



(19)



EP 2 273 988 B1

(11)

(12)

EUROPEAN PATENT SPECIFICATION

(45) Date of publication and mention
of the grant of the patent:
09.09.2015 Bulletin 2015/37

(21) Application number: **09733334.8**

(22) Date of filing: **17.04.2009**

(51) Int Cl.:
A61K 31/00 (2006.01)

(86) International application number:
PCT/US2009/041021

(87) International publication number:
WO 2009/129497 (22.10.2009 Gazette 2009/43)

(54) L-DOPA FOR TREATING AGE-RELATED MACULAR DEGENERATION

L-DOPA ZUR BEHANDLUNG DER ALTERBSEDINGTEN MAKULADEGENERATION

L-DOPA POUR LE TRAITEMENT DE LA DÉGÉNÉRESCENCE MACULAIRE LIÉE A L AGE

(84) Designated Contracting States:
**AT BE BG CH CY CZ DE DK EE ES FI FR GB GR
HR HU IE IS IT LI LT LU LV MC MK MT NL NO PL
PT RO SE SI SK TR**

(30) Priority: **18.04.2008 US 124624 P**

(43) Date of publication of application:
19.01.2011 Bulletin 2011/03

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- **LOPEZ VANESSA M ET AL: "L-DOPA is an endogenous ligand for OA1" PLOS BIOLOGY, vol. 6, no. 9, September 2008 (2008-09), pages 1861-1869, XP002548207 ISSN: 1544-9173(print) 1545-7885(ele)**
- **Ulrich Schraermeyer ET AL: "Current Understanding on the Role of Retinal Pigment Epithelium and its Pigmentation", Pigment Cell Research, vol. 12, no. 4, 1 August 1999 (1999-08-01) , pages 219-236, XP55109059, ISSN: 0893-5785, DOI: 10.1111/j. 1600-0749.1999.tb00755.x**

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Description**Background**

- 5 [0001] Age-related macular degeneration ("AMD") is an aging-associated disease resulting in the loss of vision in the macula (the center of the visual field) because of damage to the retina. AMD is a prevalent disorder of the aged, with approximately 10% of patients 66 to 74 years and 30% of patients 75 to 85 years of age having some level of macular degeneration. Currently there is no effective treatment available for most patients with AMD, and no early stage intervention.
- 10 [0002] International patent application WO 03/070269 relates to 5,6-dihydroxyindole (DHI), 5,6-dihydroxyindole-2-carboxylic acid (DHICA) and/or 5-S-cysteinylldopa (CD) as medicaments, as well as their use, and the use of tyrosinase for the preparation of a medicament for prophylaxis or therapy of diseases induced by oxidative stress.

Summary of the Invention

- 15 [0003] The present invention provides an agonist of the OA1 receptor for use in treating or preventing age-related macular degeneration (AMD), wherein the agonist of the OA1 receptor is L-DOPA.
- [0004] The L-DOPA may be provided as a composition comprising:
- 20 (a) an amount effective of L-DOPA for treating or limiting development of AMD; and
(b) an amount effective for treating or limiting development of AMD of a composition comprising a source of vitamin C, a source of vitamin E, a source of vitamin A, a source of zinc, and a source of copper.

Brief Description of the Figures

- 25 [0005]
- Figure 1(a-c)** Western blot analysis of proteins bound (B) or unbound (U) to streptavidin-conjugated beads after biotinylation of RPE *in situ*, cultured RPE (b), or COS cells transfected to express OA1-GFP (c). Blots were probed to visualize OA1 and actin after cell surface biotinylation and fractionation using streptavidin-conjugated beads. For cultured cells (b, c) cells were either maintained in 500 μ M (normal DMEM) or 1 μ M tyrosine for 3 days prior to analysis.
- 30 **Figure 1(d)** Quantification of western blot analysis by densitometry. OA1 densitometry is shown as the % of the control for paired cell cultures, transfected then split into 2 equal groups, one of which was the control, maintained in normal DMEM (control). The other group was maintained in 1 μ M tyrosine DMEM (LT) until harvest. Paired t-test analysis was used to test whether the difference was significant, and * denotes p<0.001. Actin, analyzed the same way showed no differences, and p=0.724.
- Figure 1(e-f)** Composite confocal microscopy of pigmenting RPE cells maintained in normal DMEM (e) or 1 μ M tyrosine (f) then stained with anti-OA1 antibodies and imaged at 20x. Bar = 25 μ m.
- 35 **Figure 2(a)** Representative traces of $[Ca^{2+}]_i$ during the time course of the standard experimental protocol in transfected and untransfected CHO cells. After establishment of a stable baseline for 3 minutes, the test agent was added at 1 μ M. At 5 minutes, KCl was added to serve as a control that the cells were Fura-2 loaded and patent. Identical protocols were performed for both transfected cells and paired untransfected cells.
- Figure 2(b)** Summary data for $[Ca^{2+}]_i$ in response to tyrosine, dopamine, and L-DOPA in transfected and untransfected CHO cells. Untransfected cells are shown with L-DOPA treatment. The experimental control of membrane depolarization with KCl is also shown.
- 40 Each bar represents data collected from at least 10 experiments and is presented as the mean change from baseline $[Ca^{2+}]_i$ after test agent addition. Error bars represent S.D., and t-test analyses were used to test for significant differences, * denotes p<0.01.
- 45 Analysis of pertussis toxin sensitivity of $[Ca^{2+}]_i$ increase in cells transfected to express OA1 or RPE that express the natural protein. Data represent mean of at least 6 experiments.
- Figure 2(c)** Analysis of pertussis toxin sensitivity of $[Ca^{2+}]_i$ increase in cells transfected to express OA1 or RPE that express the natural protein. Data represent mean of at least 6 experiments for each group of transfected cells and 20 individual experiments for each the treated and untreated RPE with endogenous OA1 expression. T-tests analyses were used to test for significant differences, and * denotes p<0.01.
- 50 **Figure 2 (d)** cAMP was measured in CHO transfected to express OA1. The control group represents transfected but untreated CHO cells and the basal level of cAMP in those cells. Cells were treated with 1.0 μ M L-DOPA; 0.1 μ M forskolin, L-DOPA + 0.1 μ M forskolin, and as a positive control 1 μ M forskolin. Results represent the mean cAMP levels observed in at least 6 experiments in which all experimental groups were analyzed in a paired fashion

using replicate monolayers in the same culture plate. Error bars represent the S.D. of each group, and the only significant difference observed was the increase in cAMP levels after forskolin treatment.

