Western blot showing PEDF detection in 2 wild-type and 2 OA1 –/y mice.

DETAILED DESCRIPTION OF THE INVENTION

All references cited are herein incorporated by reference in their entirety.

Within this application, unless otherwise stated, the techniques utilized may be found in any of several well-known references such as: *Molecular Cloning: A Laboratory Manual* (Sambrook, et al., 1989, Cold Spring Harbor Laboratory Press), *Gene Expression Technology* (Methods in Enzymology, Vol. 185, edited by D. Goeddel, 1991. Academic Press, San Diego, Calif.), "Guide to Protein Purification" in *Methods in Enzymology* (M. P. Deutshcer, ed., (1990) Academic Press, Inc.); *PCR Protocols: A Guide to Methods and Applications* (Innis, et al. 1990. Academic Press, San Diego, Calif.), *Culture of Animal Cells: A Manual of Basic Technique*, 2nd Ed. (R. I. Freshney. 1987. Liss, Inc. New York, N.Y.), *Gene Transfer and Expression Protocols*, pp. 109-128, ed. E. J. Murray, The Humana Press Inc., Clifton, N.J.), and the Ambion 1998 Catalog (Ambion, Austin Tex.)

As used herein, the singular forms "a", "an" and "the" include plural referents unless the context clearly dictates otherwise.

In a first aspect, the present invention provides methods for treating age-related macular degeneration (AMD), comprising administering to a subject with AMD an amount effective for treating AMD of an agonist of the OA1 receptor.

In a second aspect, the present invention provides methods for limiting development of AMD, comprising admin-

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istering to a subject at risk of developing AMD an amount effective for limiting development of AMD of an agonist of the OA1 receptor.

The human Oa1 gene, is found on the X chromosome, and has been shown to encode a 404 amino acid protein OA1 (SEQ ID NO:2), likely to be a G-protein coupled receptor (GPCR) [12,13] based upon sequence analysis [14]. As disclosed in detail herein, the inventors have identified the OA1 signaling pathway as a critical determinant of neurosensory retina survival, such that stimulation of this pathway will provide treatment for AMD as well as a means to limit AMD development for those at potential risk. While not being bound by any mechanism, the inventors believe that OA1 and tyrosinase participate in an autocrine loop through L-DOPA that regulates the secretion of at least one potent neurotrophic factor, PEDF. Thus administration of L-DOPA can be used to stimulate OA1 activity and thus upregulate PEDF expression, making it a valuable therapeutic to treat and limit development of AMD.

As discussed in detail below, such OA1 agonists can be identified, for example, using the drug discovery methods of the third and fourth aspects of the invention. Exemplary OA1 agonists are discussed in detail below.

The subject preferably is a human.

As used herein for all aspects and embodiments of the invention, "AMD" means an aging-associated disease resulting in the loss of vision in the macula (the center of the visual field) because of damage to the retina know as Age-related Macular Degeneration. As used herein, AMD encompasses both wet and dry AMD, described in more detail below.

AMD begins with characteristic drusen (yellow deposits) in the macula between the retinal pigment epithelium and the underlying choroid. Most people with these early changes (referred to as age-related maculopathy) have good vision. People with drusen can go on to develop advanced AMD. The risk is considerably higher when the drusen are large and numerous and associated with disturbance in the pigmented cell layer under the macula.

Subjects with age-related maculopathy may progress to either of the two main forms of advanced AMD, each of which can be treated or be limited in its development using the methods of the invention. "Wet" AMD causes vision loss 45 due to abnormal blood vessel growth in the choriocapillaries, through Bruch's membrane, ultimately leading to blood and protein leakage below the macula. Bleeding, leaking, and scarring from these blood vessels eventually causes irreversible damage to the photoreceptors and rapid vision 50 loss if left untreated. "Dry" AMD occurs when light-sensitive cells in the macula slowly break down, gradually causing vision loss in the affected eye. Blurring in AMD is probably due to the accumulation of drusen under the retinal pigment epithelium (RPE) which alters to focal properties of 55 the photoreceptors by moving them out of the plane of focus.

Dry AMD may occur in one or both eyes, and can advance from age-related maculopathy into intermediate or advanced stages of dry AMD.

Intermediate Dry AMD: Either many medium-sized 60 drusen or one or more large drusen. Some people see a blurred spot in the center of their vision. More light may be needed for reading and other tasks.

Advanced Dry AMD: In addition to drusen, a breakdown of light-sensitive cells and supporting tissue in the central 65 retinal area. This breakdown can cause a blurred spot in the center of vision. Over time, the blurred spot may get bigger

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and darker, taking more of the central vision; may have difficulty reading or recognizing faces until they are very close to you.

AMD symptoms include, but are not limited to blurred/ reduced central vision, central scotomas (shadows or missing areas of vision), trouble discerning one dark color from another dark color and/or one light color from another light color; slow recovery of visual function after exposure to bright light, a loss in contrast sensitivity, so that contours, shadows and color vision are less vivid, retinal pigment epithelial (RPE) disturbance (including pigment clumping and/or dropout), RPE detachment, geographic atrophy, subretinal neovascularization, and disciform scar, and distorted vision (metamorphopsia), such that a grid of straight lines appears wavy and parts of the grid may appear blank Symptoms of dry AMD and wet AMD are generally similar early during disease progression, and thus it may not be possible to determine which early-stage patients will develop dry vs. wet forms of AMD. Dry AMD develops as 20 'geographic atrophy', and early AMD become 'wet' AMD when new blood vessels sprout.

As used herein, "treat" or "treating" AMD means accomplishing one or more of the following: (a) reducing the severity of AMD; (b) limiting or preventing development of one or more symptoms characteristic of AMD, as described above; (c) inhibiting worsening of one or more symptoms characteristic of AMD, as described above; (d) limiting or preventing recurrence of AMD in patients that have previously had the disorder(s); and (e) limiting or preventing recurrence of one or more symptoms in patients that were previously symptomatic for AMD. Such treating includes treating of wet AMD and dry AMD.

As used herein, the term "limiting development of" AMD means to prevent or to minimize development of AMD in individuals at risk of developing AMD, as well as limiting progression of age-related maculopathy to AMD (wet or dry), or intermediate dry AMD to advanced dry or 'wet' AMD. In one preferred embodiment, the methods comprise treating a subject with drusen accumulation (ie: age-related maculopathy), to limit development of AMD. In another preferred embodiment, the methods comprise treating a subject with an amount effective of the OA1 agonist to decrease the rate of lines of loss of vision relative to a non-treated AMD subject, or subject at risk of AMD. In another preferred embodiment, the methods comprise treating a subject with wet AMD, or at risk of developing wet AMD, an amount effective of the OA1 agonist to decrease the rate and number of new blood vessel formation. As discussed in more detail below, OA1 stimulation causes the RPE to increase PEDF secretion, and PEDF is a potent anti-angiogenic factor. Thus, OA1 stimulation strategies may stop new blood vessel development in 'wet' AMD, in addition to its effects on retinal development discussed herein.

In another preferred embodiment, the methods comprise treating a subject that has blurred or reduced central vision with an amount of OA1 agonist effective to increase the lines of visual acuity in one or both eyes. In this embodiment, the lines of visual acuity are as measured by the standard Snellen test, where the increase or decrease in 'lines' of visual acuity are based on which smallest 'line' on a Snellen chart a patient can read clearly.

"Subjects at risk of developing AMD" mean anyone with any risk factor for development of AMD, including but not limited to being over 50 years old (in various preferred embodiments, over 60 years old, over 65 years old, over 70 years old, or over 75 years old), presence of drusen deposits, Caucasian race, having a blood relative that has or had AMD, a mutation in the complement factor H gene (CFH) of (Tyr402His), Arg80Gly variant of the complement protein C3 gene, hypertension, high cholesterol levels, obesity, smoking, a high fat intake, and mutations in the fibulin 5 gene. Thus, in a preferred embodiment, the subject to be treated has one or more of these risk factors, particularly in methods for limiting development of AMD.

The phrase "therapeutically effective amount," as used herein, refers to an amount that is sufficient or effective to limit development of or treat (prevent the progression of or reverse) AMD. The appropriate dosage range depends on the choice of the compound, the route of administration, the nature of the formulation, the nature of the subject's condition, and the judgment of the attending practitioner. For example, oral administration would be expected to require higher dosages than administration by intravenous injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization, as is well 20 understood in the art.

In a preferred embodiment, the OA1 receptor agonist comprises a compound selected from the group consisting of L-DOPA and L-DOPA analogues.

L-DOPA is [2-amino-3-(3,4-dihydroxyphenyl)propanoic ²⁵ acid] known for use in treating Parkinson's, and has the following structure.

L-DOPA is commercially available and methods for its synthesis are known to those of skill in the art.

As used herein, "L-DOPA analogues" are those L-DOPA 40 variants that retain OA1-stimulatory activity, including L-DOPA prodrugs, of which many are known in the art; exemplary such analogues are disclosed below. While not being bound by a specific mechanism of action, the inventor believes that L-DOPA binding to OA1 involves two sites of 45 binding, one involving one or both hydroxyl groups, and one involving the carboxylic acid group. In one embodiment, the L-DOPA analogues are L-DOPA prodrugs that are metabolized to L-DOPA after administration (and generally prior to binding to OA1 on the cell surface), and thus are expected 50 to retain OA1-stimulatory activity. In another embodiment, one or both hydroxyl group and/or the carboxyl group can be substituted to produce various analogues (prodrug or otherwise) for use in the methods of the invention.

In another embodiment, the L-DOPA analogues comprise 55 L-DOPA esters Exemplary L-DOPA esters, and methods for preparing them, are disclosed in WO/1997/016181; U.S. Pat. No. 4,663,349; 4,873,263, 4,873,263; 5,345,885, and 4,771, 073. In various preferred embodiments, the L-DOPA ester is selected from the group consisting of L-DOPA methyl ester, 60 L-DOPA butyl ester, L-DOPA pentyl ester, L-DOPA cyclohexyl ester, L-DOPA benzyl ester, and L-DOPA ethyl ester. In various further preferred embodiments, the L-DOPA esters are selected from the alkyl, aryl and substituted and unsubstituted aralkyl esters of L-DOPA. In a further preferred embodiment, the L-DOPA esters are represented by the following formula:

wherein R is a straight or branched chain alkyl (C_1 - C_{20}) such as methyl, ethyl, propyl, butyl, myristyl, palmityl, pentyl, tetradecyl, hexadecyl and the like; aryl(C₆-C₉) such as phenyl, tolyl and the like; substituted and unsubstituted mono, di or polyhydroxyalkyl(C₁-C₂₀) such as benzyl, alkoxybenzyl, 4-hydroxybutyl, 2-hydroxypropyl, 2,3-dihydroxypropyl, 1,3-dihydroxypropyl, 6-hydroxyhexyl and 5-hydroxypentyl and the like optionally having a substituent such as alkoxy(C_{1-5}) [methoxy, ethoxy, butoxy and the like]; carbalkoxy (C₁₋₅) [methoxycarbonyl, ethoxycarbonyl, propoxycarbonyl butoxycarbonyl and the likel; amino; mono or dialkylamino(C_{1-10}) [methylamino, methylethylamino, diethylamino and the like]; acylamino(C₁₋₅) [acetamido, butyramido and the like]; ketoalkyl (C₁₋₅) [methylketo, ethylketo, butylketo and the like]; halo [chloro, bromo and the like] or carboxamide; substituted and unsubstituted $aralkyl(C_{7-20})$ such as benzyl, alkoxybenzyl C_{8-14}) [methoxy, ethoxy, isobutoxy and the like]; phenylethyl; phenylpropyl; phenylbutyl; phenylhexyl; phenyloctyl and the like; and pharmaceutically acceptable organic or inorganic counterion salts.

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 H_2N

Synthetic processes for preparing the esters of L-DOPA and the salts thereof are known in the art, for example, in U.S. Pat. Nos. 3,891,696; 4,035,507; and 5,354,885; and Journal of Pharmaceutical Sciences, 62, p. 510 (1973), each incorporated by reference herein in their entirety.

In another embodiment, the L-DOPA analogues comprise bile acid conjugates as are known in the art. Exemplary L-DOPA bile acid conjugates, and methods for preparing are disclosed in WO/2002/028882 US20020151526. Upon oral administration, these prodrugs are cleaved within the enterohepatic system to release the parent drug and/or an active metabolite from the bile acid into the systemic circulation. Significantly, only a fraction (typically <50%)<50%) of the prodrug is cleaved during each pass through the enterohepatic cycle. Thus, the enterohepatic circulation serves as a reservoir of the drug enabling sustained systemic drug levels to be achieved. Naturally occurring bile acids such as cholic acid, chenodeoxycholic acid, ursodeoxycholic acid, deoxycholic acid, ursocholic acid and lithocholic acid are particularly preferred. The site of conjugation of these bile acids to L-DOPA or other L-DOPA analogue is preferably via the 3-hydroxy group or the C-24 carboxyl moiety. Optionally, cleavable linker functionality may be introduced between the drug and the bile acid and this linker may be selected. In a preferred embodiment, such L-DOPA bile acid conjugates are represented by the following formula

$$R^2$$

wherein R1 is selected from the group consisting of hydrogen and OH;

R2 is selected from the group consisting of hydrogen and OH;

X is selected from the group consisting of OH and D-Y—, where Y is selected from the group consisting of a covalent bond and a cleavable linker group covalently connecting D to the steroid;

D is a member selected from the group consisting of L-DOPA and other L-DOPA analogues;

W is selected from the group consisting of (a) a substituted alkyl group containing a moiety which is negatively charged at physiological pH, which moiety is selected from the group consisting of —COOH, —SO₃H, —SO₂H, —P(O)(OR6)(OH), —OP(O)(OR6)(OH), —OSO₃H and the like and pharmaceutically acceptable salts thereof.

where R6 is selected from the group consisting of alkyl, substituted alkyl, aryl and substituted aryl; and (b) a group 20 of the formula -M-Y'-D'

where M is selected from the group consisting of —CH₂OC (O)— and —CH₂CH₂C(O)—;

Y' is a covalent bond or a cleavable linker group covalently connecting D' to M;

D' is a member selected from the group consisting of L-DOPA and other L-DOPA analogues;

with the proviso that either X is —Y-D and/or W is -M-Y'-D' wherein the compound of formula (I) above is a substrate for an intestinal bile acid transporter;

or a pharmaceutically acceptable salt thereof.

In another embodiment, the L-DOPA analogues comprise di or tri-peptide derivatives. Exemplary L-DOPA di- or tri-peptide analogues, and methods for preparing them, are disclosed in U.S. Pat. No. 3,803,120 and 5,686,423. Oral absorption of the di- and tri-peptide L-DOPA prodrugs show high oral bioavailability with some compounds having the plasma concentration 60-100 fold higher than that of L-dopa. In a preferred embodiment, such L-DOPA prodrugs are represented by the following formula

$$R$$
 NH_2
 R_1
 R_2
 R_2
 R_2
 R_3
 R_4
 R_4
 R_5
 R_4
 R_5
 R_5
 R_5
 R_5

wherein n is 0 or 1; R is hydrogen or hydroxyl, preferably R is hydroxyl;

R1 is hydrogen; and

R2 is hydrogen, alkyl of from one to four carbon atoms, alkyl of from one to four carbon atoms substituted with 60 one —OH, —SH, —SCH₃, —NH₂, —NHC(=NH)NH₂, —COOH, phenyl, hydroxyphenyl, indolyl or imidazolyl group, alkyl from one to four carbon atoms substituted with one carboalkoxyl group of from one to six carbon atoms, preferably R2 is hydrogen, methyl or hydroxym-65 ethyl; or

R1 and R2 together are trimethylene.

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Preferably, R1 and R2 of the di- or tri-peptide derivative of L-DOPA (2-amino-3-(3,4-dihydroxyphenyl-)propanoic acid) of the formula (I) together is trimethylene.

In another embodiment, di-peptide derivatives of L-DOPA [2-amino-3-(3,4-dihydroxyphenyl)propanoic acid] are represented by the following formula

HOOC
$$\stackrel{R_4}{\underset{R_3}{\bigvee}}$$
 $\stackrel{O}{\underset{NH_2}{\bigvee}}$ $\stackrel{OH}{\underset{OH}{\bigvee}}$

wherein R3 is hydrogen; and R4 is phenyl or hydroxyphenyl; or R3 and R4 together is trimethylene.

In another embodiment, the L-DOPA analogues comprise amine prodrugs as are known in the art. Exemplary L-DOPA amine analogues, and methods for preparing them, are disclosed in US20060025385 and WO/2004/069146. In one preferred embodiment, such L-DOPA amine analogues are represented by

$$R_6O$$
 $*$
 R_3
 R_4
 R_3
 R_3

wherein *C denotes a chiral carbon;

R1, R2, R3 and R4 are each independently selected from the group consisting of hydrogen, alkyl having 1-30 carbon atoms, alkenyl having 1-30 carbon atoms, alkynyl having 1-30 carbon atoms, cycloalkyl, aryl, O-carboxy, C-carboxy, carbonyl, thiocarbonyl, 0-carbamyl, 0-thiocarbamyl and a fatty acid acyl, or, alternatively, R1 and R2 and/or R3 and R4 form a five- or six-membered ring; and

R5 and R6 are each independently selected from the group consisting of hydrogen, alkyl, cycloalkyl, aryl and phosphonyl,

or a pharmaceutically acceptable salt thereof.

Preferred L-DOPA amine analogues include: compounds wherein R5 and R6 are each hydrogen; compounds wherein R1 and R2 are each hydrogen; compounds wherein R3 and R4 are each hydrogen; compounds wherein at least one of R1, R2, R3 and R4, preferably R3 and/or R4 is carbonyl, e.g., acetyl. Additional preferred compounds according to the present embodiments include compounds wherein at least one of R1, R2, R3 and R4 is an alkyl, alkenyl or alkynyl having 1-30 carbon atoms, or, alternatively, at least one of R1, R2, R3 and R4 is a fatty acid acyl, derived from, for example, myristic acid, lauric acid, palmitic acid, stearic acid, oleic acid, arachidonic acid, linoleic acid or linolenic acid. Further preferred examples of L-DOPA amine analogues according to the present embodiments include α -amino-3,4-dihydroxy-benzenepropanamide, α -N-acetyl-3,4-dihydroxy-benzenepropanamide and pharmaceutically acceptable salts thereof

In a further preferred embodiment, L-DOPA prodrugs for use in the present invention, and methods for their synthesis, are disclosed in U.S. Pat. Nos. 4,065,566 and 4,035,507 and are represented by the formula

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wherein each R is independently selected from the group consisting of a hydrogen atom, an acyl group, a

group, a —CO-pyridyl group, and a —CO—R3 group, wherein R3 represents the residue of any N,N—C1-C2 dialkylamino acid or a C4-C6 cycloalkylamino acid

wherein R1 represents a member selected from the group consisting of a hydroxyl group and a —OM group, wherein M is an alkali metal (Na, K, etc.) or an ammonium ion; and wherein R2 represents a member selected from the group 40 consisting of a

group,

a —CO-pyridyl group, and a —CO—R3 group, wherein R3 represents the residue of any N,N—(C1-C2)-dialkylamino acid or a C4-C6-cycloalkylamino acid

$$(CH_2)_5$$
 N δ

Further L-DOPA prodrugs for use in the present invention, and methods for their synthesis, disclosed in U.S. Pat. 65 Nos. 4,065,566 and 4,035,507 are represented by the formula

$$R_2$$
OR
OR
OR
OR

wherein R represents an acyl group; wherein R2 represents a hydrogen atom; and wherein R1 represents a —NHCH (R4)COOR5 group, wherein R4 represents the residue of any naturally occurring amino acid, and wherein R5 represents a member selected from the group consisting of a hydrogen atom, a C1-C5 alkyl group (e.g., methyl, ethyl, propyl, butyl, pentyl), and a C1-C5 alkylaryl group (e.g., —CH2—C6H5, —CH2—CH2—C6H5, etc.), and the HX salts thereof, wherein X is a conventional pharmaceutically acceptable acid addition salt anion (e.g., chloride, bromide, perchlorate, methanesulfonate, succinate, etc.);

Preferred exemplary L-DOPA prodrugs disclosed in U.S. Pat. Nos. 4,065,566 and 4,035,507 include the following:

- Glycyl-3,4-diacetyloxy-L-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- Glycyl-3,4-diacetyloxy-L-phenylalanine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3. 3,4-diacetyloxy-L-phenylalanyl-glycine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 4. N-nicotinoyl-3,4-dihydroxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- 5. N-nicotinoyl-3,4-diacetyloxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- N-nicotinoyl-3,4-dipivalyloxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- 3,4-diacetyloxy-L-phenylalanyl-glycine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 8. 3,4-diacetyloxy-L-phenylalanyl-glycine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3,4-diacetyloxy-L-phenylalanyl-glycine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3,4-diacetyloxy-L-phenylalanyl-glycine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3,4-diacetyloxy-L-phenylalanyl-L-leucine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3,4-diacetyloxy-L-phenylalanyl-L-leucine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 13. 3,4-diacetyloxy-L-phenylalanyl-L-leucine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 14. 3,4-diacetyloxy-L-phenylalanyl-L-leucine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.

- 15. 3,4-diacetyloxy-L-phenylalanyl-L-isoleucine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3,4-diacetyloxy-L-phenylalanyl-L-isoleucine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 17. 3,4-diacetyloxy-L-phenylalanyl-L-isoleucine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 18. 3,4-diacetyloxy-L-phenylalanyl-L-isoleucine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 19. 3,4-diacetyloxy-L-phenylalanyl-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 3,4-diacetyloxy-L-phenylalanyl-phenylalaninemethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 21. 3,4-diacetyloxy-L-phenylalanyl-phenylalanine-ethyl 20 ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 22. 3,4-diacetyloxy-L-phenylalanyl-phenylalanine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- Glycyl-3,4-diacetyloxy-L-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 24. Glycyl-3,4-dipivalyloxy-L-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- Glycyl-3,4-diacetyloxy-L-phenylalanine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- Glycyl-3,4-diacetyloxy-L-phenylalanine-ethyl ester 35 and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 27. Glycyl-3,4-diacetyloxy-L-phenylalanine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 28. L-leucyl-3,4-diacetyloxy-L-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion
- L-leucyl-3,4-diacetyloxy-L-phenylalanine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 30. L-leucyl-3,4-diacetyloxy-L-phenylalanine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 31. L-leucyl-3,4-diacetyloxy-L-phenylalanine-benzyl 50 ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 32. L-isoleucyl-3,4-diacetyloxy-L-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 33. L-isoleucyl-3,4-diacetyloxy-L-phenylalanine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- L-isoleucyl-3,4-diacetyloxy-L-phenylalanine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 35. L-isoleucyl-3,4-diacetyloxy-L-phenylalanine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 36. Phenylalanyl-3,4-diacetyloxy-L-phenylalanine and its 65 HX salt, wherein X represents a pharmaceutically acceptable anion.

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- 37. Phenylalanyl-3,4-diacetyloxy-L-phenylalanine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 38. Phenylalanyl-3,4-diacetyloxy-L-phenylalanine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 39. Phenylalanyl-3,4-diacetyloxy-L-phenylalanine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 40. 3,4-diacetyloxy-L-phenylalanyl-3,4-diacetyloxy-L-phenylalanine and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 41. 3,4-diacetyloxy-L-phenylalanyl-3,4-diacetyloxy-L-phenylalanine-methyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 42. 3,4-diacetyloxy-L-phenylalanyl-3,4-diacetyloxy-L-phenylalanine-ethyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- 43. 3,4-diacetyloxy-L-phenylalanyl-3,4-diacetyloxy-L-phenylalanine-benzyl ester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- N-[N,N-dimethylamino]-glycyl-3,4-diacetyloxy-Lphenylalanine and its M salt, wherein M represents an alkali metal.
- 45. N-nicotinoyl-3,4-dinicotinoyloxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- 46. N-3-pyridylacetyl-3,4-dihydroxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- 47. N-3-pyridylacetyl-3,4-diacetyloxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- 3,4-N,N-dimethylaminoglycyl-L-phenylalanine methylester and its HX salt, wherein X represents a pharmaceutically acceptable anion.
- N[N,N-dimethylamino]glycyl-3,4-[N,N-dimethylaminoglycyl]-L-phenylalanine and its M salt, wherein M represents an alkali metal.
- N[N,N-diethylaminoglycyl]-3,4-diacetyloxy-L-phenylalanine and its M salt, wherein M represents an alkali metal.

As used herein, the term "alkyl" refers to a saturated aliphatic hydrocarbon including straight chain and branched chain groups. The alkyl group preferably has between 1 and 30 carbon atoms, more preferably between 1 and 20 carbon atoms. While lower alkyls, e.g., of between 1 and 6 carbon atoms may facilitate the formulation of the compounds, higher alkyls provides for enhanced permeability thereof through the BBB.

The alkyl group, according to the present invention, may be substituted or non-substituted. When substituted, the substitutent group can be, for example, cycloalkyl, alkenyl, aryl, heteroaryl, heteroalicyclic, hydroxy, alkoxy, aryloxy, thiohydroxy, thioalkoxy, thioaryloxy, halo, carboxy, alkoxy-carbonyl, thiocarboxy, carbamyl, and amino, as these terms are defined herein.

As used herein, the term "cycloalkyl" refers to an all-carbon monocyclic or fused ring (i.e., rings which share an adjacent pair of carbon atoms) group wherein one of more of the rings does not have a completely conjugated pielectron system. Examples, without limitation, of cycloalkyl groups are cyclopropane, cyclobutane, cyclopentane, cyclopentene, cyclohexane, cyclohexadiene, cycloheptatriene and adamantane. The cycloalkyl group, according to the present invention, may be substituted or non-substituted. When substituted, the substituent group can be, for example, alkyl, cycloalkyl, alkenyl, aryl, heteroaryl, heteroalicyclic, hydroxy, alkoxy, aryloxy, thiohydroxy, thio-

alkoxy, thioaryloxy, halo, carboxy, alkoxycarbonyl, thiocarboxy, carbamyl, and amino, as these terms are defined berein

The term "alkenyl" refers to an alkyl group which consists of at least two carbon atoms and at least one carbon-carbon 5 double bond.

The term "alkynyl" refers to an alkyl group which consists of at least two carbon atoms and at least one carbon-carbon triple bond.

As is discussed above, both the alkenyl and the alkynyl 10 groups preferably have between 1 and 30 carbon atoms.

An "aryl" group refers to an all-carbon monocyclic or fused-ring polycyclic (i.e., rings which share adjacent pairs of carbon atoms) group having a completely conjugated pi-electron system. Examples, without limitation, of aryl 15 groups are phenyl, naphthalenyl and anthracenyl. The aryl group, according to the present invention, may be substituted or non-substituted. When substituted, the substituent group can be, for example, alkyl, cycloalkyl, alkenyl, aryl, heteroaryl, heteroalicyclic, hydroxy, alkoxy, aryloxy, thiohydroxy, thioalkoxy, thioaryloxy, halo, carboxy, alkoxycarbonyl, thiocarboxy, carbamyl, and amino, as these terms are defined herein.

The term "C-carboxy" refers to a+C(=O)—OR' group, where R' is hydrogen, alkyl, cycloalkyl, alkenyl, aryl, heteroaryl (bonded through a ring carbon) or heteroalicyclic (bonded through a ring carbon) as defined herein.

The term "O-carboxy" refers to a R'—C(=O)—O—group, where R' is hydrogen, alkyl, cycloalkyl, alkenyl, aryl, heteroaryl (bonded through a ring carbon) or heteroalicyclic 30 (bonded through a ring carbon) as defined herein.

The term "carbonyl" refers to a —C(=O)—R' group, where R' is as defined hereinabove.

The term "thiocarbonyl" refers to a —C(=S)—R' group, where R' is as defined hereinabove.

An "O-carbamyl" group refers to an —OC(—O)—NR'R" group, where R' is as defined hereinabove and R" is as defined for R'.

An "O-thiocarbamyl" group refers to an —OC(=S)—NR'R" group, where R' is and R" are as defined hereinabove. 40

A "fatty acid acyl" refers to a R"C(=O)—O— group, where R" is a saturated or unsaturated hydrocarbon chain having at least 10 carbon atoms.

The term "alkoxy" refers to both an —O-alkyl and an —O-cycloalkyl group, as defined hereinabove. Representa- 45 tive examples of alkoxy groups include methoxy, ethoxy, propoxy and tert-butoxy.

The —O-alkyl and the O-cycloalkyl groups, according to the present invention, may be substituted or non-substituted. When substituted, the substituent group can be, for example, 50 cycloalkyl, alkenyl, aryl, heteroaryl, heteroalicyclic, hydroxy, alkoxy, aryloxy, thiohydroxy, thioalkoxy, thioaryloxy, halo, carboxy, alkoxycarbonyl, thiocarboxy, carbamyl, and amino, as these terms are defined herein.

The term "thioalkoxy" refers to both an —S-alkyl group, 55 and an —S-cycloalkyl group, as defined herein.

The term "hydroxy" refers to an —OH group.

The term "thiohydroxy" refers to an —SH group.

An "aryloxy" group refers to both an —O-aryl and an —O-heteroaryl group, as defined herein.

A "thioaryloxy" group refers to both an —S-aryl and an —S-heteroaryl group, as defined herein.

The term "amino" refers to a —NR'R" group, with R' and R" as defined hereinabove.

The term "alkoxycarbonyl", which is also referred to 65 herein interchangeably as "carbalkoxy", refers to a carboxy group, as defined hereinabove, where R' is not hydrogen.

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The term "heteroaryl" group includes a monocyclic or fused ring (i.e., rings which share an adjacent pair of atoms) group having in the ring(s) one or more atoms, such as, for example, nitrogen, oxygen and sulfur and, in addition, having a completely conjugated pi-electron system. Examples, without limitation, of heteroaryl groups include pyrrole, furane, thiophene, imidazole, oxazole, thiazole, pyrazole, pyridine, pyrimidine, quinoline, isoquinoline and purine.

A "heteroalicyclic" group refers to a monocyclic or fused ring group having in the ring(s) one or more atoms such as nitrogen, oxygen and sulfur. The rings may also have one or more double bonds. However, the rings do not have a completely conjugated pi-electron system.

The term "halo" refers to a fluorine, chlorine, bromine or iodine atom.

The term "phosphonyl" describes an $-P(=O)(OR')_2$ group, with R' as defined hereinabove.

In any embodiment of the first or second aspect of the invention, the methods may comprise administering two or more compounds selected from the group consisting of L-DOPA and L-DOPA analogues. In another preferred embodiment, the methods may further comprise administering a further therapeutic compound to the subject, including but not limited to an L-amino acid decarboxylase inhibitor, such as carbidopa or benserazide. Such L-amino acid decarboxylase inhibitors can be used, for example, to increase plasma half-life of L-DOPA and reduce conversion of L-DOPA to dopamine peripherally, which reduces side effects of L-DOPA treatment. In another embodiment, the methods may further comprise administering one or more other compounds useful for treating or limiting development of AMD, including but not limited to anti-angiogenic therapeutics, such as anti-vascular endothelial growth factor (VEGF) agents, including but not limited to VEGF antibodies (or fragments thereof) such as ranibizumab or bevacizumab, or VEGF aptamers, such as pegaptanib. In another embodiment, the L-DOPA or L-DOPA analogues may be present in a more complex mixture, such as in a nutritional supplement containing L-DOPA or L-DOPA analogues.

In a preferred embodiment, any one or more of the L-DOPA and/or L-DOPA analogues described herein may be used in the form of a dietary supplement. Such a supplement may combine any one or more further components that might be beneficial in treating or limiting development of AMD. In one preferred embodiment, L-DOPA and/or an L-DOPA analogue are combined with a combination of vitamin C source, vitamin E source, Vitamin A source, zinc source, and, and copper source, disclosed in U.S. Pat. No. 6,660,297 as useful in treating AMD; U.S. Pat. No. 6,660, 297 is incorporated by reference herein in its entirety. Any suitable amount of each of these additional components can be used in combination with L-DOPA and/or L-DOPA analogues in carrying out the methods of the invention. In a further preferred embodiment, this combination may further comprise lutein and/or zeaxanthin in an amount suitable to provide further protective retinal effects, preferably between 1 mg and 100 mg; between 1 mg and 50 mg, between 2 mg and 25 mg, or between 2 mg and 10 mg per day. In a further preferred embodiment of any of the above preferred embodiments, this combination may further comprise docosahexaenoic acid (DHA) and/or eicosapentaenoic acid (EPA) in an amount suitable to provide further protective retinal effects, preferably between 250 mg and 1000 mg; between 300 mg and 750 mg, between 350 mg and 750 mg, or between 350 mg and 650 mg per day. The use of such compositions for

treating AMD patients is discussed, for example, at web site www.areds2.org/ and links therein.

Ascorbic acid is the preferred source of vitamin C, although other sources such as for example sodium ascorbate could alternatively be used.

Dl-alpha tocopheryl acetate is the preferred source of vitamin E, although other sources of vitamin E, such as for example trimethyl tocopheryl acetate and/or vitamin E succinate, may be used in the alternative.

Beta-carotene is preferred in the subject composition due 10 to its ready commercial availability although alternative carotenoid proforms of vitamin A could likewise be used.

Zinc is preferred in the form of zinc oxide in subject tablets due to the fact zinc oxide provides the most concentrated form for elemental zinc and is well tolerated in the 15 digestive system. However, other forms of zinc such as for example zinc gluconate may alternatively be used or be used in combination with zinc oxide in the subject composition.

Copper in the form of cupric oxide is preferred in the subject tablets to help prevent zinc induced copper deficiency anemia, although other forms of copper such as for example copper gluconate may alternatively be used or used in combination with cupric oxide in the subject composition.

In a preferred embodiment, the amounts of each of these other components (on a per day basis) is as follows:

between 450 mg and 600 mg vitamin C (approximately 7-10 times the recommended daily allowance (RDA))

between 400 IU and 540 IU vitamin E (approximately 13-18 times the RDA);

between 17.2 mg and 28 mg beta carotene (approximately 30 6-10 times the RDA of vitamin A; beta carotene is a prodrug of vitamin A);

between 68 mg and 100 mg zinc (approximately 4-7 times the RDA for zinc); and

between 1.6 mg and 2.4 mg copper.

In a further preferred embodiment, the amounts of each of these other components (on a per day basis) is as follows:

500 mg Vitamin C;

400 IU Vitamin E;

0 mg or 15 mg beta carotene;

25 mg or 80 mg zinc oxide; and

2 mg cupric oxide.

In a further preferred embodiment, that may be combined with any other embodiments herein, other ingredients believed to be of benefit in maintaining eye health may 45 likewise be combined with L-DOPA and/or L-DOPA analogues, including but not limited to lutein and/or zeaxanthin in an amount suitable to provide further protective retinal effects, preferably between 1 mg and 100 mg; between 1 mg and 50 mg, between 2 mg and 25 mg, or between 2 mg and 50 10 mg per day; and/or docosahexaenoic acid (DHA) and/or eicosapentaenoic acid (EPA) in an amount suitable to provide further protective retinal effects, preferably between 250 mg and 1000 mg; between 300 mg and 750 mg, between 350 mg and 750 mg, or between 350 mg and 650 mg per day. 55 mgFurther examples of additional compounds that may optionally be used include but are not limited to alpha-lipoic acid and, phenolic compounds such as for example but not limited to oligomeric proanthocyanidins, anthocyanosides and combinations thereof.

L-DOPA and/or L-DOPA analogues can be administered individually or in combination, usually in the form of a pharmaceutical composition. Such compositions are prepared in a manner well known in the pharmaceutical art. L-DOPA and/or L-DOPA analogues can be administered as 65 the sole active pharmaceutical agent, or they can be used in combination with one or more other compounds useful for

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carrying out the methods of the invention, including but not limited to an anti-angiogenic therapeutics such as VEG-F, and L-amino acid decarboxylase inhibitors, such as carbidopa and benserazide. When administered as a combination, combination can be formulated as separate compositions that are given at the same time or different times, or can be given as a single composition.

The L-DOPA and/or L-DOPA analogues may be made up in a solid form (including granules, powders or suppositories) or in a liquid form (e.g., solutions, suspensions, or emulsions). The L-DOPA and/or L-DOPA analogues may be applied in a variety of solutions and may be subjected to conventional pharmaceutical operations such as sterilization and/or may contain conventional adjuvants, such as preservatives, stabilizers, wetting agents, emulsifiers, buffers etc.

The L-DOPA and/or L-DOPA analogues may be administered by any suitable route, including but not limited to oral, topical (including but not limited to eye drops and ophthalmic ointments), parenteral, intranasal, pulmonary, or rectal in dosage unit formulations containing conventional non-toxic pharmaceutically acceptable carriers, adjuvants and vehicles. The term parenteral as used herein includes percutaneous, subcutaneous, intravascular (e.g., intravenous), intramuscular, or intrathecal injection or infusion 25 techniques and the like. In addition, there is provided a pharmaceutical formulation comprising a compound of the invention and a pharmaceutically acceptable carrier. L-DOPA and/or L-DOPA analogues may be present in association with one or more non-toxic pharmaceutically acceptable carriers and/or diluents and/or adjuvants, and if desired other active ingredients. The pharmaceutical compositions containing L-DOPA and/or L-DOPA analogues may be in a form suitable for oral use, for example, as tablets, troches, lozenges, aqueous or oily suspensions, 35 dispersible powders or granules, emulsion, hard or soft capsules, or syrups or elixirs.

Eye drops can be prepared using any technique in the art, including but not limited to using a tonicity agent such as sodium chloride or concentrated glycerin, a buffer such as sodium phosphate or sodium acetate, a surfactant such as polyoxyethylene sorbitan monooleate, polyoxyl 40 stearate or polyoxyethylene hydrogenated castor oil, a stabilizer such as sodium citrate or sodium edetate, a preservative such as benzalkonium chloride or paraben as needed. The pH of the eye drops is preferably in the range of from 4 to 8. Ophthalmic ointments can be prepared with a generally used base such as white soft paraffin or liquid paraffin.

L-DOPA and/or L-DOPA analogues intended for oral use may be prepared according to any method known to the art for the manufacture of pharmaceutical compositions and such compositions may contain one or more agents selected from the group consisting of sweetening agents, flavoring agents, coloring agents and preservative agents in order to provide palatable preparations. Tablets contain the L-DOPA and/or L-DOPA analogues in admixture with non-toxic pharmaceutically acceptable excipients that are suitable for the manufacture of tablets. These excipients may be for example, inert diluents, such as calcium carbonate, sodium carbonate, lactose, calcium phosphate or sodium phosphate; granulating and disintegrating agents, for example, corn starch, or alginic acid; binding agents, for example starch, gelatin or acacia, and lubricating agents, for example magnesium stearate, stearic acid or talc. The tablets may be uncoated or they may be coated by known techniques. In some cases such coatings may be prepared by known techniques to delay disintegration and absorption in the gastrointestinal tract and thereby provide a sustained action

over a longer period. For example, a time delay material such as glyceryl monosterate or glyceryl distearate may be employed.

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Formulations for oral use may also be presented as hard gelatin capsules wherein the L-DOPA and/or L-DOPA analogue is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin, or as soft gelatin capsules wherein the active ingredient is mixed with water or an oil medium, for example peanut oil, liquid paraffin or olive oil.

Aqueous suspensions contain the L-DOPA and/or L-DOPA analogues in admixture with excipients suitable for the manufacture of aqueous suspensions. Such excipients are suspending agents, for example sodium carboxymethylcellulose, methylcellulose, hydropropyl-methylcellulose, 15 sodium alginate, polyvinylpyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents may be a naturallyoccurring phosphatide, for example, lecithin, or condensation products of an alkylene oxide with fatty acids, for example polyoxyethylene stearate, or condensation products 20 of ethylene oxide with long chain aliphatic alcohols, for example heptadecaethyleneoxycetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide 25 with partial esters derived from fatty acids and hexitol anhydrides, for example polyethylene sorbitan monooleate. The aqueous suspensions may also contain one or more preservatives, for example ethyl, or n-propyl p-hydroxybenzoate, one or more coloring agents, one or more flavoring 30 agents, and one or more sweetening agents, such as sucrose or saccharin.

Oily suspensions may be formulated by suspending the L-DOPA and/or L-DOPA analogues in a vegetable oil, for example arachis oil, olive oil, sesame oil or coconut oil, or 35 in a mineral oil such as liquid paraffin. The oily suspensions may contain a thickening agent, for example beeswax, hard paraffin or cetyl alcohol. Sweetening agents and flavoring agents may be added to provide palatable oral preparations. These compositions may be preserved by the addition of an 40 anti-oxidant such as ascorbic acid.

Dispersible powders and granules suitable for preparation of an aqueous suspension by the addition of water provide the active ingredient in admixture with a dispersing or wetting agent, suspending agent and one or more preservatives. Suitable dispersing or wetting agents or suspending agents are exemplified by those already mentioned above. Additional excipients, for example sweetening, flavoring and coloring agents, may also be present.

Pharmaceutical compositions for use in the methods of 50 the invention may also be in the form of oil-in-water emulsions. The oily phase may be a vegetable oil or a mineral oil or mixtures of these. Suitable emulsifying agents may be naturally-occurring gums, for example gum acacia or gum tragacanth, naturally-occurring phosphatides, for example soy bean, lecithin, and esters or partial esters derived from fatty acids and hexitol, anhydrides, for example sorbitan monooleate, and condensation products of the said partial esters with ethylene oxide, for example polyoxyethylene sorbitan monooleate. The emulsions may 60 also contain sweetening and flavoring agents.

Syrups and elixirs may be formulated with sweetening agents, for example glycerol, propylene glycol, sorbitol, glucose or sucrose. Such formulations may also contain a demulcent, a preservative and flavoring and coloring agents. 65 The pharmaceutical compositions may be in the form of a sterile injectable aqueous or oleaginous suspension. This

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suspension may be formulated according to the known art using those suitable dispersing or wetting agents and suspending agents that have been mentioned above. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parentally acceptable diluent or solvent, for example as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including synthetic mono- or diglycerides. In addition, fatty acids such as oleic acid find use in the preparation of injectables.

Specific methods for intranasal administration of L-DOPA and L-DOPA analogues are known in the art; see, for example, Kao et al., Pharmaceutical Research 17(8):978-984 (2000).

The dosage range depends on the choice of the compound, the route of administration, the nature of the formulation, the nature of the subject's condition, and the judgment of the attending practitioner. For example, oral administration would be expected to require higher dosages than administration by intravenous injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization, as is well understood in the art. In certain embodiments, L-DOPA and/or L-DOPAS analogues can be administered at dosages of between 10 mg/day and 1500 mg/day; in various preferred embodiments administration can be between 20 mg and 1200 mg/day, 50 mg and 1000 mg/day, 100 mg and 500 mg/day, and 200 mg and 400 mg/day.

Pharmaceutical compositions containing the compounds described herein are administered to an individual in need thereof. In a preferred embodiment, the subject is a mammal; in a more preferred embodiment, the subject is a human. In therapeutic applications, compositions are administered in an amount sufficient to carry out the methods of the invention. Amounts effective for these uses depend on factors including, but not limited to, the nature of the compound (specific activity, etc.), the route of administration, the stage and severity of the disorder, the weight and general state of health of the subject, and the judgment of the prescribing physician. The active compounds are effective over a wide dosage range. However, it will be understood that the amount of the compound actually administered will be determined by a physician, in the light of the above relevant circumstances. Therefore, the above dosage ranges are not intended to limit the scope of the invention in any way.

In a third aspect, the present invention provides compositions comprising:

- (a) an amount effective of L-DOPA or an L-DOPA analogue for treating or limiting development of AMD; and
- (b) an amount effective for treating or limiting development of AMD of a composition comprising a source of vitamin C, a source of vitamin E, a source of vitamin A, a source of zinc, and a source of copper.

The amount of L-DOPA and/or L-DOPAS analogues in the compositions is suitable to provide for administration at dosages of between 10 mg/day and 1500 mg/day; in various preferred embodiments administration can be between 20 mg and 1200 mg/day, 50 mg and 1000 mg/day, 100 mg and 500 mg/day, and 200 mg and 400 mg/day.

Ascorbic acid is the preferred source of vitamin C in the subject tablets, although other sources such as for example sodium ascorbate could alternatively be used. Dl-alpha

tocopheryl acetate is the preferred source of vitamin E in the subject tablets although other sources of vitamin E, such as for example trimethyl tocopheryl acetate and/or vitamin E succinate, may be used in the alternative. Beta-carotene is preferred in the subject composition due to its ready com- 5 mercial availability although alternative carotenoid proforms of vitamin A could likewise be used. Zinc is preferred in the form of zinc oxide in subject tablets due to the fact zinc oxide provides the most concentrated form for elemental zinc and is well tolerated in the digestive system. However, other forms of zinc such as for example zinc gluconate may alternatively be used or be used in combination with zinc oxide in the subject composition. Copper in the form of cupric oxide is preferred in the subject tablets to although other forms of copper such as for example copper gluconate may alternatively be used or used in combination with cupric oxide in the subject composition.

In one preferred embodiment of this third aspect of the invention, composition "b" provides a formulation suitable 20 to permit ingestion of the following amounts of each component:

Ascorbic acid: at least 450 mg; dI-alpha tocopheryl acetate: 400 IU; beta carotene: 17.2 mg; zinc oxide: 68 mg; and cupric oxide: 1.6 mg.