Figure 3(a) Binding kinetics between OA1 and L-DOPA were determined using radiolabeled ligand binding assays. Results represent data collected from 5 such experiments and are presented as mean specific binding +/- SEM. The hyperbolic curve fit exhibited an R^2 value of 0.994, K_d was determined to be $9.34 \times 10^{-6} M$ +/- $1.14 \times 10^{-6} M$.

Figure 3(b) Comparative binding of $5 \mu M$ [H^3] L-DOPA to OA1 transfected CHO cells was compared in the presence of 1.0 mM dopamine, tyrosine, or L-DOPA. The data represent mean total binding +/- S.D. for each group. * denotes $p < 0.05$ when comparing the results between the control group to the binding in the presence of the potential competitive ligands.

Figure 3(c) Competitive interaction between $5 \mu M$ [H^3] L-DOPA and dopamine were assessed to determine whether dopamine functions as an antagonist of OA1 activity. Results indicate that dopamine and L-DOPA compete for the same OA1 binding site, and the data fits the binding model with an r^2 value of 0.95. The K_i for dopamine was $2.388 \pm 0.266 \mu M$ (mean +/- SEM), similar to the K_d for L-DOPA.

Figure 3(d) Dose-dependent OA1 signaling through OA1. Data represent mean increase in $[Ca^{2+}]_i$ elicited by L-DOPA treatment of the cells at the concentrations given ($n=6$ for each dose). T-test analyzes were used to compare between the responses achieved at each dose, and * denotes $p < 0.01$ for the comparison at 1 and $10 \mu M$.

Figure 3(e) Scatchard plot illustrating the kinetics of a single site binding relationship based on Figure 3(a).

Figure 4(a-h) All images represent $2 \mu m$ thick confocal sections of CHO cells transfected to express OA1-GFP. β -arrestin was visualized using immuno fluorescence methods.

Prior to addition of L-DOPA (a-c) and after treatment with $1 \mu M$ L-DOPA (d-f), and the merged images (c, f) illustrate regions where the two proteins co-localize, at the resolution of white light imaging. (g,h) are low magnification of field of transfected CHO cells, with two transfected cells visible (arrows) (g). The remainder of the cell population is visualized using antibodies to β -arrestin (h) to illustrate that β -arrestin recruitment to the membrane only occurred in the OA1 expressing cells (arrows).

Figure 5 (a) PEDF concentrations were determined by ELISA of cell conditioned medium. RPE cells were control cells, without L-DOPA treatment, or OA1 stimulated cells that were treated with $1 \mu M$ L-DOPA prior to being maintained for 3 days in normal DMEM. Data are presented as the mean of 3 experiments conducted in triplicate, error bars represent S.D., and * denotes $P < 0.01$ using a paired t-test.

Figure 5(b) PEDF concentrations in conditioned medium from pigmenting RPE determined by ELISA. Cells were either control pigmenting RPE cultures or paired cultures treated with phenylthiourea (PTU) at $200 \mu M$. Data are presented as the mean of 3 experiments conducted in triplicate, error bars represent S.D., and * denotes $P < 0.01$ using a paired t-test.

Figure 5(c) PEDF concentrations in conditioned medium of pigmented RPE cells treated with PTU then treated with L-DOPA to stimulate OA1 signaling. ELISA assays were conducted prior to PTU treatment, then after PTU treatment, and then from the same cultures after L-DOPA stimulation. Results are presented as mean +/- S.D. of the value achieved related to that culture of cells. * denotes $p < 0.01$ when comparing PTU to the control (same culture tested prior to PTU), and L-DOPA/PTU compared to the PTU sample from that same culture.

Figure 6(a) Data represents mean +/- SEM bound [3H]-L-DOPA in all fractions, total, specific and non-specific. Non-specific binding was determined by measuring radiolabeled-L-DOPA bound in the presence of excess unlabeled L-DOPA (1 mM). Specific binding at each given concentration is determined by subtracting the measured non-specific binding from the measured total binding.

Figure 6(b) The figure illustrates competitive interaction between tyrosine and L-DOPA, measured using increasing concentrations of tyrosine and $5 \mu M$ [H^3] L-DOPA. Each data point represents the mean data from 5 replicate wells, and the error bars are S.D. Data illustrate that tyrosine competes for binding with L-DOPA, but with a low affinity. The results suggest tyrosine has a K_i of $52.9 \mu M$, and fits the single site binding model with an r^2 value of 0.85. Saturation could not be achieved because of the limited solubility of tyrosine.

Figure 7 Western blot and graphical representation of PEDF secretion in wild-type vs OA deficient mice.

Figure 8(a) is a graphical representation of data demonstrating that L-DOPA supplementation increases retinal ganglion cell numbers compared to what is expected in a normal wild-type mouse.

Figure 8(b) is a graphical representation of data demonstrating that L-DOPA supplementation increases photoreceptor numbers compared to what is expected in a normal wild-type mouse.

Figure 8(c) is a Western blot showing PEDF detection in 2 wild-type and 2 OA1 -/y mice.

Detailed Description of the Invention

[0006] Within this application, unless otherwise stated, the techniques utilized may be found in any of several well-known references such as: Molecular Cloning: A Laboratory Manual (Sambrook, et al., 1989, Cold Spring Harbor Laboratory Press), Gene Expression Technology (Methods in Enzymology, Vol. 185, edited by D. Goeddel, 1991. Academic

Press, San Diego, CA), "Guide to Protein Purification" in Methods in Enzymology (M.P. Deutshcer, ed., (1990) Academic Press, Inc.); PCR Protocols: A Guide to Methods and Applications (Innis, et al. 1990. Academic Press, San Diego, CA), Culture of Animal Cells: A Manual of Basic Technique, 2nd Ed. (R.I. Freshney. 1987. Liss, Inc. New York, NY), Gene Transfer and Expression Protocols, pp. 109-128, ed. E.J. Murray, The Humana Press Inc., Clifton, N.J.), and the Ambion 1998 Catalog (Ambion, Austin, TX).