In one preferred embodiment of this third aspect of the invention, composition "b" provides a formulation suitable to permit ingestion of the following amounts of each com- 30 ponent:

500 mg Vitamin C; 400 IU Vitamin E; 0 mg or 15 mg beta carotene; 25 mg or 80 mg zinc oxide; and 2 mg cupric oxide.

The preferred daily dosage of the subject composition as specified above may be administered in the form of 1, 2, 3, 4, or more dosage forms according to any suitable route of administration as disclosed above. In preferred embodi- 40 ments, the dosage form is an oral or topical dosage form, according to any embodiment of such dosage forms described herein. In another preferred embodiment the daily dosage of the subject composition is provided in the form of one dosage form taken twice daily, for a total of two dosage 45 forms a day, or in the form of two dosage forms taken twice daily, for a total of four dosage forms a day. Compared to taking the total daily dose once a day, twice daily dosing of half the total daily dose in one or more dosage forms per dose provides improved absorption and better maintenance 50 of blood levels of the essential ingredients. Accordingly, if two dosage forms of the preferred formulation of the subject composition are to be ingested each day, each dosage form is formulated to preferably provide not less than approximately 225 mg ascorbic acid, approximately 200 IU dl-alpha 55 tocopheryl acetate, approximately 8.6 mg beta-carotene, approximately 34 mg zinc oxide and approximately 0.8 mg cupric oxide upon oral administration. If four tablets of the preferred formulation of the subject composition are to be ingested each day, each tablet is formulated to preferably 60 provide not less than approximately 112.5 mg ascorbic acid, approximately 100 IU dl-alpha tocopheryl acetate, approximately 4.3 mg beta-carotene, approximately 17 mg zinc oxide, approximately 0.4 mg cupric oxide, and between 5 mg and 750 mg or L-DOPA and/or L-DOPA analogues.

In another preferred embodiment, the compositions comprise

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- (a) between 5 mg and 1500 mg L-DOPA or L-DOPA
- (b) between 450 mg and 600 mg vitamin C (approximately 7-10 times the recommended daily allowance (RDA))
- (c) between 400 IU and 540 IU vitamin E (approximately 13-18 times the RDA);
- (d) between 17.2 mg and 28 mg beta carotene (approximately 6-10 times the RDA of vitamin A; beta carotene is a prodrug of vitamin A):
- (e) between 68 mg and 100 mg of zinc (approximately 4-7 times the RDA for zinc); and
 - (f) at least 1.6 mg of copper.
- In various preferred embodiments, the composition may help prevent zinc induced copper deficiency anemia, 15 comprise between 10 mg and 1200 mg; between 25 mg and 1000 mg; between 50 mg and 500 mg, or between 100 mg and 400 mg L-DOPA or L-DOPA analogue.

In a further preferred embodiment, that may be combined with any other embodiments herein, other ingredients believed to be of benefit in maintaining eve health may likewise be combined with L-DOPA and/or L-DOPA analogues, including but not limited to lutein and/or zeaxanthin in an amount suitable to provide further protective retinal effects, preferably between 1 mg and 100 mg; between 1 mg 25 and 50 mg, between 2 mg and 25 mg, or between 2 mg and 10 mg per day; and/or docosahexaenoic acid (DHA) and/or eicosapentaenoic acid (EPA) in an amount suitable to provide further protective retinal effects, preferably between 250 mg and 1000 mg; between 300 mg and 750 mg, between 350 mg and 750 mg, or between 350 mg and 650 mg per day. The amounts necessary in any particular dosage form to provide the recited amounts can be determined by one of skill in the art based on the teachings herein and the number of dosage forms to be administered per day.

In a fourth aspect, the present invention provides in vitro methods for identifying compounds to treat AMD, comprising contacting cells with a test compound, wherein the cells

- (a) a first cell population expressing OA1; and, optionally,
- (b) a second cell population not expressing OA1; and
- (c) identifying as positive test compounds those test compounds that increase one or both of
 - (i) pigment epithelium-derived factor (PEDF) expression in the first cell population relative to one or both (A) PEDF expression in the first population of cells not contacted with the test compound, and (B) the second cell population, and
 - (ii) intracellular calcium concentration in the first cell population relative to one or both (A) intracellular calcium concentration in the first population of cells not contacted with the test compound, and (B) the second cell population

wherein the positive test compounds are candidate compounds for treating and/or limiting development of

As described above, human OA1 (SEQ ID NO:1-2 NP 000264.1) is a G-protein coupled receptor and the inventors have herein identified L-DOPA as an OA1 ligand. As disclosed in more detail below, the inventor has discovered the existence of an autocrine loop between OA1 and tyrosinase linked through L-DOPA, and this loop includes the secretion of at least one very potent retinal neurotrophic factor (PEDF) as well as an increase in intracellular calcium concentration. OA1 is a selective L-DOPA receptor whose downstream effects govern spatial patterning of the developing retina. Thus, test compounds that selectively upregulate PEDF expression and/or intracellular calcium con-

centration via stimulation of the OA1 pathway are candidate compounds for treating and/or limiting development of AMD. The methods of this aspect of the invention can be carried out with any OA1 homologue of, including but not limited to:

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Mouse: SEQ ID NO:3-4 (NM_010951); Xenopus tropicalis: SEQ ID NOS:5-6 (NM_001011018); Cow: SEQ ID NOS:7-8 (XM_001506318); Rat: SEQ ID NOS: 9-10 (NM_001106958); Platypus: SEQ ID NOS: 11-12 (XM_001506318); Xenopus laevis: SEQ ID NOS: 13-14 (NM 001096842) Chicken: SEQ ID NOS:15-16 (XM_416848); Zebrafish: SEQ ID NOS: 17-18 (NM_200822); Chimpanzee: SEQ ID NO: 19 (XR_025625); Rhesus monkey: SEQ ID NOS:21-22 (XM_001090139; 15

Macaque: SEQ ID NO: 23 (BV209253).

PEDF is pigment epithelium-derived factor (Exp Eye Res 53: 411-414), and is a known neurotrophic factor with the potential to alter neurosensory retina development, and to 20 inhibit blood vessel growth. The methods of this aspect of the invention can be carried out with any PEDF homologue of, including but not limited to:

Human: SEQ ID NOS:25-26 (NM_002615); Rat: SEQ ID NOS:27-28 (NM 031356); Zebra finch: SEQ ID NOS: 29-30 (XM_002197419); Horse: SEQ ID NOS:31-32 (NM_001143954); Xenpous tropicalis: SEQ ID NOS:33-34 (NM_203755); Mouse: SEQ ID NOS:35-36 (NM 011340); Atlantic salmon: SEQ ID NOS:37-38 (NM_001140334); 30 Sheep: SEQ ID NOS:39-40 (NM 001139447); Guinea pig: SEQ ID NOS:41-42 (EF679792); Cow: SEQ ID NOS:43-44 (NM_174140); Wild boar: SEQ ID NOS:45-46 (NM 001078662); Platypus: SEQ ID NOS:47-48 (XM_001507128); Wolf: SEQ ID NOS: 49-50 (NM_001077588); Macaque: SEQ ID NOS: 51-52 (AB174277); Chimpanzee: SEQ ID NOS: 53-54 (XM 001154665); Rhesus monkey: SEQ ID NOS: 55-56 (XM_001117361);

Flounder: SEQ ID NOS: 57-58 (DQ115406).

The first and second population of cells can be any suitable eukaryotic cell types, where the first population of cells is capable of expressing OA1 as a cell surface receptor protein. In one preferred embodiment, the first and second 45 populations of cells are of mammalian origin, such as mouse, rat, hamster, or human cells. All eukarvotic cells tested to date have been found suitable for carrying out the methods of the invention, particularly when used with embodiments involving analysis of intracellular calcium 50 concentration. Cell types tested to date for conservation of the OA1 signaling pathway disclosed herein with respect to one or both of intracellular calcium signaling and/or PEDF secretion include MCF7 (breast cancer epithelial cells), COS cells (kidney fibroblasts), MDCK cells (kidney epithelial), 55 CHO (Chinese hamster ovary), Mouse RPE, and 3T3 (mouse fibroblast), as well as those disclosed in the examples below. Such cells are commercially available from a variety of sources (LifeLine Cell Technology, Walkersville, Md.; ATCC (American Type Culture Collection)), or 60 can be isolated using methods known in the art and described below.

In one embodiment, a first portion of the first population of cells expressing OA1 as a cell surface receptor protein are contacted with the test compound, and a second portion of 65 the first population of cells are not contacted with the test compound, and those compounds that increase expression of

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PEDF and/or increased intracellular calcium concentration in the first portion relative to the second are candidate compounds for treating and/or limiting development of AMD.

Alternatively, the method may comprise use of a second population of cells not expressing OA1 as a cell surface receptor protein, and those compounds that increase expression of PEDF and/or increased intracellular calcium concentration in the first cell population relative to the second cell population are candidate compounds for treating and/or limiting development of AMD. In a preferred embodiment, the first and second populations of cells are the same cell type, with the first being engineered to recombinantly express OA1, while the second population of cells is not. In this embodiment, the second population of cells may be transfected with a similar expression vector as the first population of cells; such transfection may comprise transfection with an empty expression vector (ie: no expressed protein driven from the vector in the transfected cells), or an expression vector capable of expressing a truncated or mutated OA1 that does not insert appropriately into the cell membrane. Alternatively, cells can be transfected with an expression vector encoding an OA1 mutant known to be inactive for OA1 signaling, or an engineered form of OA1 that can signal through a different GPCR pathway (eg:

For example, one could fuse the 7 transmembrane domains of OA1 with a different intracellular c-terminal tail to change its activity without changing the ligand binding.

As used herein, an "increase in PEDF expression" or "increase in intracellular calcium concentration" is any increases in PEDF expression or intracellular calcium concentration in the first population of cells during the course of the assay above that seen in the second population of cells (or the first portion of the first population relative to the second portion). The method does not require a specific amount of increase in PEDF expression or intracellular calcium concentration over control, so long as the compound(s) promotes an increase in PEDF expression or intracellular calcium concentration above that seen in the control. In a preferred embodiment, the increase is a statistically significant increase as measured by standard statistical measurements.

Determining intracellular calcium concentrations is well known in the art and exemplary methods using Fura-2 cell loading and ratiometric imaging are described in the examples below. However, intracellular calcium concentration can be measured using any method known to those of skill in the art, including but not limited to FuraTM I (see below), or high throughput methods using FLIPerTM.

Determining expression levels of PEDF in the cell populations can be performed using any technique in the art such as those described below, including but not limited to, mRNA hybridization (Northern blot, slot blot, etc.), reverse transcription-polymerase chain reaction techniques using any suitable primer sets, fluorescence-in situ hybridization, and antibody detection in conditioned cell medium expressing/secreting PEDF (Western blot, immunocytochemistry, ELISA). PEDF antibodies are commercially available (for example, from Abcam, Cambridge, Mass.). Protein analysis can be on conditioned cell medium (since PEDF is an expressed protein); all assays can also be conducted at intracellular PEDF protein/mRNA production. In another embodiment, recombinant cells can be generated that include an expression vector driving expression of a detectable signal (GFP, luciferase, etc.) from the PEDF promoter; such cells can be used as the first cell population where

"PEDF expression" is measured via measuring the detectable fluorescent intensity or other signal driven by the PEDF promoter

As used in this fourth aspect, the term "contacting" means in vitro under suitable conditions to promote binding of OA1 5 ligands to OA1 expressed on the cell surface of the first population of cells. As used herein the "contacting" can occur at the time of initiating the culturing, or any time subsequent to initiating the culturing of the cell populations. PEDF expression and/or intracellular calcium concentration 10 can be measured at any time after contacting with the test compound as determined appropriate for a given assay. In one embodiment, a time course is carried out, measuring levels pre-contacting and at various times post-contact. In various embodiments, such measurements of calcium sig- 15 naling after contacting are made between 5 seconds and 60 minutes; more preferably 10 second and 30 minutes, 10 seconds and 10 minutes, and 10 seconds and 5 minutes. 10 seconds and 1 minutes, and 10 seconds and 30 seconds. In various embodiments, measurement of PEDF expression 20 can range between 1 minute and 72 hours, with analysis of PEDF secretion requiring later measurements than analysis of PEDF mRNA expression, PEDF intracellular protein expression, or expression of detectable signals driven by the PEDF promoter.

Any suitable cell culture conditions can be used as appropriate for a given assay. In one preferred embodiment, the contacting occurs in cell culture medium that has either a very low concentration of tyrosine (for example, between 0.1 um and 10 um tyrosine) or no tyrosine, to reduce its 30 production of endogenous L-DOPA in the cells, and to maintain the amount of OA1 present at the cell surface (since OA1 internalizes to the endosomes upon ligand binding). In one preferred embodiment, cells are cultured prior to test compound contacting in low tyrosine medium to 35 maximize OA1 expression and localization at the cell surface, followed by plating into tyrosine-free media for contacting with the test compounds. In another preferred embodiment, contacting occurs in low tyrosine medium. In another preferred embodiment, which can be combined with 40 other embodiments disclosed above, the culture media includes a tyrosinase inhibitor, including but not limited to phenylthiourea, to limit cell production of L-DOPA from tyrosine. This embodiment is particularly preferred when using pigmented cells.

In another preferred embodiment, the method may further comprise use of one or more of L-DOPA, tyrosine, and dopamine as competitors for binding to OA1. This embodiment may be carried out after identifying a test compound as an OA1 ligand, or it may be carried out in an initial screen 50 of test compounds for binding to OA1. As shown in the examples below, at concentrations of 1 mM and above, tyrosine and dopamine can compete with L-DOPA for binding to OA1. Thus, competitive assays using tyrosine and/or dopamine at concentrations between 1 mM and 100 55 mM, preferably between 1 mM and 50 mM or between 1 mM and 25 mM, can be used to further verify that the test compounds are operating via the OA1 pathway, and to measure the ability of tyrosine and dopamine to displace positive test compound binding to OA1 as compared to 60 displacement of L-DOPA. Similarly, competitive binding compared to L-DOPA (at similar molarity to the test compounds being tested) can help identify those compounds with increased avidity for OA1 compared to L-DOPA.

Any suitable test compounds can be assessed using the 65 methods of the fourth and fifth aspects (see below) of the invention, including small molecules, polypeptides, and

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nucleic acids. When the test compounds comprise polypeptide sequences, such polypeptides may be chemically synthesized or recombinantly expressed. Recombinant expression can be accomplished using standard methods in the art, as disclosed above. Such expression vectors can comprise bacterial or viral expression vectors, and such host cells can be prokaryotic or eukaryotic. Synthetic polypeptides, prepared using the well-known techniques of solid phase, liquid phase, or peptide condensation techniques, or any combination thereof, can include natural and unnatural amino acids. Amino acids used for peptide synthesis may be standard Boc (Nα-amino protected Nα-t-butyloxycarbonyl) amino acid resin with standard deprotecting, neutralization, coupling and wash protocols, or standard base-labile Nα-amino protected 9-fluorenylmethoxycarbonyl (Fmoc) amino acids. Both Fmoc and Boc Nα-amino protected amino acids can be obtained from Sigma, Cambridge Research Biochemical, or other chemical companies familiar to those skilled in the art. In addition, the polypeptides can be synthesized with other No-protecting groups that are familiar to those skilled in this art. Solid phase peptide synthesis may be accomplished by techniques familiar to those in the art and provided, such as by using automated synthesizers.

When the test compounds comprise antibodies, such antibodies can be polyclonal or monoclonal. The antibodies can be humanized, fully human, or murine forms of the antibodies. Such antibodies can be made by well-known methods, such as described in Harlow and Lane, Antibodies; A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., (1988).

When the test compounds comprise nucleic acid sequences, such nucleic acids may be chemically synthesized or recombinantly expressed as well. Recombinant expression techniques are well known to those in the art (See, for example, Sambrook, et al., 1989, supra). The nucleic acids may be DNA or RNA, and may be single stranded or double. Similarly, such nucleic acids can be chemically or enzymatically synthesized by manual or automated reactions, using standard techniques in the art. If synthesized chemically or by in vitro enzymatic synthesis, the nucleic acid may be purified prior to introduction into the cell. For example, the nucleic acids can be purified from a mixture by extraction with a solvent or resin, precipitation, electrophoresis, chromatography, or a combination thereof. Alternatively, the nucleic acids may be used with no or a minimum of purification to avoid losses due to sample processing.

When the test compounds comprise compounds other then polypeptides, antibodies, or nucleic acids, such compounds can be made by any of the variety of methods in the art for conducting organic chemical synthesis.

Test compounds identified as increasing the expression of PEDF and/or intracellular calcium concentration in the first cell population relative to the second cell population, can be further assessed for use as a candidate compound for treating or limiting development of AMD using any further technique, including but not limited to the in vivo methods of the fourth aspect of the invention, described below. In one preferred embodiment, the method may further comprise re-testing the positive test compounds in the assay in the presence of competitive amounts of tyrosine and/or dop-amine, as described above.

In a fifth aspect, the present invention provides methods for identifying compounds to treat AMD, comprising

(a) administering a test compound to a tyrosinase deficient pregnant female non-human mammal, wherein the test com-

pound is administered during embryonic photoreceptor and/ or retinal ganglion development; and

(b) comparing an effect of the test compound on photoreceptor and/or retinal ganglion development in the embryo or post-natal non-human mammal, to photoreceptor and/or 5 retinal ganglion development in an embryo or post-natal non-human mammal not administered the test compound, wherein those test compounds that increase photoreceptor and/or retinal ganglion development are candidate compounds for treating and/or limiting development of AMD. 10

The inventor has determined that OA1 signaling can be used to rescue photoreceptor and ganglion cell development in tyrosinase-deficient animals, and in the process establish the neurotrophic effect of OA1 signaling. Thus, compounds that rescue neurosensory retinal development through OA1 15 signaling are good candidates for AMD treatment. The present invention provides the first establishment of such an animal model for AMD drug screening.

As described in more detail herein, tyrosinase acts on tyrosine to create L-DOPA. Thus, a tyrosinase deficient 20 mammal does not produce L-DOPA, permitting the use of such mammals to identify activators of OA1 (via rescue of retinal development and/or increased PEDF expression) in the absence of endogenous L-DOPA. As used herein, a "tyrosinase deficient" means that the pregnant female non- 25 human mammal does not produce adequate amounts of tyrosinase to create L-DOPA in amounts adequate for normal pigment formation. In one preferred embodiment, the pregnant non-human mammal is a knockout animal (deleted for portion or all of the tyrosinase gene, or have naturally 30 occurring mutations in the tyrosinse gene or accessory genes that control, activate, or traffic tyrosinase to the melanosome) with no ability to express or traffic functional tyrosinase. Such tyrosinase knockouts are known in the art and are commercially available (Lexicon Pharmaceuticals, Jack- 35 son Laboratories, Taconic Farms. In other embodiments, the tyrosinase deficiency may be transiently induced by methods known in the art including, but not limited to, administering siRNAs targeting tyrosinase, tyrosinase antibody/aptamer treatment, etc.

The non-human mammal can be any in which tyrosinase-deficient (retinal albino) females can be obtained, which includes all mammals. In various preferred embodiments, the non-human mammal is mouse, pig, apes, and rat.

In one preferred embodiment, administration of test com- 45 pound is continued during the post-natal period of photoreceptor and/or retinal ganglion development. The embryonic and post-natal photoreceptor and/or retinal ganglion development pathways in various non-human mammals is well understood by those of skill in the art. In one exemplary 50 embodiment, mouse embryonic photoreceptor and retinal ganglion development begins on embryonic day 10 (E10) and retinal development is complete by postnatal day 14 (P14) when the pups eyes are open. Thus, in various embodiments, test compounds are first administered at about 55 day E7, E8, E9, or E10 (to facilitate its presence at the earliest stage of ocular development) and administration can continue as desired for a given assay between day P1, P2, P3, P4, P5, P6, P7, P8, P9, P10, P11, P12, P13, and day P14 or later as desired (up to one year post-natal). As will be 60 understood by those of skill in the art, administration will be to the pregnant female mother during the embryonic phase and to the pup postnatally. In another embodiment, pigmented cell development begins in earnest at approximately day E10.5 (when OA1 and tyrosinase appear), and thus in 65 one embodiment, administration of test compound may begin on about day E10, E10.5, or E11 and continue as

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desired up to about day P1, P2, P3, P4, P5, P6, P7, P8, P9, P10, P11, P12, P13, P14 or later as desired. In another embodiment, test compound administration may be limited to between day E7 and E10 or E11. In a further embodiment, retinal ganglion development begins in earnest at about day E12, and thus in one embodiment, administration of test compound may begin on about day E12 or E13 and continue as desired up to about day P1, P2, P3, P4, P5, P6, P7, P8, P9, P10, P11, P12, P13, P14 or later as desired. In another embodiment, test compound administration may be limited to between day E7 and E12 or E13. In a most preferred embodiment test compounds are first administered daily from day E7 until day P14. As will be understood by those of skill in the art, the exact timing of test compound administration will depend on the goals of the particular assay and can be determined by one of skill in the art based on the teachings herein.

The test compounds may be administered by any route suitable for use with experimental animals, including those routes of administration disclosed above for therapeutic administration of L-DOPA or L-DOPA analogues. In a preferred embodiment, the test compounds are administered in the animal's drinking water, parenterally (as discussed above) or topically (for example, in eye drops or ophthalmic ointments). Frequency of test compound administration can be as often as appropriate for a given assay; in a preferred embodiment, test compound is administered daily throughout the desired course of treatment; in other embodiments, administration is every second, third, fourth, or fifth day during the course of treatment; the frequency of administration can be determined by one of skill in the art based on the teachings herein and the specific goals of a given assay.

As used herein, an "increase in photoreceptor and/or retinal ganglion development" is any increase in photoreceptor and/or retinal ganglion development in test-compound treated vs. non-treated embryos/animals. The method does not require a specific amount of increase in photoreceptor and/or retinal ganglion development over control, so long as the compound(s) promotes an increase in photoreceptor and/or retinal ganglion development above that seen in the control. In a preferred embodiment, the increase is a statistically significant increase as measured by standard statistical measurements. In one embodiment, animals are euthanized at the appropriate time point, and retinal ganglion cells and/or photoreceptors are counted using standard methods in the art, including but not limited to those disclosed in the examples below.

Test compounds identified as increasing photoreceptor and/or retinal ganglion development, can be further assessed for use as a candidate compound for treating or limiting development of AMD using any further technique, including but not limited to re-testing the positive test compounds using the in vitro methods disclosed in the third aspect of the invention in the presence of competitive amounts of tyrosine and/or dopamine. As shown in the examples below, at concentrations of 1 mM and above, tyrosine and dopamine can compete with L-DOPA for binding to OA1. Thus, competitive assays using tyrosine and/or dopamine at concentrations between 1 mM and 100 mM, preferably between 1 mM and 50 mM or between 1 mM and 25 mM, can be used to further verify that the test compounds are operating via the OA1 pathway, and to measure the ability of tyrosine and dopamine to displace positive test compound binding to OA1 as compared to displacement of L-DOPA.

EXAMPLES

L-DOPA is an Endogenous Ligand for OA1

Background: Albinism is a genetic defect characterized by 5 a loss of pigmentation. The neurosensory retina, which is not pigmented, exhibits pathologic changes secondary to the loss of pigmentation in the retina pigment epithelium (RPE). How the loss of pigmentation in the RPE causes developmental defects in the adjacent neurosensory retina has not 10 been determined, but offers a unique opportunity to investigate the interactions between these two important tissues. One of the genes which causes albinism encodes for an orphan GPCR (OA1) expressed only in pigmented cells, including the RPE.

Methodology/Principle Findings: The function and signaling of OA1 was investigated in RPE and transfected cell lines. The results indicate that OA1 is a selective L-DOPA receptor, with no measurable second messenger activity from two closely related compounds, tyrosine and dopamine 20 Radiolabeled ligand binding confirmed that OA1 exhibited a single, saturable binding site for L-DOPA. Dopamine competed with L-DOPA for the single OA1 binding site suggesting it could function as an OA1 antagonist. OA1 response to L-DOPA was defined by several common mea- 25 sures of GPCR activation including influx of intracellular calcium and recruitment of β-arrestin. Further, inhibition of tyrosinase, the enzyme that makes L-DOPA, resulted in decreased PEDF secretion by RPE. Further, stimulation of OA1 in RPE with L-DOPA resulted in increased PEDF 30 secretion.

Conclusions/Significance: Taken together the results illustrate an autocrine loop between OA1 and tyrosinase linked through L-DOPA, and this loop includes the secretion of at least one very potent retinal neurotrophic factor. OA1 35 is a selective L-DOPA receptor whose downstream effects govern spatial patterning of the developing retina. The results suggest that the retinal consequences of albinism caused by changes in melanin synthetic machinery may be treated by L-DOPA supplementation.

Introduction: Albinism is a group of inherited genetic diseases in which there is a variable loss of pigmentation in the eye, hair or skin. When the eye is affected, there are significant alterations in neurosensory retina development that lead to low vision [1-8]. There are two broad classes of 45 albinism, ocular-cutaneous albinism (OCA) and ocular albinism (OA). OCA occurs when all pigmented tissues exhibit hypopigmentation and involves genetic mutations that result in defects in the melanin synthetic machinery [3,7-9]. OA occurs when cutaneous tissues pigment normally, but the 50 ocular tissues are hypopigmented [10,11]. Since the same proteins produce pigment in all tissues, OA most likely results from lack of expression of the melanogenic enzymes in ocular tissue rather than an inability to synthesize melanin because the other tissues pigment normally.

OA can be linked to at least one gene, Oa1, which is found on the X chromosome. Oa1 encodes a 404 amino acid protein likely to be an orphan G-protein coupled receptor (GPCR), OA1 (Genbank GPR143) [12,13] based upon sequence analysis [14]. Schiaffino et al. has demonstrated 60 that OA1 associates with several G_{α} , subunits as well as G_{β} adding further evidence that OA1 is a GPCR [14,15]. Indeed, Innamorati et. al. used a combinatorial expression strategy to illustrate GPCR-like activity from OA1, as well as β -arrestin association, even in the absence of a ligand 65 [16]. This work suggested that OA1 could signal through a G α q subunit through phospholipase C and inositol triphos-

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phate second messengers. In a yeast based expression system, Staleva and Orlow have demonstrated GPCR signaling from OA1 that appeared to be activated by a component in the melanosomal compartment [17]. Despite the significant amount of circumstantial evidence that OA1 is a GPCR, confirmation is lacking because no ligand has been identified. Other data has called into question the idea that OA1 is a GPCR. For example, the localization of OA1 as a fully intracellular protein is not typical of GPCRs and suggests that it would be a unique member of the family [14]. OA1 is primarily localized to the endolysomal compartment [14, 15,18-21] and melanosomes [11,14,22] rather than the cell surface

In this study the function of OA1 as a potential GPCR was 15 investigated, based on the hypothesis that the endosomal localization of OA1 in cultured cells was due to internalization of OA1 in response to an agent in the culture medium. Further, a ligand for OA1 was sought based on the observation that all forms OCA and OA appear to have the same retinal phenotype, indicating that tyrosinase activity and OA1 signaling are coupled upstream of retinal development. Thus, tests on whether tyrosinase activity produces the ligand for OA1 were carried out. A by-product of melanin synthesis is L-DOPA, which is released to the retina during melanin synthesis in the RPE at a critical time in retinal development [23,24]. The data suggest that OA1 is a highly selective L-DOPA receptor, and that L-DOPA causes OA1 signaling with the downstream effect of neurotrophic factor secretion by RPE. Thus, the first evidence is presented of a ligand for OA1, and provide a mechanism through which either tyrosinase or OA1 deficiency results in changes to retinal development.

Results:

Cell Surface Localization of OA1.

OA1 has previously been localized in pigment granules in situ [22], however, using transfected cells of various types, OA1 also has been localized to both the plasma membrane [16,17] and the endosomal fraction of cultured cells [14,16-18,20,21]. The investigation began by determining where 40 OA1 resides in the human tissue using cell surface biotinylation/western blot strategies. In the human eye, OA1 was present on the apical cell surface of the RPE in situ (FIG. 1 A). Quantification of cell surface, biotinylated OA1 in five human eyes indicated that at least 3.5+/-0.7% of the total OA1 resided on the apical cell surface of RPE in situ. Access to the biotinylation reagent using eye cup preparations is restricted to the apical surface, so the polarity of OA1 in the epithelium cannot be determined. Further, the total cell surface OA1 is likely underestimated because of the lack of access to the basal cell surface. Blots were also probed with antibodies against actin as a control to verify that cytoplasmic proteins were not biotinylated. In each experiment actin was only found in the unbound fraction.

Others have reported that recombinant OA1 and OA1-55 GFP is almost exclusively localized to the endosomal compartment in cultured cells [14,15,17,18,20-22]. However, when overexpressed [16], or when endocytosis is inhibited [17], OA1 accumulates at the cell surface. The observation that OA1 protein is present on the apical surface of RPE in 60 situ led us to explore the issue further.

Effects of Tyrosine on OA1 Expression and Distribution

Endosomal localization of GPCRs occurs normally after exposure to a ligand. Therefore, it was investigated whether a ligand for the receptor was present in the standard incubation medium that could drive internalization of OA1. Since the standard culture medium contains 500 µM tyrosine, and tyrosine is the starting material for pigment syn-

thesis, the effect of tyrosine on receptor distribution was evaluated. To test whether tyrosine affected OA1 distribution in cultured cells DMEM was formulated without tyrosine, and dialyzed fetal bovine serum was used. In the presence of tyrosine-free medium, OA1 was detected on the plasma membrane of cultured RPE cells both in the absence (not shown), and in medium containing low concentrations of tyrosine (1 µM, FIG. 1 B). Averaged over five experiments, 4.5+1-1% of total OA1 protein was observed on the surface of cultured RPE maintained in 1 μM tyrosine, similar to what was observed for RPE in situ. In all experiments actin was observed in the unbound protein fraction, demonstrating the absence of any cytoplasmic protein in the cell surface assay. Similarly, OA1-GFP expressed in COS illustrated a cell surface expression that was tyrosine sensitive 15 (FIG. 1 C). Quantification of six such experiments indicated significant variability in the amount of OA1 found at the cell surface using transient transfections. The range of OA1 in the bound fraction of transfected cells maintained in 1 μM tyrosine ranged between 5-40%, unlike the results with the 20 endogenous OA1 protein that were reproducibly ~5%.

Not only was the distribution of OA1 in transfected cells sensitive to tyrosine levels in the medium, total OA1-GFP expression was increased 5-fold in cells maintained in 1 μM evaluated from the paired samples. The data (FIG. 1 D) presented as optical density units indicate no difference in actin. The amount of cell surface OA1 between the normal and low tyrosine groups was also compared. Importantly, in 30 the five RPE experiments and six OA1-GFP in COS experiments, OA1 in the plasma membrane fraction of cells in standard medium was not reproducibly detected, similar to that found by others.

The distribution of OA1 in RPE cells also was evaluated 35 by confocal microscopy. OA1 has previously been characterized as an endosomal protein in cultured RPE cells as shown in (FIG. 1 E). In contrast, the distribution of OA1 in low tyrosine medium was diffuse on the plasma membrane of cultured RPE cells, with little endosomal accumulation 40 (FIG. 1 F), an observation consistent with the results obtained using biochemical methods.

L-DOPA as a Natural Agonist for OA1.

Tyrosinase function in melanogenesis begins with its activity on tyrosine to create L-DOPA, followed by a second 45 reaction to create dopaquinone that leads to pigment formation [25]. Of the intermediates between tyrosine and melanin, L-DOPA has the greatest half-life, and L-DOPA is released into the subretinal space apical to the RPE when melanin synthesis occurs [23,24]. L-DOPA is also the pre- 50 cursor to dopamine, a neurotransmitter produced by dopaneurgic neurons from tyrosine. The release of calcium from intracellular stores is a common downstream effect of GPCR activation by a ligand. Since the expression of OA1 on the cell surface appears to be sensitive to tyrosine, it was 55 examined whether tyrosine, or its metabolites L-DOPA and dopamine, could stimulate influx of Ca²⁺ into the cytoplasm in an OA1-dependent manner. CHO cells were transfected with an OA1 expression vector then maintained in DMEM containing 1 µM tyrosine for 48 hours followed by tyrosine- 60 free DMEM for 24 hours to facilitate cell surface expression of OA1. Intracellular Ca2+ was evaluated using Fura-2, and [Ca²⁺]i was determined by ratiometric imaging [26]. In the absence of any ligand, [Ca²⁺]i was not significantly different between transfected and untransfected cells (FIG. 2). Tyro- 65 sine and several tyrosine metabolites were tested at 1 µM for an effect on [Ca²⁺]i. As a positive control each experiment

was ended by treatment with 20 mM KCl to depolarize the cell and increase [Ca2+]i via activation of voltage-gated channels. This maneuver served to verify the Fura-2 loading and responsiveness of the cells being tested (FIG. 2). Only L-DOPA elicited a significant increase in [Ca²⁺]i (FIG. **2** A). Tyrosine and dopamine had no positive effect on intracellular at [Ca²⁺]i concentrations up to 1 mM (not shown). The slight negative effect of 1 µM dopamine was not statistically significant, but reproducible among the 11 experiments with dopamine (FIG. 2 B).

Over expression of GPCRs in non-native cell lines can lead to false signal transduction coupling. To verify that OA1 signaling in response to L-DOPA was indeed a natural response, OA1 was expressed in RPE cells (FIG. 2 C). Results using transfected RPE cells were similar to those achieved with transfected CHO cells. RPE cells transfected to express OA1 responded to 1.0 µM L-DOPA with an increase in [Ca²⁺]i. It was next determined whether RPE cells expressing the endogenous OA1 receptor, at endogenous levels exhibited L-DOPA responsiveness. Like all of the transfected cell experiments, RPE expressing OA1 demonstrated an increase in [Ca2+]i after treatment with 1.0 µM L-DOPA (FIG. 2 C).

To further characterize OA1 signaling activity, pertussis tyrosine. To verify that this difference related to OA1 25 toxin was used to distinguish between G_q coupled $[Ca^{2+}]i$ expression rather than cell number, actin expression was signaling and G_i linked signaling (FIG. 2 C). In all cells studied, pertussis toxin lowered the basal level of [Ca²⁺]i, indicating its activity on inhibition of the background signaling through G_i subunit activity. Pertussis toxin was used in experiments conducted in cells transfected to express OA1 including both CHO and RPE, as well as RPE expressing the endogenous OA1 protein at natural levels. In all transfected cells tested the measured [Ca2+]i response to L-DOPA was greater than in the absence of the toxin (FIG. 2), owing largely to the lower initial [Ca²⁺]i. Thus, the signaling through OA1 in response to L-DOPA that results in increase [Ca²⁺]i is not pertussis toxin sensitive and likely G_a subunit mediated. The second messenger cAMP was also measured in CHO cells transfected to express OA1 (FIG. 2 D). Using inactive cells or a submaximal forskolin treatment, the experiments were set up to measure either an increase or decrease in cAMP in response to L-DOPA. In six such experiments, no change in cAMP was observed suggesting neither G_s nor G_i subunits are involved in OA1 signaling.

Standard methods of radiolabeled ligand binding were used to characterize the interaction between OA1 and L-DOPA (FIG. 3 A). CHO cells were transfected to express OA1, then binding of L-DOPA was quantified in a concentration-dependent manner, and the results were further characterized by Scatchard Plot analysis (FIG. 3E). Results illustrate saturable binding of L-DOPA to OA1 expressing cells with a Kd of 9.35×10⁻⁶M. No specific binding was observed in untransfected CHO cells, indicating that the cells do not have an endogenous L-DOPA receptor (not shown). All binding parameters, total, specific, and nonspecific are shown as supplemental data (FIG. 6A). Tyrosine exhibited the potential to interact with OA1, but neither tyrosine nor dopamine stimulated OA1 signaling (see FIG. 2). Competitive ligand binding was used to determine whether either tyrosine or dopamine competed with L-DOPA for OA1 binding. At high concentrations (1 mM), both tyrosine and dopamine competed with L-DOPA for OA1 binding (FIG. 3 B). To further characterize this the kinetics of the competition between L-DOPA and either dopamine (FIG. 3 C) or tyrosine (FIG. 6B) was examined Dopamine exhibited competitive binding to a single site

with L-DOPA with a Ki of $2.33 \times 10^{-6} + 1 - 0.2 \times 10^{-6}$ M. Similar experiments with tyrosine demonstrated inhibition of L-DOPA binding only at high concentrations (FIG. 6B). Saturation kinetics were not possible with tyrosine because of its low affinity and insolubility at the high concentrations. 5

Given the relatively low affinity of OA1 for L-DOPA it was determined whether its signaling activity was dosedependent in the range of this binding affinity. The concentrations in which binding data suggested the steepest rise in association between L-DOPA and OA1, 1.0-10 µM were 10 tested, and results illustrate a concentration dependent GPCR response as measured by [Ca²⁺]i (FIG. 3 C). Thus, the activation kinetics of L-DOPA and OA1 matched the concentration range observed in radiolabeled ligand binding experiments.

In response to ligand binding, GPCRs recruit β-arrestin to the plasma membrane which is followed by internalization of the ligand-receptor complex [27-33]. The effect of L-DOPA on β -arrestin localization was then tested (FIG. 4). tyrosine DMEM for 48 hours prior to analysis to allow cell surface expression of the protein. Cells were then treated with 1 µM L-DOPA followed by rapid fixation on ice in cold methanol. Initially, under resting conditions in the absence of an agonist, OA1-GFP was found at the cell surface and 25 β -arrestin was diffuse in the cytoplasm (FIG. 4 A-C), with no co-localization between the proteins. After stimulation with L-DOPA, OA1 and β-arrestin were co-localized at the plasma membrane (FIG. 4 D-F). Untransfected cells showed no response to L-DOPA treatment (FIG. 4 G,H), illustrating 30 that the L-DOPA effect on β-arrestin distribution was OA1 dependent, similar to results obtained for [Ca²⁺]i. Effects of 1-DOPA on PEDF Secretion

Mutations in OA1 cause defects in the development of the neurosensory retina. In previous work it has been shown that 35 pigmented RPE secrete significantly more PEDF than nonpigmented RPE [34], and PEDF is a neurotrophic factor with the potential of altering neurosensory retina development [35-41]. Mutations in OA1 cause a loss of pigmentation in the RPE, suggesting that OA1 activity governs RPE pig- 40 mentation. Thus, it was determined whether L-DOPA stimulation of pigmented RPE cells caused increased secretion of PEDF (FIG. 5). This assay is made somewhat more difficult because pigmenting RPE cells produce L-DOPA, which is the agonist for OA1, and OA1 is not readily detectable in 45 nonpigmented cultures of RPE. Thus, pigmented RPE were used to determine whether L-DOPA stimulation increases PEDF expression/secretion. RPE cells were placed in tyrosine-free medium for 24 hours then treated with 1 μ M L-DOPA for one hour. After treatment, the cells were 50 returned to standard medium without exogenous L-DOPA for three days. Control cells were not treated with L-DOPA, but the medium was changed at the same time the experimental cells were returned to normal medium. Conditioned medium was collected after three days and PEDF was 55 measured. Results illustrate a significant increase in the secretion of PEDF in pigmented cells treated with L-DOPA when compared to paired, control monolayers of pigmented RPE (FIG. 5 A). Importantly, this significant increase occurred in cells which were pigmenting and therefore 60 expressed OA1 and had a basal level of PEDF expression.

To determine whether pigmented RPE cells secrete PEDF through an autocrine loop involving tyrosinase activity and OA1 signaling, a specific tyrosinase inhibitor phenylthiourea (PTU) was used to inhibit pigmentation and L-DOPA 65 production (FIG. 5 B). In these experiments, pigmented RPE cells were either maintained in DMEM, or DMEM contain-

ing 200 µM PTU for three days, then PEDF secretion was measured. Pigmented RPE secreted substantial PEDF, but PTU caused a significant decrease in PEDF secretion indicating that tyrosinase activity is necessary for the high level of PEDF secretion observed in pigmented RPE cells. To verify that it was the lack of L-DOPA in the PTU treated cells that caused the decreased PEDF secretion, 3 different cultures of pigmented RPE were used, and exposed to PTU for 48 hours, then treated with 1.0 µM L-DOPA in the continued presence of PTU; PEDF was measured after 72 hours (FIG. 5 C). The data are presented as percent of control for this experiment because the cultures used varied in both pigmentation and PEDF expression before the experiment began. PTU treated RPE responded to the added L-DOPA by increasing PEDF secretion, indicating that the effect of PTU on PEDF secretion is caused be the lack of L-DOPA production when tyrosinase is inhibited. Discussion:

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There is a complex inter-tissue relationship between the Cells were transfected to express OA1 then cultured in 1 uM 20 RPE and the neurosensory retina. One aspect of this relationship is centered on RPE pigmentation, and defects in melanin synthesis which result in significant neurosensory retina alterations [8,23,42]. The data suggest that OA1 and tyrosinase participate in an autocrine loop through L-DOPA that regulates the secretion of at least one potent neurotrophic factor, PEDF. The data also suggest that the pathologic changes in retinal development that occur in albinism may result from changes in the activity of the OA1 signaling pathway. Reduced OA1 signaling activity can be caused either directly through OA1 mutations or indirectly through changes in L-DOPA production by tyrosinase activity. Thus, it is hypothesized that the similar retinal phenotypes that accompany the diverse forms of albinism can be reconciled to a single common pathway, OA1 signaling.

> In the study, OA1 on the apical surface of human RPE in situ was observed. Previous reports have suggested that OA1 in mice is localized to the melanosome [22], and in cultured cells to the endosomal compartment [15-18,20-22, 43]. The results from in situ RPE preparations indicate that OA1 is distributed to the apical surface of the RPE. The limited quantities of OA1 on the surface of the RPE (~3.5% of total OA1) may account for the lack of observation of the protein in previous studies where immunogold electron microscopy was used. Like many cell surface GPCRs, OA1 is not an abundant protein.

> The endosomal localization of OA1 reported in previous studies using cultured cells was reproduced in this study for both the endogenous protein and the transgenic protein. When tested in normal culture medium little detectable OA1 protein on the cell surface was found, in agreement with all previous work. However, reduction of tyrosine in the medium caused a modest increase in cell surface receptor accumulation of both the endogenous and recombinant OA1 proteins. This suggests that the distribution of OA1 to the cell surface in cultured cells is sensitive to tyrosine. A previous study has demonstrated OA1 could be localized to the cell surface when endocytosis in inhibited [17] and OA1 on the apical surface of human RPE was observed in situ. The data suggest OA1 is a cell surface GPCR, but is a target for endocytosis that may be stimulated by tyrosine or tyrosine metabolites. In this regard, the results differ from past reports of OA1 localization that have classified OA1 as a unique type of intracellular GPCR. Most GPCRs are cell surface proteins that are internalized by a variety of signals, and the data suggest OA1 is similar to most other GPCRs.

> OA1 signaling activity was stimulated by L-DOPA, but not by either its precursor, tyrosine, or its neuronal metabo-

lite dopamine. This result suggests an exquisitely sensitive receptor activity able to distinguish between closely related molecules, after all L-DOPA and tyrosine differ by a sole hydroxyl group. OA1 is sensitive to tyrosine, as tyrosine causes an intracellular localization of OA1 in cultured cells. 5 However, no signaling response to tyrosine was noted, and competition binding studies suggest that tyrosine has a low affinity for OA1. The data suggest that the continuous exposure of cells to high concentrations of tyrosine present in normal medium is sufficient to result in internalization of 10 OA1, but it is unlikely to result in measurable OA1 activation. Strong evidence of a single site competitive interaction between L-DOPA and dopamine was found. The Ki observed for dopamine was similar to the Kd observed for L-DOPA, suggesting that the affinity for the two tyrosine 15 metabolites is similar. The results illustrated a slight, but reproducible, decrease in OA1 signaling from dopamine, suggesting that dopamine may be an effective antagonist or inverse agonist for OA1.

As an orphan GPCR, its signaling pathway has not 20 previously been identified. In this study it was illustrated that OA1 signaling in response to L-DOPA causes an increase in [Ca²⁺]i. The data illustrate that the increased [Ca²⁺]i observed in response to L-DOPA was insensitive to pertussis toxin and no effects on cAMP were found, indicating that 25 OA1 is likely signaling through a G_{α} subunit. Previous work has suggested that OA1 can associate with multiple subunits in transfected cells including members of the G_o, G_i, and G_a subunit families. Innamorati et al. has shown that spontaneous activity of overexpressed OA1 is likely signaled 30 Human RPE In Situ—Human eyecups were produced by through a Gq subunit [16]. The data indicate that liganddependent signaling from endogenous OA1 in RPE most likely occurs through a G_q mediated pathway, and no promiscuous coupling activities were observed when comparing OA1 over expression in CHO and RPE to natural OA1 expressed in RPE. Interestingly, two overactive mutant forms of Gq subunits cause hyperpigmentation in skin and hair [44], but whether they have an effect in RPE is unknown. RPE and cutaneous melanocytes use the same enzymes to produce pigmentation but differ in their control 40 of melanogenesis. A recent report suggests that OA1 may signal through Gai3, because the retinal phenotype of $OA1^{-/-}$ and $G\alpha i3^{-/-}$ are similar [45]. That study provided no data regarding interaction or signaling between Gai3 and OA1, and the results do not support OA1 signaling through 45 Gai3. However, both OA1 and Gai3 could have activity in convergent pathways that govern some part of the complex system of retinal development.