[0007] As used herein, the singular forms "a", "an" and "the" include plural referents unless the context clearly dictates otherwise.

[0008] The present invention provides an agonist of the OA1 receptor for use in treating or preventing age-related macular degeneration (AMD), wherein the agonist of the OA1 receptor is L-DOPA.

[0009] The human *Oa1* gene, is found on the X chromosome, and has been shown to encode a 404 amino acid protein OA1 (**SEQ ID NO:2**), likely to be a G-protein coupled receptor (GPCR) [12,13] based upon sequence analysis [14]. As disclosed in detail herein, the inventors have identified the OA1 signaling pathway as a critical determinant of neuro-sensory retina survival, such that stimulation of this pathway will provide treatment for AMD as well as a means to limit AMD development for those at potential risk. While not being bound by any mechanism, the inventors believe that OA1 and tyrosinase participate in an autocrine loop through L-DOPA that regulates the secretion of at least one potent neurotrophic factor, PEDF. Thus administration of L-DOPA can be used to stimulate OA1 activity and thus upregulate PEDF expression, making it a valuable therapeutic to treat and limit development of AMD.

[0010] The subject preferably is a human.

[0011] As used herein for all aspects and embodiments of the invention, "AMD" means an aging-associated disease resulting in the loss of vision in the macula (the center of the visual field) because of damage to the retina known as Age-related Macular Degeneration. As used herein, AMD encompasses both wet and dry AMD, described in more detail below.

[0012] AMD begins with characteristic drusen (yellow deposits) in the macula between the retinal pigment epithelium and the underlying choroid. Most people with these early changes (referred to as age-related maculopathy) have good vision. People with drusen can go on to develop advanced AMD. The risk is considerably higher when the drusen are large and numerous and associated with disturbance in the pigmented cell layer under the macula.

[0013] Subjects with age-related maculopathy may progress to either of the two main forms of advanced AMD, each of which can be treated or be limited in its development using the methods of the invention. "Wet" AMD causes vision loss due to abnormal blood vessel growth in the choriocapillaries, through Bruch's membrane, ultimately leading to blood and protein leakage below the macula. Bleeding, leaking, and scarring from these blood vessels eventually causes irreversible damage to the photoreceptors and rapid vision loss if left untreated. "Dry" AMD occurs when light-sensitive cells in the macula slowly break down, gradually causing vision loss in the affected eye. Blurring in AMD is probably due to the accumulation of drusen under the retinal pigment epithelium (RPE) which alters to focal properties of the photoreceptors by moving them out of the plane of focus.

[0014] Dry AMD may occur in one or both eyes, and can advance from age-related maculopathy into intermediate or advanced stages of dry AMD.

[0015] Intermediate Dry AMD: Either many medium-sized drusen or one or more large drusen. Some people see a blurred spot in the center of their vision. More light may be needed for reading and other tasks.

[0016] Advanced Dry AMD: In addition to drusen, a breakdown of light-sensitive cells and supporting tissue in the central retinal area. This breakdown can cause a blurred spot in the center of vision. Over time, the blurred spot may get bigger and darker, taking more of the central vision; may have difficulty reading or recognizing faces until they are very close to you.

[0017] AMD symptoms include, but are not limited to blurred/reduced central vision, central scotomas (shadows or missing areas of vision), trouble discerning one dark color from another dark color and/or one light color from another light color; slow recovery of visual function after exposure to bright light, a loss in contrast sensitivity, so that contours, shadows and color vision are less vivid, retinal pigment epithelial (RPE) disturbance (including pigment clumping and/or dropout), RPE detachment, geographic atrophy, subretinal neovascularization, and disciform scar, and distorted vision (metamorphopsia), such that a grid of straight lines appears wavy and parts of the grid may appear blank. Symptoms of dry AMD and wet AMD are generally similar early during disease-progression, and thus it may not be possible to determine which early-stage patients will develop dry vs. wet forms of AMD. Dry AMD develops as 'geographic atrophy', and early AMD become 'wet' AMD when new blood vessels sprout.

[0018] As used herein, "treat" or "treating" AMD means accomplishing one or more of the following: (a) reducing the severity of AMD; (b) limiting or preventing development of one or more symptoms characteristic of AMD, as described above; (c) inhibiting worsening of one or more symptoms characteristic of AMD, as described above; (d) limiting or preventing recurrence of AMD in patients that have previously had the disorder(s); and (e) limiting or preventing recurrence of one or more symptoms in patients that were previously symptomatic for AMD. Such treating includes treating of wet AMD and dry AMD.

[0019] As used herein, the term "limiting development of" AMD means to prevent or to minimize development of AMD in individuals at risk of developing AMD, as well as limiting progression of age-related maculopathy to AMD (wet or dry),

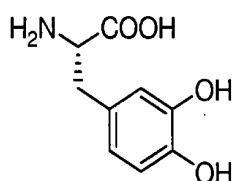
or intermediate dry AMD to advanced dry or 'wet' AMD. In one preferred embodiment, the treatment comprises treating a subject with drusen accumulation (ie: age-related maculopathy), to limit development of AMD. In another preferred embodiment, the treatment comprises treating a subject with an amount effective of the OA1 agonist L-DOPA to decrease the rate of lines of loss of vision relative to a non-treated AMD subject, or subject at risk of AMD. In another preferred embodiment, the treatment comprises treating a subject with wet AMD, or at risk of developing wet AMD, an amount effective of the OA1 agonist L-DOPA to decrease the rate and number of new blood vessel formation. As discussed in more detail below, OA1 stimulation causes the RPE to increase PEDF secretion, and PEDF is a potent anti-angiogenic factor. Thus, OA1 stimulation strategies may stop new blood vessel development in 'wet' AMD, in addition to its effects on retinal development discussed herein.