The response of OA1 to L-DOPA was measured in three ways, increased [Ca²⁺]i, recruitment of β-arrestin to plasma 50 membrane OA1, and the increased secretion of PEDF. In addition, inhibiting the activity of tyrosinase in pigmented RPE inhibits L-DOPA production, and results in a decreased secretion of PEDF. Taken together, these studies present a strong argument for a productive ligand:receptor relation- 55 ship between L-DOPA and OA1. Further, the data suggest selectivity among tyrosine and its metabolites, with only L-DOPA being a productive ligand for OA1. We have determined the binding kinetics between OA1 and L-DOPA, and observed a typical one site receptor:ligand relationship between the two. The binding affinity between OA1 and L-DOPA, with a Kd in the µM range, is not uncommon for an endogenous ligand:receptor relationship. Future identification of a specific, high affinity antagonist for OA1 will aid in further biochemical characterization of the interaction 65 between OA1 and L-DOPA, and be useful in determining whether dopamine is an inverse agonist.

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This study illustrated the selective activation of OA1, an orphan GPCR, by L-DOPA, an intermediate product of melanin synthesis. This study has also illustrated that OA1 activity stimulates PEDF secretion by RPE, a molecule that has the potential to support normal retinal development [40,41]. In humans, this suggests that pharmacologic intervention through OA1 activation could be useful for albinism caused by defects in the melanogenic machinery (OCA 1-4). Unfortunately, the data also suggest that OA1 is necessary for such pharmacologic intervention, and mutations in Oal are the most common cause of albinism.

Methods:

Cell Culture

RPE—Cells were isolated as described [46] and maintained in Dulbecco's modified essential medium (DMEM) supplemented with 5% fetal bovine serum (FBS). For experiments in which tyrosine concentrations were lowered, custom manufactured DMEM produced without tyrosine by JRH Biosciences (Lenexa, Kans.) was used. Dialyzed FBS was purchased from Invitrogen, (San Diego, Calif.).

COS-7 and CHO-Cells were obtained from ATCC and cultured in DMEM supplemented with 5% FBS. For analysis of OA1 distribution, cells were cultured in tyrosine-free DMEM supplemented with 1 μM tyrosine, 5% dialyzed FBS for 2-4 days, then tyrosine-free media as described for the experiment.

Cell Surface Biotinylation

dissection ~2 mm anterior to the equator and removals of the anterior segment. The vitreous and retina were removed without impairing the underlying RPE monolayer, and the retina was cut at the optic nerve head. The resulting eyecups with RPE exposed were rinsed three times with reaction buffer (100 mM NaCl, 50 mM NaHCO3, pH 8.0) then filled with Sulfo-NHS-LC-Biotin (1 mg/ml) two times for thirty minutes. The reaction was stopped with TG buffer (25 mM Tris, 192 mM Glycine, pH 8.3) then the cells were harvested in lysis buffer (2 mM EDTA, 1% Triton X and 1% Tween 20 in Tris Base Saline Buffer) containing Halt Protease Inhibitor Cocktail. Intact cells and pigment granules were removed by centrifugation at 14,000 rpm for 20 minutes. Biotinylated proteins were captured overnight with immobilized streptavidin beads and then mixed with 4× reducing buffer (250 mM Tris, pH 6.8, 8% SDS, 40% Glycerol, 20% Beta-mercaptoethanol, 0.08% bromophenol blue). The OA1 protein was separated on a 10% SDS-PAGE gel and identified by a using a polyclonal rabbit OA1 antibody for western blot analysis. Paired western blots were probed with a monoclonal antibody directed against actin.

Cultured Cells—RPE and transfected cells were maintained in DMEM containing tyrosine concentrations described for the experiment. Cultures were rinsed three times in reaction buffer, then biotinylated as described above for the in situ preparation.

Cloning of Oa1

A cDNA library was constructed from pooled tissue from 6 human donor eyes. Total RNA was harvested using Trizol reagent, then cDNA was synthesized using Poly-T primers for the first strand synthesis, and random hexamers for the second strand. Following cDNA synthesis, RNA was removed using RNase A. The coding sequence for OA1 was obtained by PCR using terminal primers that added restriction sites to the 5' and 3' ends and removed the native stop codon. The PCR product was ligated in frame with GFP in

the pEGFP N-1 vector (Clontech). The sequence was verified by automated sequencing in both directions over the entire sequence.

Immunocytochemistry

Cells on slides were fixed with 3% paraformaldehyde at 5 RT, rinsed with 0.1% Triton X-100 in 10% milk in TBST then blocked with 10% milk in TBST. β-arrestin was visualized using a polyclonal antibody directed against β-arrestin, and incubated overnight at 4° C. Cover slips were mounted using 50% glycerol and immunostaining was analyzed by optical sectioning using a Nikon Eclipse E800 laser scanning confocal microscope powered by Compix Confocal Imaging Systems software (Simple PCI Version 4.0.6.1605). Three-dimensional analysis of OA1-GFP and β -arrestin distribution was performed in Image J 1.32. Measurement of [Ca²⁺]i

OA1-GFP expressing CHO cells plated on glass cover slips were rinsed in Ca²⁺ containing HEPES buffered Hanks Balanced Salt Solution (HBSS) (pH 7.45), then incubated with 2.5 µM Fura-2 (solubilized in anhydrous dimethylsul- 20 foxide and 0.002% pluronic acid) for 20 minutes at 37° C., 5% CO₂. The Fura-2 loaded cells were rinsed with HBSS for 15 minutes at 37° C., 5% CO₂ to allow for full cleavage of the dye to its active form. Each cover slip was incubated in 1 ml of HBSS in a chamber held at 37° C. on the stage of 25 wild type mice, and showed that wild-type mice secreted an inverted Olympus IX70 microscope equipped with a 40×1.35 NA UV-fluor objective.

Using a filter wheel, excitation light from a 200 W Xe bulb was passed alternately through 340 and 380 nm filters. A 10 nm bandpass filter, centered at 510 nm, selected for the 30 emitted fluorescence which was passed to a CCD camera (Photometrics CH-250). For each experiment, image pairs were taken every minute for the first three minutes, which established a stable baseline. Then L-DOPA (1 µM final concentration) was added and image sets were taken every 35 30 seconds for the next three minutes. Finally, KCl (20 mM final concentration) was added one minute before completion of each experiment as a positive control to establish that the cells were loaded with Fura-2. The same was repeated independently for tyrosine and dopamine (both at 1 µM final 40 concentration). Using a Silicon Graphics Personal IRIS computer, the 340/380 nm ratio was computed for each pixel within a cell, and then analyzed using Microsoft Excel version 4.0 (Microsoft, Redmond, Wash.). Once the 340/380 nm ratio was determined, each ratio was normalized to 1 45 (ratio at time zero divided by itself), then the free ion concentration was calculated using the following equation:

$$[Ca_i]\#=Kd\#*(R-R_{min}\#)/R_{max}\#-R)$$

in which R, R_{min} , and R_{max} are the measured, minimum, and 50 maximum ratios, respectively. R_{max} represents the ratio of fluorescence intensity of ion-sensitive wavelengths under fully deprotonated conditions, whereas R_{min} is the ratio for the dye when it is fully protonated. In the case of Fura-2, R increases with increasing Ca^{2+} ; hence R_{min} represents Fura-2 55 in the absence of Ca^{2+} (Ca^{2+} <1 nM) whereas R_{max} represents the Ca²⁺-Fura-2 chelate as previously described [26]. R_{min} , R^{max} and Kd were determined in independent experiments in Fura-2 loaded cells, and subsequently utilized for calculation of free Ca²⁺ for the experimental procedures. Radiolabeled Ligand Binding

CHO cells were transfected to express OA1-GFP were plated into 24-well plates. Cells were chilled to -2 C, then rinsed in cold binding buffer, 25 mM Tris, 150 mM NaCl, 5 mM EDTA, 5 µM digitonin (pH 7.45). Cells were incubated 65 for two hours in binding buffer containing [3H]-L-DOPA (Moravek Biochemicals, Brea, Calif.) at concentrations

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between $10^{-4} \mathrm{M}$ to $10^{-9} \mathrm{M}$. The temperature was not allowed to exceed -2° C. at any step of the assay. Controls included assays conducted on nontransfected CHO and specific binding was determined by competition with excess unlabelled L-DOPA at 10⁻³M. Bound L-DOPA was quantified by scintillation spectroscopy.

Measurement of cAMP

Cells were pretreated with forskolin (15 minutes) then challenged with L-DOPA using an assay setup as previously described [47]. After 1 minute of ligand exposure, cells are scraped into ice-cold buffer, boiled then centrifuged. Equivalent volumes, 50 μl, of supernate and ³H-cAMP (New England Nuclear) then combined with 100 µl cold PKA. After 2 hours, the solution is passed over activated charcoal, 15 and supernates are counted in a scintillation counter. Results are compared to those achieved using a standard curve, instead of cytosol, produced using 50 µl of cAMP 0.25-32.0 pmole/50 µl.

Example 2

The OA1 Loop Functions in vivo

PEDF secretion in OA deficient mice was compared to significantly more PEDF than OA1 -/y mice. The culture medium (C.M.) used contains PEDF, and it is likely that PEDF in the CM from OA1 -/y is from the medium used, not the RPE. Results (FIG. 7) are quantified and summarized in the graph. The difference, even with the background PEDF in the CM for both groups is significant. T-test analysis results are presented

Tyrosinase deficient pregnant mice were maintained under normal conditions (No L-DOPA), or supplemented with 1.0 mg/ml L-DOPA in there drinking water, beginning on embryonic day 7 for their pups. Animals were maintained on supplemental until post-natal day 14, when ocular development is over and the eyes are open.

Two cell types are reduced in number in albinism: retinal ganglion cells and photoreceptors. FIG. 8A demonstrates that L-DOPA supplementation increases retinal ganglion cell numbers compared to what is expected in a normal wildtype mouse. FIG. 8B shows the same result for photoreceptors. Photoreceptors are not counted directly as they are too dense. Rather, the area occupied by photoreceptor nuclei is measured as a measure of photoreceptor numbers. L-DOPA supplementation increased the photoreceptor nuclear area, so the number of photoreceptors were increased. Again, this appeared to restore the albino animal to normal levels.

As shown in FIG. 8C, Four paired littermate animals, 2 wild-type and 2 OA1 -/y (female OA1 deficient) were euthanized and the retinas from each animal were loaded independently in a lane, then proteins were western blotted to detect PEDF, which was readily observed in the retina from wild-type mice. In contrast, PEDF is not readily detected in the retinas from the OA1 -/y mice.

In summary this data illustrate that OA1 -/y mice make less PEDF than wild type mice. L-DOPA stimulation in tyrosinase defective mice rescues the two most prominent neurosensory retina defects of albinism: a loss of photoreceptor cells and retinal ganglion cells. Finally, PEDF levels are reduced in the retinas of mice lacking OA1. Thus, it is concluded that the OA1 autocrine loop functions in vivo, and can be stimulated with oral L-DOPA.

The data together illustrate that the linkage between RPE pigmentation and AMD are likely through the signaling activity of OA1. The data illustrate that the ligand for OA1

is L-DOPA, and that OA1 signaling from L-DOPA controls the expression of PEDF. PEDF is the most potent neurotrophic factor made by RPE. Thus, the identification of L-DOPA as the ligand for OA1, which controls PEDF expression, ties together L-DOPA and neurotrophic activity in the RPE. Because L-DOPA is produced as a by-product of pigment production, this established for the first time a linkage between RPE pigmentation and neurotrophic activity. This system is defined as the OA1 autocrine loop. Tyrosinase makes pigment and releases L-DOPA. Released L-DOPA binds to and initiates signaling through OA1. OA1 signaling controls the expression of both tyrosinase and

To date the data illustrate this model biochemically, in $_{15}$ cultured cells, and in vivo. The fact that retinal development in an albino animal can be rescued using dietary L-DOPA indicates that dietary L-DOPA can be used to stimulate RPE trophic factor expression in vivo. AMD is clearly tied to an RPE defect somehow related to its pigmentation. Blue-eyed 20 individuals get AMD at a much greater frequency than dark-eyed individuals, so the level of RPE pigmentation controls the AMD process. The level of RPE pigmentation is controlled by OA1 signaling and is part of the same OA1 autocrine loop described above. Thus, AMD is related to 25 17. Staleva L, Orlow S J (2006) Ocular albinism 1 protein: OA1 signaling in RPE. Therefore, those with lower RPE pigmentation will have lower tyrosinase, lower L-DOPA, lower OA1 signaling, and lower PEDF production. We can use dietary L-DOPA or related compounds as ligands for OA1 and stimulate that activity. The final determinant of the health of the neurosensory retina is PEDF, but we can use OA1 signaling to increase the OA1 loop activity, and increase the neurotrophic activity of the RPE. The effect of OA1 signaling will be to foster neuron survival.

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Ala Ser Leu Leu Lys Gly Arg Gln Gly Ile Tyr Thr Glu Asn 245 250	Glu Arg 255				
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Leu Ile Ile Cys Trp Leu Ser Asn Ile Ile Asn Glu Ser Leu 275 280 285	Leu Phe				
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Arg Thr Ala Ala Lys Thr Thr Trp Phe Ile Met Gly Ile Leu 305 310 315	Asn Pro 320				
Ala Gln Gly Phe Leu Leu Ser Leu Ala Phe Tyr Gly Trp Thr 325 330	Gly Cys 335				
Ser Leu Gly Phe Gln Ser Pro Arg Lys Glu Ile Gln Trp Glu 340 345 350					
Thr Thr Ser Ala Ala Glu Gly Ala His Pro Ser Pro Leu Met 355 360 365	Pro His				
Glu Asn Pro Ala Ser Gly Lys Val Ser Gln Val Gly Gly Gln 370 375 380	Thr Ser				
Asp Glu Ala Leu Ser Met Leu Ser Glu Gly Ser Asp Ala Ser 385 390 395	Thr Ile 400				
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Tyr Tyr Pro Ser Val Ser Arg Cys Glu Arg Gly Leu Asp His Ala Ile 180 \$185\$

Pro His Tyr Val Thr Thr Tyr Leu Pro Leu Leu Leu Val Leu Val Ala 195 200 205

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Asn Pro Ile Leu Phe His Lys Thr Val Thr Ser Val Ala Ser Leu Leu	
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1	AIA	ser	PIO	5	цец	GIU	1111	rne	10	Сув	PIO	ASII	AIG	Asp 15	PIO	
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Glu Arg Arg Met Gly Ala Arg Ile Lys Thr Arg Phe Phe Lys Ile Met 245 250 255	
Leu Val Phe Ile Val Cys Trp Phe Ser Asn Val Ile Asn Glu Ser Leu 260 265 270	
Leu Phe Tyr Leu Glu Met Gln Pro Asp Ile Asn Ser Ser Ser Leu Lys 275 280 285	
Gln Val Arg Asn Ala Ala Lys Thr Thr Trp Phe Met Met Gly Ile Leu 290 295 300	

57 58

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Gly Cys Arg Leu Thr Leu Pro Gly Pro Ser Lys Glu Ile Gln Trp Asp 335

Ser Met Thr Thr Ser Ala Thr Glu Gly Ala Pro Pro Ser Pro Gly Gly 345

Pro Gln Glu Pro Gly Gly Gly 366

Gly Thr His Thr Ser Asp Glu Ala Leu Ser Leu Leu Ser Glu Gly 366

Gly Gly Ser Thr Ile Glu 370

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Cys Leu Gly Ser Ala Ser Ala Asn Leu Leu Leu Ser Ile Phe Gln Leu 35 40 45												
Leu Pro Lys Arg Gly Gln Gly Pro Arg Lys Leu Thr Gln Thr Ser Ser 50 55 60												
50 55 60 Ala Ser Ile Leu Leu Phe Ile Ser Ala Cys Asp Leu Leu Gly Cys Leu												
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Ala Ser Ile Leu Leu Phe Ile Ser Ala Cys Asp Leu Leu Gly Cys Leu 65 70 70 75 80 Gly Val Ile Phe Arg Ser Thr Val Trp Leu Gly Phe Pro Asp Phe Val 85 90 95 Gly Asn Ile Ser Val Val Asn Gly Thr Asp Gly Trp Pro Ser Ala Phe												
Ala Ser Ile Leu Leu Phe Ile Ser Ala Cys Asp Leu Leu Gly Cys Leu 70 75 80 Gly Val Ile Phe Arg Ser Thr Val Trp Leu Gly Phe Pro Asp Phe Val 95 Gly Asn Ile Ser Val Val Asn Gly Thr Asp Gly Trp Pro Ser Ala Phe 100 Cys Val Gly Ser Ala Met Trp Ile Gln Leu Leu Tyr Ser Ala Cys Phe												
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<210> SEQ ID NO 14
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Cys Leu Gly Ser Gly Leu Val Ser Leu Leu Leu Thr Ile Val Gln Leu 35 40 45

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<211> LENGTH: 400

<212> TYPE: PRT

<213 > ORGANISM: Xenopus laevis

<400> SEOUENCE: 14

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Ser	Pro	Gly	Phe 100	Ile	Ser	Asn	Met	Ser 105	Leu	Met	Asn	Thr	Ser 110	Asp	Ile
Trp	Pro	Ser 115	Ser	Phe	CÀa	Val	Gly 120	Ser	Ala	Met	Trp	Ile 125	Gln	Leu	Phe
Tyr	Ser 130	Ala	Ser	Phe	Trp	Trp 135	Leu	Phe	Cys	Tyr	Ala 140	Ile	Asp	Ala	Tyr
Leu 145	Val	Val	Arg	Arg	Ser 150	Ala	Gly	Ile	Ser	Thr 155	Ile	Val	Leu	Tyr	His 160
Met	Met	Thr	Trp	Gly 165	Leu	Ala	Leu	Met	Leu 170	Cys	Val	Glu	Gly	Val 175	Ala
Met	Leu	Tyr	Tyr 180	Pro	Ser	Val	Ser	Asn 185	Cys	Glu	Asn	Gly	Leu 190	Glu	His
Ala	Ile	Pro 195	His	Tyr	Val	Thr	Thr 200	Tyr	Ala	Pro	Leu	Leu 205	Ile	Val	Met
Phe	Ala 210	Asn	Pro	Ile	Leu	Phe 215	Arg	Arg	Thr	Val	Ala 220	Ala	Val	Ala	Ser
Leu 225	Leu	Lys	Gly	Arg	Gln 230	Gly	Ile	Tyr	Thr	Glu 235	Asn	Glu	Arg	Arg	Leu 240
Gly	Thr	Glu	Ile	Gln 245	Leu	Arg	Phe	Phe	Lys 250	Ile	Met	Leu	Val	Phe 255	Met
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Glu	Met	Gln 275	Pro	Asp	Ile	ГÀа	Thr 280	Asp	Gln	Leu	Lys	Asn 285	Val	Arg	Asn
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Asp	Phe	Asn	Phe	Arg 325	Gln	Lys	Glu	Thr	Ala 330	Trp	Glu	Arg	Val	Ser 335	Thr
Ser	Ser	Leu	Thr 340	Glu	Ala	Ala	His	Asn 345	Gly	Thr	Asn	Gly	Ser 350	Phe	Leu
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Ser	Gln 370	Gln	Thr	Asp	Glu	Ala 375	Leu	Ser	Ile	Leu	Ser 380	Glu	Gly	Asn	Gly
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Cys Ile Gly Ser Ala s	Ser Ala Ser Leu Leu Leu 40	Thr Ile Leu Gln Leu 45	
Leu Pro Lys Lys Gly (In Ser Leu Arg Lys Met 55	Pro Lys Ala Ser Ser 60	
	eu Leu Ile Ser Val Cys 70 75	Asp Ile Leu Gly Gly	
Ser Gly Val Ile Phe 2	arg Ser Ser Val Trp Leu	Gly Phe Pro Ser Phe	

 Ile Ala Asn Ile Ser Val Ala Asn 100
 Asn 61y Thr Asp Ile Trp Pro Ser Ala 105
 Ile Trp Pro Ser Ala 100
 Ile Trp Pro Ser Ala 110
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Arg Ser Ala Gly Arg Ser Thr Ile Val Leu Tyr 145 150 155	
Gly Leu Ala Val Leu Leu Cys Met Glu Gly Val 165 170	Met Leu Leu Tyr Tyr 175
Pro Ser Leu Ser Ser Cys Glu Arg Gly Leu Glu 180 185	His Ala Ile Pro His
Tyr Ile Thr Thr Tyr Ala Pro Leu Leu Val	Leu Val Val Asn Pro 205
Val Leu Phe Arg Arg Thr Val Thr Ala Val Ala 210 215	Ser Leu Leu Lys Gly 220
Arg Gln Gly Ile Tyr Thr Glu Asn Glu Arg Arg 225 230 235	
Gln Met Arg Phe Phe Lys Ile Met Leu Val Phe 245 250	Thr Val Cys Trp Ser 255
Ser Asn Ile Ile Asn Glu Ser Leu Leu Phe Tyr 260 265	Leu Glu Met Gln Pro 270
Asp Ile Asn Glu Thr Pro Leu Lys Asn Ile Arg 275 280	Ser Ala Ala Leu Ile 285
Thr Trp Ile Ile Met Gly Val Leu Asn Pro Met 290 295	Gln Gly Phe Leu Phe 300
Thr Leu Ala Phe Tyr Gly Trp Thr Gly Trp Lys 305 310 315	
Gln Lys Arg Glu Ile Pro Trp Glu Ser Met Ser 325 330	Ser Ser Thr Val Gly 335
Asp Asn Asp Tyr Pro Ser Pro Val Asn Tyr Gln 340 345	Ser Asn Val His Asp 350
Ser Lys Lys Ile Ser Thr Thr Asp Ser Gln Gln 355 360	Thr Asp Glu Ala Ile 365
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aaaaaaaaa	aaaaaaaaa	aaaaaaaaa	aa			1712

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<213> ORGANISM: Danio rerio

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Cys Val Cys Ser Ala Ala Leu Ser Ser Gly Leu Ala Leu Leu Gln Ile

Leu Pro Lys Arg Arg Ser Phe Arg Pro Gln Ala His Ser Ser Arg Ala

Ala Ser Ser Ser Arg Ile Leu Thr Ile Ile Ser Val Cys Asp Ile Leu

Gly Cys Thr Gly Ile Ile Ile Arg Ser Ser Leu Trp Ile Gly Leu Pro

Asn Leu Val Ser Glu Ile Ser Asp Gly Asn Ser Ser Ser Val Trp Pro 100 105

Gln Val Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Phe Phe Ser 120

Ala Ser Phe Trp Trp Thr Phe Cys Tyr Ala Val Asp Val Phe Leu Val 135

Val Lys Arg Ser Ala Gly Ile Ser Thr Ile Ile Leu Tyr His Met Ile 150 155

Thr Trp Gly Leu Thr Leu Leu Cys Val Glu Gly Val Ala Met Leu 165 170

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Tyr Tyr Pro Ser Ile Ser Ser Cys Glu Asn Gly Leu Gln His Ala Ile 180 \$180\$

			100					103					190			
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Asn	Pro 210	Val	Leu	Phe	Thr	Arg 215	Thr	Val	Ser	Ala	Val 220	Thr	Ser	Leu	Leu	
Lys 225	Gly	Gln	Gln	Gly	Ile 230	Tyr	Thr	Glu	Asn	Glu 235	Arg	Arg	Leu	Gly	Ser 240	
Glu	Ile	Lys	Ile	Arg 245	Phe	Phe	Lys	Ile	Met 250	Leu	Val	Phe	Phe	Ile 255	Cys	
Trp	Leu	Pro	Asn 260	Ile	Ile	Asn	Glu	Ser 265	Leu	Leu	Phe	Tyr	Leu 270	Glu	Met	
Gln	Asp	Asp 275	Val	Lys	Ser	Ser	Asp 280	Leu	Lys	Asn	Ile	Arg 285	Asn	Ala	Ala	
Leu	Ile 290	Thr	Trp	Phe	Ile	Met 295	Gly	Ile	Leu	Asn	Pro 300	Met	Gln	Gly	Phe	
Leu 305	Asn	Thr	Leu	Ala	Phe 310	His	Gly	Trp	Thr	Gly 315	Leu	Asp	Leu	Asp	Phe 320	
Ser	Arg	Gln	Arg	Arg 325	Arg	Glu	Leu	Pro	Trp 330	Asp	Ser	Ala	Ser	Thr 335	Ser	
Leu	Ala	Gly	Gly 340	Phe	Thr	Pro	Val	Val 345	Gly	Ser	Ser	Leu	Ile 350	Tyr	Gln	
Ser	His	Val 355	Gln	Glu	Ile	Lys	Tys	Asn	Leu	Ser	Ala	Asn 365	Gly	Gly	Gln	
Gln	Pro 370	Ser	Asp	Ala	Ile	Ser 375	Val	Leu	Ser	Glu	Asp 380	Ser	Glu	Ser	Ser	
Thr 385	Val	Glu	Ile	His	Ile 390	Ser	Ser	Glu	Gln	Arg 395	Glu	Phe	Glu	Glu	Leu 400	
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aact	ttgg	gtc t	ctct	gtct	t go	etete	gtege	c cca	aggct	gga	aato	gactt	tg (gttt	cctct	600
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aacatgaaga	cctcaatgaa	gtagccaaac	tcttcaacat	tcatgaagat	tgaatcccta	1920	
tcatggaata	cacaagcact	tttccatccc	tgacagggaa	tgggtggata	aaagaacatt	1980	
ttttattcag	catactttt	ctttatgtag	gagcaggaat	cgaacaagcc	tctgtgaata	2040	
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geege						2105	

<210> SEQ ID NO 28

<211> LENGTH: 612

<212> TYPE: PRT

<213 > ORGANISM: Rattus norvegicus

<400> SEQUENCE: 28

Met Phe Arg Cys Gly Gly Leu Ala Gly Ala Phe Lys Gln Lys Leu Val 1 $$ 5 $$ 10 $$ 15

Pro Leu Val Arg Ser Val Cys Val Gln Arg Pro Lys Gln Arg Asn Arg

Leu Pro Gly Asn Leu Phe Gln Gln Trp Arg Val Pro Leu Glu Leu Gln $35\,$

Met Ala Arg Gln Met Ala Ser Ser Gly Pro Ser Gly Gly Lys Met Asp 50 $\,$ 60

Asn Ser Val Leu Val Leu Ile Val Gly Leu Ser Thr Ile Gly Ala Gly 65 70 75 80

Ala Tyr Ala Tyr Lys Thr Ile Lys Glu Asp Gln Lys Arg Tyr Asn Glu 85 90 95

Arg Ile Met Gly Leu Gly Leu Ser Pro Glu Glu Lys Gln Arg Arg Ala

_															
			100					105					110		
Ile	Ala	Ser 115	Ala	Ala	Glu	Gly	Gly 120	Ser	Val	Pro	Pro	Ile 125	Arg	Val	Pro
Ser	His 130	Val	Pro	Phe	Leu	Leu 135	Ile	Gly	Gly	Gly	Thr 140	Ala	Ala	Phe	Ala
Ala 145	Ala	Arg	Ser	Ile	Arg 150	Ala	Arg	Asp	Pro	Gly 155	Ala	Arg	Val	Leu	Ile 160
Val	Ser	Glu	Asp	Pro 165	Glu	Leu	Pro	Tyr	Met 170	Arg	Pro	Pro	Leu	Ser 175	Lys
Glu	Leu	Trp	Phe 180	Ser	Asp	Asp	Pro	Asn 185	Val	Thr	ГÀа	Thr	Leu 190	Gln	Phe
Arg	Gln	Trp 195	Asn	Gly	Lys	Glu	Arg 200	Ser	Ile	Tyr	Phe	Gln 205	Pro	Pro	Ser
Phe	Tyr 210	Val	Ser	Ala	Gln	Asp 215	Leu	Pro	His	Ile	Glu 220	Asn	Gly	Gly	Val
Ala 225	Val	Leu	Thr	Gly	Lys 230	ГÀа	Val	Val	His	Leu 235	Asp	Val	Arg	Gly	Asn 240
Met	Val	Lys	Leu	Asn 245	Asp	Gly	Ser	Gln	Ile 250	Thr	Phe	Glu	Lys	Сув 255	Leu
Ile	Ala	Thr	Gly 260	Gly	Thr	Pro	Arg	Ser 265	Leu	Ser	Ala	Ile	Asp 270	Arg	Ala
Gly	Ala	Glu 275	Val	Lys	Ser	Arg	Thr 280	Thr	Leu	Phe	Arg	Lys 285	Ile	Gly	Asp
Phe	Arg 290	Ala	Leu	Glu	ГÀа	Ile 295	Ser	Arg	Glu	Val	300 TÀa	Ser	Ile	Thr	Val
305	_	-	_		310		Ser			315	-			_	320
ГÀа	Ser	Gln	Ala	Ser 325	Gly	Ile	Glu	Val	Ile 330	Gln	Leu	Phe	Pro	Glu 335	Lys
Gly	Asn	Met	Gly 340	ГÀЗ	Ile	Leu	Pro	Glu 345	Tyr	Leu	Ser	Asn	Trp 350	Thr	Met
Glu	Lys	Val 355	Lys	Arg	Glu	Gly	Val 360	ГÀа	Val	Met	Pro	Asn 365	Ala	Ile	Val
Gln	Ser 370	Val	Gly	Val	Ser	Gly 375	Gly	ГÀа	Leu	Leu	Ile 380	ГÀЗ	Leu	Lys	Asp
Gly 385	Arg	Lys	Val	Glu	Thr 390	Asp	His	Ile	Val	Thr 395	Ala	Val	Gly	Leu	Glu 400
Pro	Asn	Val	Glu	Leu 405	Ala	ГÀа	Thr	Gly	Gly 410	Leu	Glu	Ile	Asp	Ser 415	Asp
Phe	Gly	Gly	Phe 420	Arg	Val	Asn	Ala	Glu 425	Leu	Gln	Ala	Arg	Ser 430	Asn	Ile
Trp	Val	Ala 435	Gly	Asp	Ala	Ala	Cys 440	Phe	Tyr	Asp	Ile	Lys 445	Leu	Gly	Arg
Arg	Arg 450	Val	Glu	His	His	Asp 455	His	Ala	Val	Val	Ser 460	Gly	Arg	Leu	Ala
Gly 465	Glu	Asn	Met	Thr	Gly 470	Ala	Ala	Lys	Pro	Tyr 475	Trp	His	Gln	Ser	Met 480
Phe	Trp	Ser	Asp	Leu 485	Gly	Pro	Asp	Val	Gly 490	Tyr	Glu	Ala	Ile	Gly 495	Leu
Val	Asp	Ser	Ser 500	Leu	Pro	Thr	Val	Gly 505	Val	Phe	Ala	Lys	Ala 510	Thr	Ala
Gln	Asp	Asn 515	Pro	Lys	Ser	Ala	Thr 520	Glu	Gln	Ser	Gly	Thr 525	Gly	Ile	Arg

89 90

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 Ser
 Glu
 Ser
 Glu
 Thr
 Glu
 Ser
 Glu
 Ala
 Ser
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 Ala
 Ser
 Glu
 Ala
 Ser
 Glu
 Ala
 Fro
 Pro
 Ala
 Val
 Pro
 Glu
 Glu
 Glu
 Asp
 Tyr
 Gly
 560

 Lys
 Gly
 Val
 Ile
 Phe
 Tyr
 Leu
 Arg
 Asp
 Lys
 Val
 Val
 Gly
 Ile
 Val
 Ile
 Ile

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<210> SEQ ID NO 29

<211> LENGTH: 1486

<212> TYPE: DNA

<213> ORGANISM: Taeniopygia guttata

<400> SEQUENCE: 29

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Pro	Val	Asn	Lys 20	Leu	Ala	Ala	Ala	Val 25	Ser	Asn	Phe	Gly	Tyr 30	Asp	Leu
Tyr	Arg	Gln 35	Gln	Ser	Ile	Arg	Thr 40	Ala	Thr	Ala	Asn	Val 45	Leu	Leu	Ser
Pro	Phe 50	Ser	Leu	Ala	Thr	Ala 55	Leu	Ser	Gly	Leu	Ser 60	Leu	Gly	Ala	Gly
Glu 65	Arg	Thr	Glu	Asp	Val 70	Ile	Ser	Arg	Ala	Leu 75	Phe	Tyr	Asp	Leu	Leu 80
Asn	ГÀа	Ala	Glu	Val 85	His	Asp	Thr	Tyr	90 Lys	Glu	Leu	Leu	Ser	Ser 95	Val
Thr	Gly	Pro	Glu 100	Lys	Ser	Met	Lys	Ser 105	Ala	Ser	Arg	Ile	Ile 110	Leu	Glu
ГÀа	Arg	Leu 115	Arg	Ala	Arg	Pro	Gly 120	Phe	His	Ser	Gln	Leu 125	Glu	Lys	Ser
Tyr	130 Lys	Met	Arg	Pro	Arg	Ala 135	Leu	Ser	Gly	Asn	Thr 140	Gln	Leu	Asp	Leu
Gln 145	Glu	Ile	Asn	Thr	Trp 150	Val	Arg	Gln	Gln	Thr 155	ГÀа	Gly	Arg	Ile	Met 160
Arg	Phe	Met	Lys	Asp 165	Met	Pro	Thr	Asp	Val 170	Ser	Ile	Leu	Leu	Ala 175	Gly
Ala	Ala	Phe	Phe 180	Lys	Gly	Thr	Trp	Lys 185	Thr	Lys	Phe	Asp	Thr 190	Lys	Arg
Thr	Ala	Leu 195	Gln	Asp	Phe	His	Leu 200	Asp	Glu	Asp	Arg	Thr 205	Val	Lys	Val
Ser	Met 210	Met	Ser	Asp	Pro	Lys 215	Ala	Ile	Leu	Arg	Tyr 220	Gly	Phe	Asp	Ser
Glu 225	Leu	Asn	Cys	Lys	Ile 230	Ala	Gln	Leu	Pro	Leu 235	Thr	Glu	Gly	Ile	Ser 240
Ala	Met	Phe	Phe	Leu 245	Pro	Thr	ГЛа	Val	Thr 250	Gln	Asn	Met	Thr	Leu 255	Ile
Glu	Glu	Ser	Leu 260	Thr	Ser	Glu	Phe	Val 265	His	Asp	Val	Asp	Lys 270	Glu	Leu
Lys	Thr	Val 275	His	Ala	Val	Leu	Ser 280		Pro	Lys	Leu	Lys 285	Leu	Asn	His
Glu	Glu 290	Ala	Leu	Gly	Ser	Thr 295	Leu	ГÀа	Glu	Thr	Arg 300	Leu	Gln	Ser	Leu
Phe 305	Thr	Ser	Pro	Asp	Phe 310	Ser	ГЛа	Ile	Ser	Ala 315	ГÀа	Pro	Leu	Arg	Leu 320
Ser	His	Val	Gln	His 325	Lys	Ala	Met	Leu	Glu 330	Leu	Gly	Glu	Asp	Gly 335	Glu
Arg	Ser	Thr	Pro 340	Asn	Ala	Gly	Ala	Asn 345	Ala	Ala	Arg	Leu	Thr 350	Phe	Pro
Ile	Glu	Tyr 355	His	Val	Asp	Arg	Pro 360	Phe	Leu	Leu	Val	Leu 365	Arg	Asp	Asp
Thr	Thr 370	Gly	Thr	Leu	Leu	Phe	Ile	Gly	Lys	Ile	Leu 380	Asp	Pro	Arg	Gly

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385 <210> SEQ ID NO 31 <211> LENGTH: 1464 <212> TYPE: DNA <213> ORGANISM: Equus caballus <400> SEQUENCE: 31 ttaaaagttt tgtgcttgct ggagccccct cagtgtgcag acctaggctg ggcgcggagc 60 tgcagcacac ccacaggccc cgggatgcag gccctaatgc tactcctctg gactggagcc ctccttgggc atggcagctg ccagaacaac gccggcggcc cagaggaggg ctccccagac cctgacatca caggggcacc agtggaggag gaggatcctt tcctcaaggt ccctgtgaac aagetggcag eggeegtete caactttgge tatgacetgt acegegegaa atecageatg ageoccaceg ccaatgtget cetgteecca etcagegtgg ccacageact etetgeeett tcgctggggg cggaacagcg gacagagtcc agcattcacc tggctctcta ctatgacctg 420 atcaaqaacc caqacatcca cqqcacctac aaqqaactcc ttqcqtccqt cactqcccc 480 aataagaact tcaagagcgc ttcccgaatc atcttcgaga agaagctgcg catcaaatcc 540 600 agetttqtta caccactqqa qaaqtcatat qqqaccaqqc ccaaqateet qaetqqcaac tctcgcacgg atcttcagga gattaacaac tgggtgcagg cccagatgaa agggaaaatt 660 gctaggtcca caagggaagt gcccagtgaa atcagcattc tccttctcgg tgtggcttac 720 ttcaaggggc agtgggtaac aaagtttgac tccagaaaga cttccctcca ggatttccac 780 ttggatgagg agaggaccgt gacagtcccc acgatgtcag atccgaaggc cattctacgc 840 tacggettgg attetgatet caactgtaag ategeceage tacceetgae eggaageatg 900 agcategtet tetteetgee teagaaagtg acceagaace tgaceatgat agaagagage 960 ctcacctccg agttccttca tgacatagac cgagagctga agactgtgca ggcagtcctg 1020 accatececa agetgaaget gagttatgag ggtgaagtea etaagteeet geaggagata 1080 aagctgcaat cettgtttga ttcaccagae tttagcaaga tcacaggcaa acetetcaag 1140 cttactcaag tggaacatcg tgctggcttt gagtggaatg aggatggggc aaccaacccc 1200 agecaaggge eccageetge ecaceteace tteecettgg actaceacet taaccaacet 1260 1320 ttcatctttg tactgaggga cacggacaca ggggcccttc tcttcatagg caaaattctg gaccccaggg gcacttaatg ctctagctta atgttcaaat accctagatg aagaaaaccc 1380 tagagggatg gcagattata tattacgtga aggctgccct ataatgtttc aatgtatcct 1440 tttcaataaa agtgctttat cctt 1464 <210> SEQ ID NO 32 <211> LENGTH: 417 <212> TYPE: PRT <213 > ORGANISM: Equus caballus <400> SEQUENCE: 32 Met Gln Ala Leu Met Leu Leu Leu Trp Thr Gly Ala Leu Leu Gly His 10 Gly Ser Cys Gln Asn Asn Ala Gly Gly Pro Glu Glu Gly Ser Pro Asp 25 Pro Asp Ile Thr Gly Ala Pro Val Glu Glu Glu Asp Pro Phe Leu Lys

40

55

50

Val Pro Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp

Leu Tyr Arg Ala Lys Ser Ser Met Ser Pro Thr Ala Asn Val Leu Leu Ser Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln Arg Thr Glu Ser Ser Ile His Leu Ala Leu Tyr Tyr Asp Leu Ile Lys Asn Pro Asp Ile His Gly Thr Tyr Lys Glu Leu Leu Ala Ser Val Thr Ala Pro Asn Lys Asn Phe Lys Ser Ala Ser Arg Ile Ile Phe Glu Lys Lys Leu Arg Ile Lys Ser Ser Phe Val Thr Pro Leu Glu Lys Ser Tyr Gly Thr Arg Pro Lys Ile Leu Thr Gly Asn Ser Arg Thr Asp Leu Gln Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Ile 185 Ala Arg Ser Thr Arg Glu Val Pro Ser Glu Ile Ser Ile Leu Leu Leu 200 Gly Val Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr Ser Leu Gln Asp Phe His Leu Asp Glu Glu Arg Thr Val Thr 230 Val Pro Thr Met Ser Asp Pro Lys Ala Ile Leu Arg Tyr Gly Leu Asp 250 Ser Asp Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met 265 260 Ser Ile Val Phe Phe Leu Pro Gln Lys Val Thr Gln Asn Leu Thr Met 280 Ile Glu Glu Ser Leu Thr Ser Glu Phe Leu His Asp Ile Asp Arg Glu 295 Leu Lys Thr Val Gln Ala Val Leu Thr Ile Pro Lys Leu Lys Leu Ser Tyr Glu Gly Glu Val Thr Lys Ser Leu Gln Glu Ile Lys Leu Gln Ser Leu Phe Asp Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Leu Lys Leu Thr Gln Val Glu His Arg Ala Gly Phe Glu Trp Asn Glu Asp Gly 360 Ala Thr Asn Pro Ser Gln Gly Pro Gln Pro Ala His Leu Thr Phe Pro Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr Asp Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly 405 410 Thr <210> SEQ ID NO 33 <211> LENGTH: 1503 <212> TYPE: DNA

<213 > ORGANISM: Xenopus (Silurana) tropicalis

<400> SEQUENCE: 33

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                                                                 180
ctacaagagt ccaatcaaca ggcttgcctc ttctgcatct aactttggat atgacctata
                                                                 240
tcgtatgcaa gcaaacaaaa atcccaacag caatatcatt atttcaccac tgagcattgc
                                                                 300
tacatctctg tcaagtcttt ccttgggggg tggacaaaga actgaatcat taatccagcg
ttctctatac tatgaccttc tcaatgatcc tgaagtccat gctacatata aagacttgct
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gcagacacaa gggaaagtgg tgaagttett caaagagatt ccaactagtg tgagcattet
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tgtccagcgt gaattccacc tcgatgaaca gacatctgtc actgttccaa tgatgtcatc
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taaaaacatc cccqtqaqat acqqcttaqa ctctqatttt aactqcaaqa ttqttcaqct
                                                                 840
tecteteact ggtggggtta gcatcatgtt ttteetgeea aacacagtea eccagaaett
                                                                 900
qactatqatt qaaqaqqcc tqacatctqa qtttqtccat qacataqacc aqqcactqca
                                                                 960
gectateaac tiggteetaa gegteeetaa actaaagetg aactatgaag eigagettaa
                                                                1020
qqaaqcactq caqqaatcaa aqctccaatc ccttttcqcc actcctqact tcaqcaaaat
                                                                1080
ctcctcaaag ccattaaagc tctcctatgt cgtacataaa gccaccttgg aattgaacga
                                                                1140
ggaaggagca gagacagcgc caaaaccaga ggacagccac cgcaattact ttcctttgga
                                                                1200
gtatcactta gatcatcctt tcttgtttgt tctccgtgcc aatgacaacg gcgctctcct
                                                                1260
1320
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1440
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aaa
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<210> SEQ ID NO 34
<211> LENGTH: 409
<212> TYPE: PRT
<213> ORGANISM: Xenopus (Silurana) tropicalis
<400> SEQUENCE: 34
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Glu Asp Pro Phe Tyr Lys Ser Pro Ile Asn Arg Leu Ala Ser Ser Ala
                          40
Ser Asn Phe Gly Tyr Asp Leu Tyr Arg Met Gln Ala Asn Lys Asn Pro
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Ser Leu Ser Leu Gly Gly Gln Arg Thr Glu Ser Leu Ile Gln Arg 85 90 95 Ser Leu Tvr Tvr Asp Leu Leu Asn Asp Pro Glu Val His Ala Thr Tvr

Asn Ser Asn Ile Ile Ile Ser Pro Leu Ser Ile Ala Thr Ser Leu Ser

75

70

Ser Leu Tyr Tyr Asp Leu Leu Asn Asp Pro Glu Val His Ala Thr Tyr \$100\$ \$100\$ \$105

Lys Asp Leu Leu Ala Ser Phe Thr Ser Gln Ala Ser Gly Leu Lys Ser 115 120 125												
Thr Trp Arg Ile Met Leu Glu Arg Arg Leu Arg Leu Arg Met Asp Phe 130 135 140												
Val Thr Gln Val Glu Lys Phe Tyr Gly Asn Lys Pro Lys Val Leu Thr 145 150 155 160												
Gly Ser Thr Arg Leu Asp Leu Gln Glu Ala Asn Asp Phe Ile Gln Lys 165 170 175												
Gln Thr Gln Gly Lys Val Val Lys Phe Phe Lys Glu Ile Pro Thr Ser 180 185 190												
Val Ser Ile Leu Leu Gly Thr Thr Tyr Leu Lys Gly Gln Trp Ala 195 200 205												
Tyr Lys Phe Asn Pro Arg Glu Thr Val Gln Arg Glu Phe His Leu Asp 210 215 220												
Glu Gln Thr Ser Val Thr Val Pro Met Met Ser Ser Lys Asn Ile Pro 225 230 235 240												
Val Arg Tyr Gly Leu Asp Ser Asp Phe Asn Cys Lys Ile Val Gln Leu 245 250 255												
Pro Leu Thr Gly Gly Val Ser Ile Met Phe Phe Leu Pro Asn Thr Val												
Thr Gln Asn Leu Thr Met Ile Glu Glu Gly Leu Thr Ser Glu Phe Val 275 280 285												
His Asp Ile Asp Gln Ala Leu Gln Pro Ile Asn Leu Val Leu Ser Val 290 295 300												
Pro Lys Leu Lys Leu Asn Tyr Glu Ala Glu Leu Lys Glu Ala Leu Gln 305 310 315 320												
Glu Ser Lys Leu Gln Ser Leu Phe Ala Thr Pro Asp Phe Ser Lys Ile 325 330 335												
Ser Ser Lys Pro Leu Lys Leu Ser Tyr Val Val His Lys Ala Thr Leu 340 345 350												
Glu Leu Asn Glu Glu Gly Ala Glu Thr Ala Pro Lys Pro Glu Asp Ser 355 360 365												
His Arg Asn Tyr Phe Pro Leu Glu Tyr His Leu Asp His Pro Phe Leu 370 375 380												
Phe Val Leu Arg Ala Asn Asp Asn Gly Ala Leu Leu Phe Ile Gly Lys												
Val Met Asp Pro Lys Gly Phe Ser Phe												
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aaccacagtt ccgggatgca ggccctggtg ctactcctct ggactggagc cctgctcggg 18												
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agtacttaat gtctcagtgc tctacagaac ccccagaggg aagctgatta tacattccag	1440												
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Asp Ser Thr Gly Glu Pro Val Glu Glu Glu Asp Pro Phe Phe Lys Val 35 40 45													
Pro Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu 50 60													
Tyr Arg Leu Arg Ser Ser Ala Ser Pro Thr Gly Asn Val Leu Leu Ser 65 70 75 80													
Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu 85 90 95													
His Arg Thr Glu Ser Val Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile 100 105 110													
Thr Asn Pro Asp Ile His Ser Thr Tyr Lys Glu Leu Leu Ala Ser Val													
Thr Ala Pro Glu Lys Asn Leu Lys Ser Ala Ser Arg Ile Val Phe Glu													
130 135 140													
Arg Lys Leu Arg Val Lys Ser Ser Phe Val Ala Pro Leu Glu Lys Ser 145 150 155 160													
Tyr Gly Thr Arg Pro Arg Ile Leu Thr Gly Asn Pro Arg Val Asp Leu													
165 170 175													