[0020] In another preferred embodiment, the treatment comprises treating a subject that has blurred or reduced central vision with an amount of the OA1 agonist L-DOPA effective to increase the lines of visual acuity in one or both eyes. In this embodiment, the lines of visual acuity are as measured by the standard Snellen test, where the increase or decrease in 'lines' of visual acuity are based on which smallest 'line' on a Snellen chart a patient can read clearly.

[0021] "Subjects at risk of developing AMD" means anyone with any risk factor for development of AMD, including but not limited to being over 50 years old (in various preferred embodiments, over 60 years old, over 65 years old, over 70 years old, or over 75 years-old), presence of drusen deposits, Caucasian race, having a blood relative that has or had AMD, a mutation in the complement factor H gene (CFH) of (Tyr402His), Arg80Gly variant of the complement protein C3 gene, hypertension, high cholesterol levels, obesity, smoking, a high fat intake, and mutations in the fibulin 5 gene. Thus, in a preferred embodiment, the subject to be treated has one or more of these risk factors, particularly in treatments for limiting development of AMD.

[0022] The phrase "therapeutically effective amount," as used herein, refers to an amount that is sufficient or effective to limit development of or treat (prevent the progression of or reverse) AMD. The appropriate dosage range depends on the choice of the compound, the route of administration, the nature of the formulation, the nature of the subject's condition, and the judgment of the attending practitioner. For example, oral administration would be expected to require higher dosages than administration by intravenous injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization, as is well understood in the art.

[0023] L-DOPA is [2-amino-3-(3,4-dihydroxyphenyl)propanoic acid] known for use in treating Parkinson's, and has the following structure.



[0024] L-DOPA is commercially available and methods for its synthesis are known to those of skill in the art.

[0025] While not being bound by a specific mechanism of action, the inventor believes that L-DOPA binding to OA1 involves two sites of binding, one involving one or both hydroxyl groups, and one involving the carboxylic acid group.

[0026] In one embodiment of the invention, the treatment or prevention may comprise administering a further therapeutic compound to the subject, including but not limited to an L-amino acid decarboxylase inhibitor, such as carbidopa or benserazide. Such L-amino acid decarboxylase inhibitors can be used, for example, to increase plasma half-life of L-DOPA and reduce conversion of L-DOPA to dopamine peripherally, which reduces side effects of L-DOPA treatment. In another embodiment, the treatment or prevention may further comprise administering one or more other compounds useful for treating or limiting development of AMD, including but not limited to anti-angiogenic therapeutics, such as anti-vascular endothelial growth factor (VEGF) agents, including but not limited to VEGF antibodies (or fragments thereof) such as ranibizumab or bevacizumab, or VEGF aptamers, such as pegaptanib. In another embodiment, the L-DOPA may be present in a more complex mixture, such as in a nutritional supplement containing L-DOPA.

[0027] In a preferred embodiment, the L-DOPA may be used in the form of a dietary supplement. Such a supplement may combine any one or more further components that might be beneficial in treating or limiting development of AMD. In one preferred embodiment, L-DOPA is combined with a combination of vitamin C source, vitamin E source, Vitamin A source, zinc source, and a copper source, disclosed in US Patent No. 6,660,297 as useful in treating AMD. Any suitable amount of each of these additional components can be used in combination with L-DOPA in carrying out the treatment or prevention described in relation to the invention. In a further preferred embodiment, this combination may further comprise lutein and/or zeaxanthin in an amount suitable to provide further protective retinal effects, preferably between 1 mg and 100 mg; between 1 mg and 50 mg, between 2 mg and 25 mg, or between 2 mg and 10 mg per day. In a further preferred embodiment of any of the above preferred embodiments, this combination may further comprise docosahexaenoic acid (DHA) and/or eicosapentaenoic acid (EPA) in an amount suitable to provide further protective retinal effects,

preferably between 250 mg and 1000 mg; between 300 mg and 750 mg, between 350 mg and 750 mg, or between 350 mg and 650 mg per day. The use of such compositions for treating AMD patients is discussed, for example, at web site www.areds2.org/ and links therein.

[0028] Ascorbic acid is the preferred source of vitamin C, although other sources such as for example sodium ascorbate could alternatively be used.

[0029] DL-alpha tocopheryl acetate is the preferred source of vitamin E, although other sources of vitamin E, such as for example trimethyl tocopheryl acetate and/or vitamin E succinate, may be used in the alternative.

[0030] Beta-carotene is preferred in the subject composition due to its ready commercial availability although alternative carotenoid proforms of vitamin A could likewise be used.

[0031] Zinc is preferred in the form of zinc oxide in subject tablets due to the fact zinc oxide provides the most concentrated form for elemental zinc and is well tolerated in the digestive system. However, other forms of zinc such as for example zinc gluconate may alternatively be used or be used in combination with zinc oxide in the subject composition.

[0032] Copper in the form of cupric oxide is preferred in the subject tablets to help prevent zinc induced copper deficiency anemia, although other forms of copper such as for example copper gluconate may alternatively be used or used in combination with cupric oxide in the subject composition.

[0033] In a preferred embodiment, the amounts of each of these other components (on a per day basis) is as follows:

between 450 mg and 600 mg vitamin C (approximately 7-10 times the recommended daily allowance (RDA));
between 400 IU and 540 IU vitamin E (approximately 13-18 times the RDA);

between 17.2 mg and 28 mg beta carotene (approximately 6-10 times the RDA of vitamin A; beta carotene is a prodrug of vitamin A);

between 68 mg and 100 mg zinc (approximately 4-7 times the RDA for zinc); and

between 1.6 mg and 2.4 mg copper.

[0034] In a further preferred embodiment, the amounts of each of these other components (on a per day basis) is as follows:

500 mg Vitamin C;

400 IU Vitamin E;

0 mg or 15 mg beta carotene;

25 mg or 80 mg zinc oxide; and

2 mg cupric oxide.

[0035] In a further preferred embodiment, that may be combined with any other embodiments herein, other ingredients believed to be of benefit in maintaining eye health may likewise be combined with L-DOPA, including but not limited to lutein and/or zeaxanthin in an amount suitable to provide further protective retinal effects, preferably between 1 mg and 100 mg; between 1 mg and 50 mg, between 2 mg and 25 mg, or between 2 mg and 10 mg per day; and/or docosahexaenoic acid (DHA) and/or eicosapentaenoic acid (EPA) in an amount suitable to provide further protective retinal effects, preferably between 250 mg and 1000 mg; between 300 mg and 750 mg, between 350 mg and 750 mg, or between 350 mg and 650 mg per day. Further examples of additional compounds that may optionally be used include but are not limited to alpha-lipoic acid and, phenolic compounds such as for example but not limited to oligomeric proanthocyanidins, anthocyanosides and combinations thereof.

[0036] L-DOPA can be administered individually or in combination, usually in the form of a pharmaceutical composition. Such compositions are prepared in a manner well known in the pharmaceutical art. L-DOPA can be administered as the sole active pharmaceutical agent, or it can be used in combination with one or more other compounds useful for carrying out the treatment or prevention described in relation to the invention, including but not limited to anti-angiogenic therapeutics such as VEG-F, and L-amino acid decarboxylase inhibitors, such as carbidopa and benserazide. When administered as a combination, combination can be formulated as separate compositions that are given at the same time or different times, or can be given as a single composition.

[0037] The L-DOPA may be made up in a solid form (including granules, powders or suppositories) or in a liquid form (e.g., solutions, suspensions, or emulsions). The L-DOPA may be applied in a variety of solutions and may be subjected to conventional pharmaceutical operations such as sterilization and/or may contain conventional adjuvants, such as preservatives, stabilizers, wetting agents, emulsifiers, buffers etc.

[0038] The L-DOPA may be administered by any suitable route, including but not limited to oral, topical (including but not limited to eye drops and ophthalmic ointments), parenteral, intranasal, pulmonary, or rectal in dosage unit formulations containing conventional non-toxic pharmaceutically acceptable carriers, adjuvants and vehicles. The term parenteral as used herein includes percutaneous, subcutaneous, intravascular (e.g., intravenous), intramuscular, or intrathecal injection or infusion techniques and the like. In addition, there is provided a pharmaceutical formulation comprising a compound

of the invention and a pharmaceutically acceptable carrier. L-DOPA may be present in association with one or more non-toxic pharmaceutically acceptable carriers and/or diluents and/or adjuvants, and if desired other active ingredients. The pharmaceutical compositions containing L-DOPA may be in a form suitable for oral use, for example, as tablets, troches, lozenges, aqueous or oily suspensions, dispersible powders or granules, emulsion, hard or soft capsules, or 5 syrups or elixirs.

[0039] Eye drops can be prepared using any technique in the art, including but not limited to using a tonicity agent such as sodium chloride or concentrated glycerin, a buffer such as sodium phosphate or sodium acetate, a surfactant such as polyoxyethylene sorbitan monooleate, polyoxyl 40 stearate or polyoxyethylene hydrogenated castor oil, a stabilizer such as sodium citrate or sodium edetate, a preservative such as benzalkonium chloride or paraben as needed.

10 The pH of the eye drops is preferably in the range of from 4 to 8. Ophthalmic ointments can be prepared with a generally used base such as white soft paraffin or liquid paraffin.

[0040] L-DOPA intended for oral use may be prepared according to any method known to the art for the manufacture of pharmaceutical compositions and such compositions may contain one or more agents selected from the group consisting of sweetening agents, flavoring agents, coloring agents and preservative agents in order to provide palatable 15 preparations. Tablets contain the L-DOPA in admixture with non-toxic pharmaceutically acceptable excipients that are suitable for the manufacture of tablets. These excipients may be for example, inert diluents, such as calcium carbonate, sodium carbonate, lactose, calcium phosphate or sodium phosphate; granulating and disintegrating agents, for example, corn starch, or alginic acid; binding agents, for example starch, gelatin or acacia, and lubricating agents, for example magnesium stearate, stearic acid or talc. The tablets may be uncoated or they may be coated by known techniques. In 20 some cases such coatings may be prepared by known techniques to delay disintegration and absorption in the gastrointestinal tract and thereby provide a sustained action over a longer period. For example, a time delay material such as glycercyl monostearate or glycercyl distearate may be employed.

[0041] Formulations for oral use may also be presented as hard gelatin capsules wherein the L-DOPA is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin, or as soft gelatin capsules wherein 25 the active ingredient is mixed with water or an oil medium, for example peanut oil, liquid paraffin or olive oil.

[0042] Aqueous suspensions contain the L-DOPA in admixture with excipients suitable for the manufacture of aqueous suspensions. Such excipients are suspending agents, for example sodium carboxymethylcellulose, methylcellulose, hydropropyl-methylcellulose, sodium alginate, polyvinylpyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents may be a naturally-occurring phosphatide, for example, lecithin, or condensation products of an alkylene 30 oxide with fatty acids, for example polyoxyethylene stearate, or condensation products of ethylene oxide with long chain aliphatic alcohols, for example heptadecaethylenoxyacetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with partial esters derived from fatty acids and hexitol anhydrides, for example polyethylene sorbitan 35 monooleate. The aqueous suspensions may also contain one or more preservatives, for example ethyl, or n-propyl p-hydroxybenzoate, one or more coloring agents, one or more flavoring agents, and one or more sweetening agents, such as sucrose or saccharin.

[0043] Oily suspensions may be formulated by suspending the L-DOPA in a vegetable oil, for example arachis oil, olive oil, sesame oil or coconut oil, or in a mineral oil such as liquid paraffin. The oily suspensions may contain a thickening 40 agent, for example beeswax, hard paraffin or cetyl alcohol. Sweetening agents and flavoring agents may be added to provide palatable oral preparations. These compositions may be preserved by the addition of an anti-oxidant such as ascorbic acid.