Gln Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Ile Ala 180 185 190

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Arg	Ser	Thr	Arg	Glu	Met	Pro	Ser 200	Ala	Leu	Ser	Ile	Leu 205	Leu	Leu	Gly
		193					200					203			
Val	Ala 210	Tyr	Phe	ГÀа	Gly	Gln 215	Trp	Val	Thr	ГÀа	Phe 220	Asp	Ser	Arg	ГÀв
Thr 225	Thr	Leu	Gln	Asp	Phe 230	His	Leu	Asp	Glu	Asp 235	Arg	Thr	Val	Arg	Val 240
Pro	Met	Met	Ser	Asp 245	Pro	ГЛа	Ala	Ile	Leu 250	Arg	Tyr	Gly	Leu	Asp 255	Ser
Asp	Leu	Asn	Сув 260	Lys	Ile	Ala	Gln	Leu 265	Pro	Leu	Thr	Gly	Ser 270	Met	Ser
Ile	Ile	Phe 275	Phe	Leu	Pro	Leu	Thr 280	Val	Thr	Gln	Asn	Leu 285	Thr	Met	Ile
Glu	Glu 290	Ser	Leu	Thr	Ser	Glu 295	Phe	Ile	His	Asp	Ile 300	Asp	Arg	Glu	Leu
Lys 305	Thr	Ile	Gln	Ala	Val 310	Leu	Thr	Val	Pro	Lys 315	Leu	Lys	Leu	Ser	Phe 320
Glu	Gly	Glu	Leu	Thr 325	Lys	Ser	Leu	Gln	Asp 330	Met	ГÀа	Leu	Gln	Ser 335	Leu
Phe	Glu	Ser	Pro 340	Asp	Phe	Ser	Lys	Ile 345	Thr	Gly	rys	Pro	Val 350	Lys	Leu
Thr	Gln	Val 355	Glu	His	Arg	Ala	Ala 360	Phe	Glu	Trp	Asn	Glu 365	Glu	Gly	Ala
Gly	Ser 370	Ser	Pro	Ser	Pro	Gly 375	Leu	Gln	Pro	Val	Arg 380	Leu	Thr	Phe	Pro
Leu 385	Asp	Tyr	His	Leu	Asn 390	Gln	Pro	Phe	Leu	Phe 395	Val	Leu	Arg	Asp	Thr 400
Asp	Thr	Gly	Ala	Leu 405	Leu	Phe	Ile	Gly	Arg 410	Ile	Leu	Asp	Pro	Ser 415	Ser
<21	0> SE L> LE 2> TY	ENGTI	H: 18												

<213 > ORGANISM: Salmo salar

<400> SEQUENCE: 37

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		-contir	nued	
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cgggagcact gctctttatt ggcaaggtgg	tcaacccacg	caatctgagg	atataaacac	1260
agacacacac tgccttctaa gcaggtccta	ggaggggatc	agccatcgtt	aagcttaagc	1320
ttctgtgtgt cataaatgca caatatgaga	gggtggataa	gcagctagat	ttacccattg	1380
atcatataat acagtttctt aatcatgtat	ggaaaccatg	cataacattc	agactaaaag	1440
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atttgtgatt gaaaagtaca gctctcataa	ttttaaata	gaggcacatt	ctttaacccc	1560
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tttctgtttg tctgtctgac tccatagatg	gaattgtata	actttatcca	gttgacatac	1680
aatagctgct tccagtaaag ggttgggtta	ttttggaaag	aaattggact	cttggatgct	1740
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aaaaaaaga				1810
<210> SEQ ID NO 38 <211> LENGTH: 405 <212> TYPE: PRT <213> ORGANISM: Salmo salar				
<400> SEQUENCE: 38				
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Ser Tyr Ala Gl
n Leu Leu Glu Thr Glu Ala Ala Gly Gly Glu Glu Glu 25

Ala Val Glu Leu Phe Thr Thr Pro Arg Ala Lys Met Ala Ala Ala Thr 35 40 45

Ser Asp Phe Gly Tyr Asn Leu Phe Arg Ala Leu Ala Gly Arg Asn Pro

Asn Thr Asn Val Phe Leu Ala Pro Ile Ser Ile Ser Ala Val Leu Thr 65 70 75 75 80

Gln Leu Ser Met Gly Ala Ser Pro Asp Arg Ser Glu Arg Trp Leu Tyr 85 90 95

Arg Ala Leu Arg Tyr His Thr Leu Gln Asp Pro Gln Leu His Asp Thr 105

Ile Ala Ala Arg Val Tyr Leu Ala Arg Arg Leu Arg Leu Lys Gln Glu 135

Tyr Phe Gly Val Val Glu Lys Gln Tyr Gly Val Arg Pro Lys Ala Leu 150 155

Met Gly Gly Ala Lys Asp Val Asn Glu Ile Asn Asp Trp Val Lys Gln 170

Gln Thr Gly Gly Lys Val Asp Arg Phe Met Ser Lys Pro Leu Gly Arg 185

Asn Ser Gly Val Val Pro Leu Gly Ala Ala Tyr Phe Lys Val Lys Trp

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195 200 205														
Met Thr Arg Phe Ser Gln Ser Gly Val Met Glu Asp Phe Gln Leu Val 210 215 220														
Gly Glu Ala Pro Ala Arg Ile Ser Met Met Gln Gln Asp Asn Tyr Pro 225 230 235 240														
Val Lys Met Gly Val Asp Pro Asp Leu Gly Cys Thr Ile Ala Gln Ile 245 250 255														
Gln Met Gln Asp Asp Val Ser Met Phe Val Phe Leu Pro Asp Asp Val 260 265 270														
Thr Gln Asn Met Thr Leu Val Glu Glu Ser Leu Thr Ala Glu Phe Val 275 280 285														
Gln Asp Leu Ser Met Thr Leu His Pro Val Gln Thr Ala Leu Thr Leu 290 295 300														
Pro Val Leu Lys Phe Ser Tyr Ser Thr Asp Leu Leu Pro Leu Leu Thr 305 310 315 320														
Asp Leu Gly Leu Asp Glu Phe Leu Ala Asp Thr Asp Leu Thr Lys Ile 325 330 335														
Thr Ser Gln Ala Ala Lys Leu Gly Ser Leu Asn His Lys Val Val Met 340 345 350														
Glu Met Ala Pro Glu Gly Thr Gln Tyr Ala Ser Ser Leu Pro Ala Ser 355 360 365														
Thr Pro Leu Ser Tyr Cys Val Asp His Pro Phe Leu Phe Leu Val Arg 370 375 380														
Asp Glu Ala Ser Gly Ala Leu Leu Phe Ile Gly Lys Val Val Asn Pro 385 390 395 400														
Arg Asn Leu Arg Ile 405														
405 <210> SEQ ID NO 39 <211> LENGTH: 1422 <212> TYPE: DNA <213> ORGANISM: Ovis aries														
<400> SEQUENCE: 39														
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tecetggece etgagageae aggggeaece gtggaggaag aggateeett etteaaggte 180														
cccgtgaaca agctggcggc agccgtctcc aacttcggct acgacctgta ccgcgtgaga 240														
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tetgecetgt egetgggtge ggaacagegg acagaateca geatteaeeg ggetetgtae 360														
tacgacetga teagtaacee agacateeae ggeacetaea aggaceteet tgeeteegte 420														
actgcccccc agaagaacct taaaagtgct tcccggatta tctttgagag gaagctgcgg 480														
ataaaagcca gettegteec acceetegag aagteatatg ggaccaggee cagaateetg 540														
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gatttccact tggatgaggg gaggaccgtg aaagttccca tgatgtcaga ccctaaggcc 780														
gttttacggt acggcttgga ttctgatctc aactgcaaga tcgcccagct gcccttgacc 840														
gggagcacaa gtatcatctt cttcctgcct cagaaagtga cccagaactt gaccttgata 900														

-cc	ntinued
- CC	ntinued

											_	COII	CIII	ueu		
gaaq	gagaç	gec 1	caco	ctct	ga gt	tcat	tcat	gad	cataç	gacc	gaga	aacto	gaa q	gacto	gttcag	960
gcaç	gteet	ga (ccatt	ccca	aa go	ctgaa	agete	g agt	tato	gaag	gcga	aacto	cac (gaagt	ctgtg	1020
cag	gagct	ga a	agcta	acaat	cc cc	ctgtt	tgat	gca	accaç	gact	ttaç	gcaaq	gat (cacaç	ggcaaa	1080
ccta	atcaa	aac 1	tact	caaq	gt gg	gaaca	atcgo	ato	ggat	tcg	agt	ggaat	ga 🤅	ggato	ggggcg	1140
ggta	actaa	act o	ccago	cca	gg gg	gtcca	agcct	gco	eeged	ctca	ccti	ccct	ct q	ggact	atcac	1200
ctta	aacca	aac (ettt	catci	t to	gtact	gagg	g gad	cacaç	gaca	cag	gggc	cct 1	cctct	tcata	1260
ggca	aaaat	tc 1	ggad	ccca	ag ag	ggca	cttaa	a tao	ctcaa	ectt	aat	gttca	aaa 1	cacco	ccagaa	1320
gaaa	aaaaa	aca (ctago	9993	at go	gcaga	attat	ata	attat	atg	aag	gctg	ccc (ctaco	gtttca	1380
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Gly	His	Cys	Gln 20	Asn	Ala	Gly	Pro	Glu 25	Ala	Gly	Ser	Leu	Ala 30	Pro	Glu	
Ser	Thr	Gly 35	Ala	Pro	Val	Glu	Glu 40	Glu	Asp	Pro	Phe	Phe 45	Lys	Val	Pro	
Val	Asn 50	Lys	Leu	Ala	Ala	Ala 55	Val	Ser	Asn	Phe	Gly 60	Tyr	Asp	Leu	Tyr	
Arg 65	Val	Arg	Ser	Gly	Glu 70	Ser	Pro	Thr	Thr	Asn 75	Val	Leu	Leu	Ser	Pro 80	
Leu	Ser	Val	Ala	Thr 85	Ala	Leu	Ser	Ala	Leu 90	Ser	Leu	Gly	Ala	Glu 95	Gln	
Arg	Thr	Glu	Ser 100	Ser	Ile	His	Arg	Ala 105	Leu	Tyr	Tyr	Asp	Leu 110	Ile	Ser	
Asn	Pro	Asp 115	Ile	His	Gly	Thr	Tyr 120	Lys	Asp	Leu	Leu	Ala 125	Ser	Val	Thr	
Ala	Pro 130	Gln	Lys	Asn	Leu	Lys 135	Ser	Ala	Ser	Arg	Ile 140	Ile	Phe	Glu	Arg	
Lys 145	Leu	Arg		_	Ala 150		Phe		Pro			Glu	Lys	Ser	Tyr 160	
Gly	Thr	Arg	Pro	Arg 165	Ile	Leu	Thr	Gly	Asn 170	Ser	Arg	Ile	Asp	Leu 175	Gln	
Glu	Ile	Asn	Asn 180	Trp	Val	Gln	Ala	Gln 185	Met	Lys	Gly	Lys	Ile 190	Ala	Arg	
Ser	Thr	Arg 195	Glu	Ile	Pro	Ser	Gly 200	Ile	Ser	Ile	Leu	Leu 205	Leu	Gly	Val	
Ala	Tyr 210	Phe	Lys	Gly	Gln	Trp 215	Val	Thr	Lys	Phe	Asp 220	Ser	Arg	Lys	Thr	
Ser 225	Leu	Glu	Asp	Phe	His 230	Leu	Asp	Glu	Gly	Arg 235	Thr	Val	Lys	Val	Pro 240	
Met	Met	Ser	Asp	Pro 245	Lys	Ala	Val	Leu	Arg 250	Tyr	Gly	Leu	Asp	Ser 255	Aap	
Leu	Asn	Cys	Lys 260	Ile	Ala	Gln	Leu	Pro 265	Leu	Thr	Gly	Ser	Thr 270	Ser	Ile	
Ile	Phe	Phe 275	Leu	Pro	Gln	Lys	Val 280	Thr	Gln	Asn	Leu	Thr 285	Leu	Ile	Glu	

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 Glu
 Ser 290
 Leu
 Thr
 Ser 295
 Glu
 Phe 295
 Ile His Asp 300
 Arg 300
 Arg Glu
 Leu Lys 295

 Thr
 Val
 Glu
 Yal
 Leu 310
 Thr
 Ile Pro
 Leu 315
 Leu Lys Leu Glu
 Tyr
 Glu 320

 Gly
 Glu
 Leu Thr Lys 325
 Ser Val
 Gln
 Glu Leu Leu Lus 330
 Leu Lus Leu Gln
 Ser Leu 325
 Phe 325
 Leu Gln
 Leu Lus Leu Gln
 Ser Leu Gln
 Ser Jas 25
 Phe 325
 Glu Jus Leu Jus Leu Gln
 Ser Jus 25
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 Phe 325
 Glu Jus 25
 Phe 325
 Fur Jus 25
 Phe 325
 Phe 325

<210> SEQ ID NO 41

<211> LENGTH: 1465

<212> TYPE: DNA

<213> ORGANISM: Cavia porcellus

<400> SEQUENCE: 41

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<210> SEQ ID NO 42 <211> LENGTH: 418 <212> TYPE: PRT <213> ORGANISM: Cavia porcellus																
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Gly	Ser	Сув	Gln 20	Asp	Ile	Ala	Ser	Asn 25	Pro	Glu	Asp	Ser	Pro 30	Ser	Pro	
Glu	Ser	Thr 35	Gly	Glu	Pro	Val	Glu 40	Glu	Glu	Asp	Pro	Phe 45	Phe	Lys	Val	
Pro	Val 50	Asn	Lys	Leu	Ala	Ala 55	Ala	Ile	Ser	Asn	Phe 60	Gly	Tyr	Asp	Leu	
Tyr 65	Arg	Val	Arg	Ser	Ile 70	Glu	Ser	Pro	Thr	Thr 75	Asn	Val	Leu	Leu	Ser 80	
Pro	Leu	Ser	Val	Ala 85	Thr	Ala	Leu	Ser	Ala 90	Leu	Ser	Leu	Gly	Ala 95	Glu	
Gln	Arg	Thr	Glu 100	Ala	Thr	Ile	His	Arg 105	Ala	Leu	Tyr	Tyr	Asp 110	Met	Ile	
Ser	Asn	Pro 115	Asp	Ile	His	Ser	Thr 120	Tyr	Lys	Glu	Leu	Leu 125	Ala	Thr	Val	
Thr	Ala 130	Pro	Gln	Lys	Asn	Leu 135	Lys	Ser	Ala	Ser	Arg 140	Ile	Val	Phe	Glu	
Arg 145	Lys	Leu	Arg	Ile	Lуs 150	Ser	Ser	Leu	Val	Ala 155	Leu	Leu	Glu	Lys	Ser 160	
Tyr	Ser	Thr	Arg	Pro 165	Arg	Ile	Leu	Thr	Gly 170	Asn	Pro	Arg	Ile	Asp 175	Leu	
Gln	Glu	Ile	Ser 180	Asn	Trp	Val	Gln	Ala 185	Gln	Met	ГÀа	Gly	Lys 190	Ile	Thr	
Arg	Ser	Thr 195	Arg	Glu	Val	Pro	Ser 200	Gly	Ile	Ser	Ile	Leu 205	Leu	Leu	Gly	
Val	Ala 210	Tyr	Phe	Lys	Gly	Gln 215	Trp	Val	Thr	Lys	Phe 220	Asp	Ser	Arg	Lys	
Thr 225	Ser	Leu	Gln	Asp	Phe 230	His	Leu	Asp	Glu	Glu 235	Arg	Thr	Val	Lys	Val 240	
Pro	Met	Met	Ser	Asp 245	Pro	ГÀа	Ala	Ile	Ile 250	Arg	Tyr	Gly	Leu	Asp 255	Thr	
Asp	Leu	Asn	260	ГÀа	Ile	Ala	Gln	Leu 265	Pro	Leu	Thr	Gly	Ser 270	Met	Ser	
Ile	Ile	Phe 275	Phe	Leu	Pro	Met	Arg 280	Ala	Thr	Gln	Asn	Leu 285	Thr	Met	Ile	
Glu	Glu 290	Ser	Leu	Thr	Ser	Glu 295	Phe	Val	His	Asp	Ile 300	Asn	Arg	Glu	Leu	
305	Ala	Val	Gln	Ala	Val 310	Leu	Ser	Ile	Pro	Arg 315	Leu	Lys	Leu	Ser	Phe 320	
Glu	Gly	Glu	Leu	Thr 325	Lys	Ser	Leu	Gln	Glu 330	Met	Lys	Leu	His	Ser 335	Leu	
Phe	Glu	Ser	Pro 340	Asp	Phe	Ser	Lys	Ile 345	Thr	Gly	Lys	Pro	Ile 350	Lys	Leu	
Thr	Gln	Val	Glu	His	Arg	Ala	Gly	Phe	Glu	Trp	Asn	Glu	Glu	Gly	Ala	

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355 360 365 Pro Gly Thr Ser Thr Asn Ser Asp Leu Gln Pro Thr Gly Phe Thr Phe 370 375 380 Ser Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp 385 390 Thr Asp Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg 405 410 Ser Thr <210> SEQ ID NO 43 <211> LENGTH: 1408 <212> TYPE: DNA <213 > ORGANISM: Bos taurus <400> SEQUENCE: 43 gaggtgcacc cacaggcccc gagatgcagg ccctcgtgct actcctctgg actggagccc 60 120 tgcttgggtt tggccgctgc cagaacgccg gccaggaggc gggctctctg acccctgaga gcacggggc accagtggag gaagaggatc ccttcttcaa ggtccctgtg aacaagctgg 180 eggeageggt etceaactte ggetacgace tgtacegegt gagateeggt gagageecea 240 coqccaatqt qctqctqtct ccqctcaqcq tqqccacqqc qctctctqcc ctqtcqctqq 300 gtgcggaaca gcggacagaa tccaacattc accgggctct gtactacgac ctgatcagta 360 420 acccaqacat ccacqqcacc tacaaqqacc tccttqcctc cqtcaccqcc ccccaqaaqa accttaagag tgcttcccgg attatctttg agaggaagct gcggataaaa gccagcttca 480 teccaeceet ggagaagtea tatgggaeea ggeeeagaat eetgaeegge aactetegag 540 tagaccttca ggagattaac aactgggtgc aggcccagat gaaagggaaa gtcgctaggt 600 ccacgaggga gatgcccagt gagatcagca ttttcctcct gggcgtggct tacttcaagg 660 ggcagtgggt aacaaagttt gactccagaa aaacttccct ggaggatttc tacttggatg 720 aggagaggac cgtgaaagtc cccatgatgt cagaccctca ggccgtttta cggtacggct 780 tggattctga tctcaactgc aagatcgccc agctgccctt gaccgggagc acaagtatca 840 tcttcttcct gcctcagaaa gtgacccaga acttgacctt gatagaagag agcctcacct 900 ctgagttcat tcatgacata gaccgagaac tgaagactgt tcaggcggtc ctgaccattc 960 ccaagetgaa getgagttat gaaggegaae teaegaagte egtgeaggag etgaagetge aatccctgtt tgatgcacca gactttagca agatcacagg caaacctatc aaacttactc aagtggaaca tcgcgtcgga tttgagtgga atgaggatgg ggcgggtact aactccagcc 1140 caggggtcca gcctgcccgc ctcaccttcc ctctggacta tcaccttaac caacctttca tetttgtaet gagggaeaca gaeacagggg ceettetett cataggeaaa attetggaee 1260 ccaggggcac ttagtactcc aactaaatgt tcaaataccc cagaagaaaa aaacactaga 1320 gggatggcag attatatatt atacgaaggc tgcccctaca tttcaatgta tactttgcaa 1380 taaaagtgct ttatccttaa aaaaaaaa 1408 <210> SEO TD NO 44 <211> LENGTH: 416 <212> TYPE: PRT <213 > ORGANISM: Bos taurus