[0044] Dispersible powders and granules suitable for preparation of an aqueous suspension by the addition of water provide the active ingredient in admixture with a dispersing or wetting agent, suspending agent and one or more preservatives. Suitable dispersing or wetting agents or suspending agents are exemplified by those already mentioned 45 above. Additional excipients, for example sweetening, flavoring and coloring agents, may also be present.

[0045] Pharmaceutical compositions for use in the treatment or prevention described in relation to the invention may also be in the form of oil-in-water emulsions. The oily phase may be a vegetable oil or a mineral oil or mixtures of these. Suitable emulsifying agents may be naturally-occurring gums, for example gum acacia or gum tragacanth, naturally-occurring phosphatides, for example soy bean, lecithin, and esters or partial esters derived from fatty acids and hexitol, 50 anhydrides, for example sorbitan monooleate, and condensation products of the said partial esters with ethylene oxide, for example polyoxyethylene sorbitan monooleate. The emulsions may also contain sweetening and flavoring agents.

[0046] Syrups and elixirs may be formulated with sweetening agents, for example glycerol, propylene glycol, sorbitol, glucose or sucrose. Such formulations may also contain a demulcent, a preservative and flavoring and coloring agents. The pharmaceutical compositions may be in the form of a sterile injectable aqueous or oleaginous suspension. This 55 suspension may be formulated according to the known art using those suitable dispersing or wetting agents and suspending agents that have been mentioned above. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parentally acceptable diluent or solvent, for example as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution and isotonic sodium

chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including synthetic mono- or diglycerides. In addition, fatty acids such as oleic acid find use in the preparation of injectables.

[0047] Specific methods for intranasal administration of L-DOPA are known in the art; see, for example, Kao et al., 5 Pharmaceutical Research 17(8):978-984 (2000).

[0048] The dosage range depends on the choice of the compound, the route of administration, the nature of the formulation, the nature of the subject's condition, and the judgment of the attending practitioner. For example, oral administration would be expected to require higher dosages than administration by intravenous injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization, as is well understood in the art.

10 In certain embodiments, L-DOPA can be administered at dosages of between 10 mg/day and 1500 mg/day; in various preferred embodiments administration can be between 20 mg and 1200 mg/day, 50 mg and 1000 mg/day, 100 mg and 500 mg/day, and 200 mg and 400 mg/day.

15 **[0049]** Pharmaceutical compositions containing the compounds described herein are administered to an individual in need thereof. In a preferred embodiment, the subject is a mammal; in a more preferred embodiment, the subject is a human. In therapeutic applications, compositions are administered in an amount sufficient to carry out the treatment or prevention described in relation to the invention. Amounts effective for these uses depend on factors including, but not limited to, the nature of the compound (specific activity, etc.), the route of administration, the stage and severity of the disorder, the weight and general state of health of the subject, and the judgment of the prescribing physician. The active compound is effective over a wide dosage range. However, it will be understood that the amount of the compound actually administered will be determined by a physician, in the light of the above relevant circumstances. Therefore, the above dosage ranges are not intended to limit the scope of the invention in any way.

20 **[0050]** The L-DOPA for use in the present invention may be provided as a composition comprising:

- 25 (a) an amount effective of L-DOPA for treating or limiting development of AMD; and
- (b) an amount effective for treating or limiting development of AMD of a composition comprising a source of vitamin C, a source of vitamin E, a source of vitamin A, a source of zinc, and a source of copper.

30 **[0051]** The amount of L-DOPA in the compositions is suitable to provide for administration at dosages of between 10 mg/day and 1500 mg/day; in various preferred embodiments administration can be between 20 mg and 1200 mg/day, 50 mg and 1000 mg/day, 100 mg and 500 mg/day, and 200 mg and 400 mg/day.

35 **[0052]** Ascorbic acid is the preferred source of vitamin C in the subject tablets, although other sources such as for example sodium ascorbate could alternatively be used. Dl-alpha tocopheryl acetate is the preferred source of vitamin E in the subject tablets although other sources of vitamin E, such as for example trimethyl tocopheryl acetate and/or vitamin E succinate, may be used in the alternative. Beta-carotene is preferred in the subject composition due to its ready commercial availability although alternative carotenoid proforms of vitamin A could likewise be used. Zinc is preferred in the form of zinc oxide in subject tablets due to the fact zinc oxide provides the most concentrated form for elemental zinc and is well tolerated in the digestive system. However, other forms of zinc such as for example zinc gluconate may alternatively be used or be used in combination with zinc oxide in the subject composition. Copper in the form of cupric oxide is preferred in the subject tablets to help prevent zinc induced copper deficiency anemia, although other forms of copper such as for example copper gluconate may alternatively be used or used in combination with cupric oxide in the subject composition.

40 **[0053]** In one preferred embodiment, composition "b" provides a formulation suitable to permit ingestion of the following amounts of each component:

- 45 Ascorbic acid: at least 450 mg;
- dl-alpha tocopheryl acetate: 400 IU;
- beta carotene: 17.2 mg;
- zinc oxide: 68 mg; and
- cupric oxide: 1.6 mg.

50 **[0054]** In one preferred embodiment, composition "b" provides a formulation suitable to permit ingestion of the following amounts of each component:

- 55 500 mg Vitamin C;
- 400 IU Vitamin E;
- 0 mg or 15 mg beta carotene;
- 25 mg or 80 mg zinc oxide; and
- 2 mg cupric oxide.