<400> SEOUENCE: 44

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Ser	Thr	Gly 35	Ala	Pro	Val	Glu	Glu 40	Glu	Asp	Pro	Phe	Phe 45	Lys	Val	Pro
Val	Asn 50	Lys	Leu	Ala	Ala	Ala 55	Val	Ser	Asn	Phe	Gly 60	Tyr	Asp	Leu	Tyr
Arg 65	Val	Arg	Ser	Gly	Glu 70	Ser	Pro	Thr	Ala	Asn 75	Val	Leu	Leu	Ser	Pro 80
Leu	Ser	Val	Ala	Thr 85	Ala	Leu	Ser	Ala	Leu 90	Ser	Leu	Gly	Ala	Glu 95	Gln
Arg	Thr	Glu	Ser 100	Asn	Ile	His	Arg	Ala 105	Leu	Tyr	Tyr	Asp	Leu 110	Ile	Ser
Asn	Pro	Asp 115	Ile	His	Gly	Thr	Tyr 120	Lys	Asp	Leu	Leu	Ala 125	Ser	Val	Thr
Ala	Pro 130	Gln	Lys	Asn	Leu	Lуs 135	Ser	Ala	Ser	Arg	Ile 140	Ile	Phe	Glu	Arg
Lys 145	Leu	Arg	Ile	ГÀа	Ala 150	Ser	Phe	Ile	Pro	Pro 155	Leu	Glu	ГЛа	Ser	Tyr 160
Gly	Thr	Arg	Pro	Arg 165	Ile	Leu	Thr	Gly	Asn 170	Ser	Arg	Val	Asp	Leu 175	Gln
Glu	Ile	Asn	Asn 180	Trp	Val	Gln	Ala	Gln 185	Met	Lys	Gly	Lys	Val 190	Ala	Arg
Ser	Thr	Arg 195	Glu	Met	Pro	Ser	Glu 200	Ile	Ser	Ile	Phe	Leu 205	Leu	Gly	Val
Ala	Tyr 210	Phe	Lys	Gly	Gln	Trp 215	Val	Thr	Lys	Phe	Asp 220	Ser	Arg	Lys	Thr
Ser 225	Leu	Glu	Asp	Phe	Tyr 230	Leu	Asp	Glu	Glu	Arg 235	Thr	Val	ГÀв	Val	Pro 240
Met	Met	Ser	Asp	Pro 245	Gln	Ala	Val	Leu	Arg 250	Tyr	Gly	Leu	Asp	Ser 255	Asp
Leu	Asn	CÀa	Lys 260	Ile	Ala	Gln	Leu	Pro 265	Leu	Thr	Gly	Ser	Thr 270	Ser	Ile
Ile	Phe	Phe 275	Leu	Pro	Gln	ГÀа	Val 280	Thr	Gln	Asn	Leu	Thr 285	Leu	Ile	Glu
Glu	Ser 290					295					300				
Thr 305	Val	Gln	Ala	Val	Leu 310		Ile	Pro	ГÀа	Leu 315	Lys	Leu	Ser	Tyr	Glu 320
Gly	Glu	Leu	Thr	Lys 325	Ser	Val	Gln	Glu	Leu 330	ГÀа	Leu	Gln	Ser	Leu 335	Phe
Asp	Ala	Pro	Asp 340	Phe	Ser	Lys	Ile	Thr 345	Gly	ГÀа	Pro	Ile	Lys 350	Leu	Thr
Gln	Val	Glu 355	His	Arg	Val	Gly	Phe 360	Glu	Trp	Asn	Glu	Asp 365	Gly	Ala	Gly
Thr	Asn 370	Ser	Ser	Pro	Gly	Val 375	Gln	Pro	Ala	Arg	Leu 380	Thr	Phe	Pro	Leu
Asp 385	Tyr	His	Leu	Asn	Gln 390	Pro	Phe	Ile	Phe	Val 395	Leu	Arg	Asp	Thr	Asp 400
Thr	Gly	Ala	Leu	Leu 405	Phe	Ile	Gly	Lys	Ile 410	Leu	Asp	Pro	Arg	Gly 415	Thr

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<212> TYPE: DNA <213> ORGANISM: Sus scrofa <400> SEOUENCE: 45 agtgcacgga cctaggctgg gcgtggagct gcagcgcacc cacaggcccc gggatgcagg 60 ccctcgtgct actcctctgg actggagccc tcctcgggtc tggcagctgc cagaacgctg 120 gcccggagga gggctccccg gcccctgaca cggtgggggc gccagtggag gaggaggatc 180 cettetteaa ggteeetgtg aacaagetgg eggeggeegt etecaacttt ggttaegace tgtaccgagt gagatccagc gagagcccca ccgccaacgt gctcctgtct cccctcagcg tggccacggc gctctctgcc ctgtctctgg gagccgaaca gcggacagaa tccagcctcc accgggctct ctactatgac ctgatcagca acccggacct ccacggcacc tacaaggagc 480 teettqetqe eqteactqee ecceaqaaqa aceteaaqaq tqetteeeqq ateatetttq agaagaagct gcggataaaa gccagctttg ttgcacccct ggaaaagtca tacgggacca 540 ggcccagaat tctgaccggc aactcccgct tggaccttca ggaggttaac aactgggtgc 600 660 aggeteagae gaaagggaaa gtegeeaggt ceaegeggga aetgeeegge gaaateagea tecteettet tqqtqtqqet taetteaaqq qqcaqtqqqt aaccaaqttt qaetecaqqa 720 agacqtcqct qqaqqatttc cacttqqatq aqqaqaqac cqtqaaqqtq cccatqatqt 780 cagaccetaa ggccgtttta cgctacgget tggattetga tetcaactge aagattgeec 840 agetgeeett gaeeggaage atgagtatea tettetteet geetetgaaa gtgaeeeaga 900 acctgaccat gatagaagag agcctcacct ctgagttcat tcacgacata gaccgagaac 960 tgaagacggt tcaagcggtc ctgaccgtcc ccaagctgaa gctgagttac gaaggcgaac 1020 tcacgaagtc tgtgcaggaa ctgaagctgc aatccttgtt tgattcacca gactttagca 1080 agatcacggg caaacctatc aaacttactc aagtggaaca tcgcattggc tttgagtgga 1140 acgaggatgg gggaagcgcc acctccagcc cagggccccg cctcaccttc cccctggact 1200 atcaccttaa ccagcctttc atctttgtac tgagggacac agacacagga gcccttctct 1260 tcataggcaa gattctggac cccaggagca cttaatgctc tagtttaatg ttcaaatatc 1320 ccagaagaag aaaactctag acagatggca gattatatat tacacgaaag ctgcacatat 1380 gtttcaatgt atactttgca ataaaagtgc tttatccc 1418 <210> SEQ ID NO 46 <211> LENGTH: 413 <212> TYPE: PRT <213> ORGANISM: Sus scrofa <400> SEQUENCE: 46 Met Gln Ala Leu Val Leu Leu Trp Thr Gly Ala Leu Leu Gly Ser Gly Ser Cys Gln Asn Ala Gly Pro Glu Glu Gly Ser Pro Ala Pro Asp 25 Thr Val Gly Ala Pro Val Glu Glu Glu Asp Pro Phe Phe Lys Val Pro Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu Tyr 55 Arg Val Arg Ser Ser Glu Ser Pro Thr Ala Asn Val Leu Leu Ser Pro

Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln

_																
Arg	Thr	Glu	Ser 100	Ser	Leu	His	Arg	Ala 105	Leu	Tyr	Tyr	Asp	Leu 110	Ile	Ser	
Asn	Pro	Asp 115	Leu	His	Gly	Thr	Tyr 120	Lys	Glu	Leu	Leu	Ala 125	Ala	Val	Thr	
Ala	Pro 130	Gln	Lys	Asn	Leu	Lys 135	Ser	Ala	Ser	Arg	Ile 140	Ile	Phe	Glu	Lys	
Lys 145	Leu	Arg	Ile	Lys	Ala 150	Ser	Phe	Val	Ala	Pro 155	Leu	Glu	Lys	Ser	Tyr 160	
Gly	Thr	Arg	Pro	Arg 165	Ile	Leu	Thr	Gly	Asn 170	Ser	Arg	Leu	Asp	Leu 175	Gln	
Glu	Val	Asn	Asn 180	Trp	Val	Gln	Ala	Gln 185	Thr	ГЛа	Gly	Lys	Val 190	Ala	Arg	
Ser	Thr	Arg 195	Glu	Leu	Pro	Gly	Glu 200	Ile	Ser	Ile	Leu	Leu 205	Leu	Gly	Val	
Ala	Tyr 210	Phe	Lys	Gly	Gln	Trp 215	Val	Thr	Lys	Phe	Asp 220	Ser	Arg	Lys	Thr	
Ser 225	Leu	Glu	Asp	Phe	His 230	Leu	Asp	Glu	Glu	Arg 235	Thr	Val	Lys	Val	Pro 240	
Met	Met	Ser	Asp	Pro 245	Lys	Ala	Val	Leu	Arg 250	Tyr	Gly	Leu	Asp	Ser 255	Asp	
Leu	Asn	CAa	Lys 260	Ile	Ala	Gln	Leu	Pro 265	Leu	Thr	Gly	Ser	Met 270	Ser	Ile	
Ile	Phe	Phe 275	Leu	Pro	Leu	Lys	Val 280	Thr	Gln	Asn	Leu	Thr 285	Met	Ile	Glu	
Glu	Ser 290	Leu	Thr	Ser	Glu	Phe 295	Ile	His	Asp	Ile	Asp 300	Arg	Glu	Leu	Lys	
Thr 305	Val	Gln	Ala	Val	Leu 310	Thr	Val	Pro	Lys	Leu 315	rya	Leu	Ser	Tyr	Glu 320	
Gly	Glu	Leu	Thr	Lув 325	Ser	Val	Gln	Glu	Leu 330	Lys	Leu	Gln	Ser	Leu 335	Phe	
Asp	Ser	Pro	Asp 340	Phe	Ser	ГÀа	Ile	Thr 345	Gly	Lys	Pro	Ile	350	Leu	Thr	
Gln	Val	Glu 355	His	Arg	Ile	Gly	Phe 360	Glu	Trp	Asn	Glu	Asp 365	Gly	Gly	Ser	
Ala	Thr 370	Ser	Ser	Pro	Gly	Pro 375	Arg	Leu	Thr	Phe	Pro 380	Leu	Asp	Tyr	His	
Leu 385	Asn	Gln	Pro	Phe	Ile 390	Phe	Val	Leu	Arg	Asp 395	Thr	Asp	Thr	Gly	Ala 400	
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_		_								_					ggcccc	60
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															gtggtg	180 240
															ccaac	300
															cggacg	
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cgactgacct	tccccctgga	ctaccaccto	aaccagcctt	tcatctttgt cttgcgggac	1260
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Ala Thr Gly	Thr Ala V	al Val Glu 40	Glu Glu Asp	Pro Phe Phe Lys Val 45	
Pro Val Asr 50	ı Lys Leu A	la Ala Ala 55	Val Ser Asn	Phe Gly Tyr Asp Leu 60	
Tyr Arg Glr		er Ser Ser	Pro Thr Thr	Asn Val Leu Leu Ser 80	
Pro Leu Sei	. Val Ala T 85	hr Ala Leu	Ser Ser Leu 90	Ser Leu Gly Ala Gly 95	
Pro Arg Thi	Glu Ser L	eu Ile His	Arg Ala Leu 105	Tyr Tyr Asp Leu Ile 110	
His Asn Pro	_	is Gly Thr	Tyr Lys Glu	Leu Leu Ala Thr Val 125	
Thr Ala Pro	Gln Lys A	sn Leu Lys 135	Thr Ala Ser	Arg Leu Val Leu Glu 140	
	_		Phe Val Gly	Leu Leu Glu Lys Ser	
	Arg Pro L		Thr Gly Asn	Thr Arg Thr Asp Leu	
	165		170	175	
		-		Lys Gly Lys Met Gly	

Arg Thr Leu Lys Glu Leu Pro Ser Gly Ile Ser Val Leu Leu Gly 195 200 205

Ile Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Pro Lys Lys 210 215 220	
Thr Ser Leu Gln Asp Phe His Leu Asp Glu Asp Arg Thr Val Lys Val 225 230 235 240	
Pro Met Met Ser Asp Pro Lys Ala Ile Ile Arg Tyr Gly Leu Asp Ser 245 250 255	
Asp Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Glu Gly Ser Met Ser 260 265 270	
Val Ile Phe Phe Leu Pro Leu Lys Ala Thr Gln Asn Leu Thr Leu Ile 275 280 285	
Glu Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu 290 295 300	
Lys Thr Ile Gln Ala Val Leu Thr Val Pro Lys Leu Gln Leu Ser Phe 305 310 315 320	
Glu Gly Glu Val Ser Lys Thr Phe Gln Glu Ile Lys Leu Gln Ser Leu 325 330 335	
Phe Asn Ser Pro Asp Leu Ser Lys Ile Thr Pro Arg Pro Ile Lys Leu 340 345 350	
Thr His Val Val His Arg Ser Ser Leu Glu Trp Ser Glu Asp Gly Val 355 360 365	
Gly Asp Ala Pro Ser Pro Ala Leu Leu Pro Ala Arg Leu Thr Phe Pro 370 375 380	
Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr 385 390 395 400	
Asp Thr Gly Thr Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly 405 410 415	
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cetetggace ggagecetee tggggeacag eagetgeeag aaegatgegg geggeeeeea	180
aggactetee ageteeegae gegacagggg tgeeegtgga ggaggaggae eeettettea	240
gggtccccgt gaataagctg gcagcagcca tctccaactt cggctatgac ctgtaccgtg	300
taaggtccag cttcagccct gctgccaatg tgctgctgtc accactcagc gtggccaccg	360
cactetetge getetegetg ggageggaac ageggacaga atceaceatt cacegggete	420
totactacga cotgatoago aaccoggaca tocacagoac otataaggag otoottgoot	480 540
ctgtcactgc cccggagaag aacttcaaga gtgcttcccg gattgtcttt gagaggaagc	600
tgcggataaa atccagcttt gttgcaccac tggagaagtc ctatagcacc aggcccagaa	660
tectgacegg caaceetege etggacette aggaggttaa caactgggtg caggeceaga	720
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	78∩
togaggattt coacttogat gaggagagga ctotgaaagt coccatgato toagaggeta	780 840
tcgaggattt ccacttggat gaggagagga ctgtgaaagt ccccatgatg tcagacccta aggccatctt acgctatggc ttggactctg atctcagctg taagattgcc cagctgccct	780 840 900

	060
tgaccggcag catgagtatc atctttttcc tgcctctgaa agtaacccag aacttgacca	960
tgatagaaga gagcetcace tetgagttea tteatgaeat agacegagag etgaagaeaa	1020
ttcaagcagt cctgaccatc cccaagctga agctgagtta tgaaggcgaa gtcacgaagt ccctgcagga aatgaaactg caatccttgt ttgattcacc agacttcagc aagatcacag	1080
	1140
gcaaacctat taaacttacc caagtggaac atcgagctgg cttcgagtgg aacgaggatg	1200
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atcacctgaa ccgacctttc atctttgtgc tgagagacac agacacaggg gcccttctct	1320
tcataggcaa aatcetggac cecaggggca tttaatgete eggtttttaa tgttecaata	1380
ccctagaaga acaaaaccct caacggatgg cagatgacat attacatgaa ggctgcccct	1484
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<400> SEQUENCE: 50	
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Ala Ala Ala Ile Ser Asn Phe Gly Tyr Asp Leu Tyr Arg Val Arg Ser 35 40 45	
Ser Phe Ser Pro Ala Ala Asn Val Leu Leu Ser Pro Leu Ser Val Ala 50 55 60	
Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln Arg Thr Glu Ser 65 70 75 80	
Thr Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile Ser Asn Pro Asp Ile 85 90 95	
His Ser Thr Tyr Lys Glu Leu Leu Ala Ser Val Thr Ala Pro Glu Lys 100 105 110	
Asn Phe Lys Ser Ala Ser Arg Ile Val Phe Glu Arg Lys Leu Arg Ile 115 120 125	
Lys Ser Ser Phe Val Ala Pro Leu Glu Lys Ser Tyr Ser Thr Arg Pro 130 135 140	
Arg Ile Leu Thr Gly Asn Pro Arg Leu Asp Leu Gln Glu Val Asn Asn 145 150 155 160	
Trp Val Gln Ala Gln Met Lys Gly Lys Ile Ala Arg Ser Thr Arg Glu 165 170 175	
Ile Pro Ser Gly Ile Ser Ile Leu Leu Leu Gly Val Ala Tyr Phe Lys 180 185 190	
Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr Ser Leu Glu Asp 195 200 205	
Phe His Leu Asp Glu Glu Arg Thr Val Lys Val Pro Met Met Ser Asp 210 215 220	
Pro Lys Ala Ile Leu Arg Tyr Gly Leu Asp Ser Asp Leu Ser Cys Lys 225 230 235 240	
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<210> SEQ ID NO 51

<211> LENGTH: 1579 <212> TYPE: DNA

<213 > ORGANISM: Macaca fascicularis

<400> SEQUENCE: 51

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<400> SEQUENCE	: 52				
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Pro Asp Ser Th 35	r Gly Ala Le	u Val Glu Gl 40	u Glu Asp Pr 45	o Phe Phe Lys	
Val Pro Val As 50	n Lys Leu Al 55		l Ser Asn Ph 60	e Gly Tyr Asp	
Leu Tyr Arg Va 65	l Arg Ser Se 70	r Met Ser Pro	o Thr Thr As 75	n Val Leu Leu 80	
Ser Pro Leu Se	r Val Ala Th 85	r Ala Leu Se 90	r Ala Leu Se	r Leu Gly Ala 95	
Glu Gln Arg Th 10		l Ile His Arg 105	g Ala Leu Ty	r Tyr Asp Leu 110	
Ile Ser Ser Pr 115	o Asp Ile Hi	s Gly Thr Ty: 120	r Lys Glu Le 12	-	
Val Thr Ala Pr 130	o Gln Lys As 13	-	r Ala Ser Ar 140	g Ile Val Phe	
Glu Lys Lys Le 145	u Arg Ile Ly 150	s Ser Ser Ph	e Val Ala Pr 155	o Leu Glu Lys 160	
Ser Tyr Gly Th	r Arg Pro Ar 165	g Val Leu Th: 17	_	o Arg Leu Asp 175	
Leu Gln Glu Il 18		p Val Gln Al. 185	a Gln Met Ly	s Gly Lys Leu 190	
Ala Arg Ser Th 195	r Lys Glu Le	u Pro Asp Gl	u Ile Ser Il 20		
Gly Val Ala Ty 210	r Phe Lys Gl 21		l Thr Lys Ph 220	e Asp Pro Arg	
Lys Thr Ser Le 225	u Glu Asp Ph 230	e His Leu As _]	p Glu Glu Ar 235	g Thr Val Arg 240	
Val Pro Met Me	t Ser Asp Pr 245	o Lys Ala Il 25		r Gly Leu Asp 255	
Ser Asp Leu Se 26		e Ala Gln Le 265	u Pro Leu Th	r Gly Ser Met 270	
Ser Ile Ile Ph 275	e Phe Leu Pr	o Leu Lys Va 280	l Thr Gln As 28		
Ile Glu Glu Se 290	r Leu Thr Se 29		e His Asp Il 300	e Asp Arg Glu	
Leu Lys Thr Va 305	l Gln Ala Va 310	l Leu Thr Le	u Pro Lys Le 315	u Lys Leu Ser 320	
Tyr Glu Gly Gl	u Val Thr Ly 325	s Ser Leu Gl:	_	s Leu Gln Ser 335	

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<211> LENGTH: 193 <212> TYPE: DNA

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Asp Phe Gly Tyr Asn Leu Phe Arg Ser Leu Ala Ser Arg Asp Thr Thr

Thr Asn Val Phe Leu Ala Pro Ile Ser Val Ser Ala Ala Leu Thr Gln 70

Leu Ser Met Gly Gly Ser Glu Leu Ala Glu Arg Gln Leu Phe Arg Ala

Leu Arg Phe His Thr Leu Gln Asp Pro Gln Leu His Asn Thr Leu Lys $100 \hspace{1.5cm} 100 \hspace{1.5cm} 105 \hspace{1.5cm} 110 \hspace{1.5cm}$

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We claim:

- 1. A method for identifying candidate compounds to treat and/or limit development of age-related macular degeneration (AMD), comprising contacting a first population of cells expressing OA1 with a test compound, and identifying as positive test compounds those test compounds that increase one or both of
 - (i) pigment epithelium-derived factor (PEDF) expression 45 in the first cell population relative to one or both (A) PEDF expression in the first population of cells not contacted with the test compound, and (B) a second cell population not expressing OA1, and
 - population relative to one or both (A) intracellular calcium concentration in the first population of cells not contacted with the test compound, and (B) the second cell population not expressing OA1;
 - wherein the positive test compounds are candidate com- 55 in the presence of between 0 uM and 10 uM tyrosine. pounds for treating and/or limiting development of
- 2. The method of claim 1 wherein the identifying comprises (i) detecting pigment epithelium-derived factor (PEDF) expression in the first cell population relative to one 60 or both (a) PEDF expression in the first population of cells not contacted with the test compound, and (b) the second cell population, and (ii) identifying as positive test compounds those test compounds that increase PEDF expression in the first cell population relative to one or both (a) PEDF expression in the first population of cells not contacted with the test compound, and (b) the second cell population.

- 3. The method of claim 1 wherein the identifying comprises (i) detecting levels of intracellular calcium concentration in the first cell population relative to one or both (a) intracellular calcium concentration in the first population of cells not contacted with the test compound, and (b) the second cell population; and (ii) identifying as positive test compounds those test compounds that increase intracellular calcium concentration in the first cell population relative to one or both (a) intracellular calcium concentration in the first population of cells not contacted with the test compound, and (b) the second cell population.
- 4. The method of claim 1, wherein the first cell population and the second cell population are selected from the group (ii) intracellular calcium concentration in the first cell 50 consisting of mouse, rat, hamster, and human cells.
 - 5. The method of claim 1, wherein the first cell population and the second cell population are retinal pigment epithelial
 - 6. The method of claim 1, wherein the contacting occurs
 - 7. The method of claim 1, wherein the contacting occurs in the presence of a tyrosinase inhibitor.
 - 8. The method of claim 2, wherein the first cell population and the second cell population are selected from the group consisting of mouse, rat, hamster, and human cells.
 - 9. The method of claim 2, wherein the first cell population and the second cell population are retinal pigment epithelial cells.
 - 10. The method of claim 2, wherein the contacting occurs in the presence of between 0 uM and 10 uM tyrosine.
 - 11. The method of claim 2, wherein the contacting occurs in the presence of a tyrosinase inhibitor.

- 12. The method of claim 3, wherein the first cell population and the second cell population are selected from the group consisting of mouse, rat, hamster, and human cells.
- 13. The method of claim 3, wherein the first cell population and the second cell population are retinal pigment 5 epithelial cells.
- 14. The method of claim 3, wherein the contacting occurs in the presence of between 0 uM and 10 uM tyrosine.
- 15. The method of claim 3, wherein the contacting occurs in the presence of a tyrosinase inhibitor.
- **16**. The method of claim **5**, wherein the contacting occurs in the presence of between 0 uM and 10 uM tyrosine.
- 17. The method of claim 5, wherein the contacting occurs in the presence of a tyrosinase inhibitor.
- 18. The method of claim 9, wherein the contacting occurs 15 in the presence of between 0 uM and 10 uM tyrosine.
- 19. The method of claim 9, wherein the contacting occurs in the presence of a tyrosinase inhibitor.
- **20**. The method of claim **13**, wherein the contacting occurs in the presence of between 0 uM and 10 uM tyrosine, 20 and/or wherein the contacting occurs in the presence of a tyrosinase inhibitor.

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