[0055] The preferred daily dosage of the subject composition as specified above may be administered in the form of 1, 2, 3, 4, or more dosage forms according to any suitable route of administration as disclosed above. In preferred embodiments, the dosage form is an oral or topical dosage form, according to any embodiment of such dosage forms described herein. In another preferred embodiment the daily dosage of the subject composition is provided in the form of one dosage form taken twice daily, for a total of two dosage forms a day, or in the form of two dosage forms taken twice daily, for a total of four dosage forms a day. Compared to taking the total daily dose once a day, twice daily dosing of half the total daily dose in one or more dosage forms per dose provides improved absorption and better maintenance of blood levels of the essential ingredients. Accordingly, if two dosage forms of the preferred formulation of the subject composition are to be ingested each day, each dosage form is formulated to preferably provide not less than approximately 225 mg ascorbic acid, approximately 200 IU dl-alpha tocopheryl acetate, approximately 8.6 mg beta-carotene, approximately 34 mg zinc oxide and approximately 0.8 mg cupric oxide upon oral administration. If four tablets of the preferred formulation of the subject composition are to be ingested each day, each tablet is formulated to preferably provide not less than approximately 112.5 mg ascorbic acid, approximately 100 IU dl-alpha tocopheryl acetate, approximately 4.3 mg beta-carotene, approximately 17 mg zinc oxide, approximately 0.4 mg cupric oxide, and between 5 mg and 750 mg of L-DOPA.

[0056] In another preferred embodiment, the compositions comprise

- (a) between 5 mg and 1500 mg L-DOPA;
- (b) between 450 mg and 600 mg vitamin C (approximately 7-10 times the recommended daily allowance (RDA));
- (c) between 400 IU and 540 IU vitamin E (approximately 13-18 times the RDA);
- (d) between 17.2 mg and 28 mg beta carotene (approximately 6-10 times the RDA of vitamin A; beta carotene is a prodrug of vitamin A);
- (e) between 68 mg and 100 mg of zinc (approximately 4-7 times the RDA for zinc); and
- (f) at least 1.6 mg of copper.

[0057] In various preferred embodiments, the composition may comprise between 10 mg and 1200 mg; between 25 mg and 1000 mg; between 50 mg and 500 mg, or between 100 mg and 400 mg L-DOPA.

[0058] In a further preferred embodiment, that may be combined with any other embodiments herein, other ingredients believed to be of benefit in maintaining eye health may likewise be combined with L-DOPA, including but not limited to lutein and/or zeaxanthin in an amount suitable to provide further protective retinal effects, preferably between 1 mg and 100 mg; between 1 mg and 50 mg, between 2 mg and 25 mg, or between 2 mg and 10 mg per day; and/or docosahexaenoic acid (DHA) and/or eicosapentaenoic acid (EPA) in an amount suitable to provide further protective retinal effects, preferably between 250 mg and 1000 mg; between 300 mg and 750 mg, between 350 mg and 750 mg, or between 350 mg and 650 mg per day. The amounts necessary in any particular dosage form to provide the recited amounts can be determined by one of skill in the art based on the teachings herein and the number of dosage forms to be administered per day.

[0059] As described above, human OA1 (**SEQ ID NO:1-2 NP 000264.1**) is a G-protein coupled receptor and the inventors have herein identified L-DOPA as an OA1 ligand. As disclosed in more detail below, the inventor has discovered the existence of an autocrine loop between OA1 and tyrosinase linked through L-DOPA, and this loop includes the secretion of at least one very potent retinal neurotrophic factor (PEDF) as well as an increase in intracellular calcium concentration. OA1 is a selective L-DOPA receptor whose downstream effects govern spatial patterning of the developing retina. Thus, test compounds that selectively up-regulate PEDF expression and/or intracellular calcium concentration via stimulation of the OA1 pathway are candidate compounds for treating and/or limiting development of AMD. OA1 homologues include, but are not limited to:

- Mouse: SEQ ID NO:3-4 (NM_010951);
- Xenopus tropicalis: SEQ ID NOS:5-6 (NM_001011018);
- Cow: SEQ ID NOS:7-8 (XM_001506318);
- Rat: SEQ ID NOS: 9-10 (NM_001106958);
- Platypus: SEQ ID NOS: 11-12 (XM_001506318);
- Xenopus laevis: SEQ ID NOS: 13-14 (NM_001096842)
- Chicken: SEQ ID NOS:15-16 (XM_416848);
- Zebrafish: SEQ ID NOS: 17-18 (NM_200822);
- Chimpanzee: SEQ ID NO: 19 (XR_025625);
- Rhesus monkey: SEQ ID NOS:21-22 (XM_001090139; and
- Macaque: SEQ ID NO: 23 (BV209253).

[0060] PEDF is pigment epithelium-derived factor (Exp Eye Res 53: 411-414), and is a known neurotrophic factor with

the potential to alter neurosensory retina development, and to inhibit blood vessel growth. PEDF homologues include, but are not limited to:

5	Human:	SEQ ID NOS:25-26 (NM_002615);
	Rat:	SEQ ID NOS:27-28 (NM_031356);
	Zebra finch:	SEQ ID NOS: 29-30 (XM_002197419);
	Horse:	SEQ ID NOS:31-32 (NM_001143954);
	Xenopus tropicalis:	SEQ ID NOS:33-34 (NM_203755);
10	Mouse:	SEQ ID NOS:35-36 (NM_011340);
	Atlantic salmon:	SEQ ID NOS:37-38 (NM_001140334);
	Sheep:	SEQ ID NOS:39-40 (NM_001139447);
	Guinea pig:	SEQ ID NOS:41-42 (EF679792);
	Cow:	SEQ ID NOS:43-44 (NM_174140);
15	Wild boar:	SEQ ID NOS:45-46 (NM_001078662);
	Platypus:	SEQ ID NOS:47-48 (XM_001507128);
	Wolf:	SEQ ID NOS:49-50 (NM_001077588);
	Macaque:	SEQ ID NOS: 51-52 (AB174277);
20	Chimpanzee:	SEQ ID NOS: 53-54 (XM_001154665);

Rhesus monkey: SEQ ID NOS: 55-56 (XM_001117361); and
 Flounder: SEQ ID NOS: 57-58 (DQ115406).

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[0061] The inventor has determined that OA1 signaling can be used to rescue photoreceptor and ganglion cell development in tyrosinase-deficient animals, and in the process establish the neurotrophic effect of OA1 signaling. Thus, compounds that rescue neurosensory retinal development through OA1 signaling are good candidates for AMD treatment. Described herein is the first establishment of such an animal model for AMD drug screening.

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Examples: L-DOPA is an Endogenous Ligand for OA1

[0062] Background: Albinism is a genetic defect characterized by a loss of pigmentation. The neurosensory retina, which is not pigmented, exhibits pathologic changes secondary to the loss of pigmentation in the retina pigment epithelium (RPE). How the loss of pigmentation in the RPE causes developmental defects in the adjacent neurosensory retina has not been determined, but offers a unique opportunity to investigate the interactions between these two important tissues. One of the genes which causes albinism encodes for an orphan GPCR (OA1) expressed only in pigmented cells, including the RPE.

[0063] Methodology/Principle Findings: The function and signaling of OA1 was investigated in RPE and transfected cell lines. The results indicate that OA1 is a selective L-DOPA receptor, with no measurable second messenger activity from two closely related compounds, tyrosine and dopamine. Radiolabeled ligand binding confirmed that OA1 exhibited a single, saturable binding site for L-DOPA. Dopamine competed with L-DOPA for the single OA1 binding site suggesting it could function as an OA1 antagonist. OA1 response to L-DOPA was defined by several common measures of GPCR activation including influx of intracellular calcium and recruitment of β -arrestin. Further, inhibition of tyrosinase, the enzyme that makes L-DOPA, resulted in decreased PEDF secretion by RPE. Further, stimulation of OA1 in RPE with L-DOPA resulted in increased PEDF secretion.

[0064] Conclusions/Significance: Taken together the results illustrate an autocrine loop between OA1 and tyrosinase linked through L-DOPA, and this loop includes the secretion of at least one very potent retinal neurotrophic factor. OA1 is a selective L-DOPA receptor whose downstream effects govern spatial patterning of the developing retina. The results suggest that the retinal consequences of albinism caused by changes in melanin synthetic machinery may be treated by L-DOPA supplementation.

[0065] Introduction: Albinism is a group of inherited genetic diseases in which there is a variable loss of pigmentation in the eye, hair or skin. When the eye is affected, there are significant alterations in neurosensory retina development that lead to low vision [1-8]. There are two broad classes of albinism, ocular-cutaneous albinism (OCA) and ocular albinism (OA). OCA occurs when all pigmented tissues exhibit hypopigmentation and involves genetic mutations that result in defects in the melanin synthetic machinery [3,7-9]. OA occurs when cutaneous tissues pigment normally, but the ocular tissues are hypopigmented [10,11]. Since the same proteins produce pigment in all tissues, OA most likely

results from lack of expression of the melanogenic enzymes in ocular tissue rather than an inability to synthesize melanin because the other tissues pigment normally.

[0066] OA can be linked to at least one gene, *Oa1*, which is found on the X chromosome. *Oa1* encodes a 404 amino acid protein likely to be an orphan G-protein coupled receptor (GPCR), OA1 (Genbank GPR143) [12,13] based upon sequence analysis [14]. Schiaffino *et al.* has demonstrated that OA1 associates with several G_{α} subunits as well as G_{β} adding further evidence that OA1 is a GPCR [14,15]. Indeed, Innamorati *et. al.* used a combinatorial expression strategy to illustrate GPCR-like activity from OA1, as well as β -arrestin association, even in the absence of a ligand [16]. This work suggested that OA1 could signal through a $G_{\alpha}q$ subunit through phospholipase C and inositol triphosphate second messengers. In a yeast based expression system, Staleva and Orlow have demonstrated GPCR signaling from OA1 that appeared to be activated by a component in the melanosomal compartment [17]. Despite the significant amount of circumstantial evidence that OA1 is a GPCR, confirmation is lacking because no ligand has been identified. Other data has called into question the idea that OA1 is a GPCR. For example, the localization of OA1 as a fully intracellular protein is not typical of GPCRs and suggests that it would be a unique member of the family [14]. OA1 is primarily localized to the endolysosomal compartment [14,15,18-21] and melanosomes [11,14,22] rather than the cell surface.

[0067] In this study the function of OA1 as a potential GPCR was investigated, based on the hypothesis that the endosomal localization of OA1 in cultured cells was due to internalization of OA1 in response to an agent in the culture medium. Further, a ligand for OA1 was sought based on the observation that all forms OCA and OA appear to have the same retinal phenotype, indicating that tyrosinase activity and OA1 signaling are coupled upstream of retinal development. Thus, tests on whether tyrosinase activity produces the ligand for OA1 were carried out. A by-product of melanin synthesis is L-DOPA, which is released to the retina during melanin synthesis in the RPE at a critical time in retinal development [23,24]. The data suggest that OA1 is a highly selective L-DOPA receptor, and that L-DOPA causes OA1 signaling with the downstream effect of neurotrophic factor secretion by RPE. Thus, the first evidence is presented of a ligand for OA1, and provide a mechanism through which either tyrosinase or OA1 deficiency results in changes to retinal development.

25 Results:

Cell Surface Localization of OA1.

[0068] OA1 has previously been localized in pigment granules *in situ* [22], however, using transfected cells of various types, OA1 also has been localized to both the plasma membrane [16,17] and the endosomal fraction of cultured cells [14,16-18,20,21]. The investigation began by determining where OA1 resides in the human tissue using cell surface biotinylation/western blot strategies. In the human eye, OA1 was present on the apical cell surface of the RPE *in situ* (Fig. 1 A). Quantification of cell surface, biotinylated OA1 in five human eyes indicated that at least 3.5 +/- .7% of the total OA1 resided on the apical cell surface of RPE *in situ*. Access to the biotinylation reagent using eye cup preparations is restricted to the apical surface, so the polarity of OA1 in the epithelium cannot be determined. Further, the total cell surface OA1 is likely underestimated because of the lack of access to the basal cell surface. Blots were also probed with antibodies against actin as a control to verify that cytoplasmic proteins were not biotinylated. In each experiment actin was only found in the unbound fraction.

[0069] Others have reported that recombinant OA1 and OA1-GFP is almost exclusively localized to the endosomal compartment in cultured cells [14,15,17,18,20-22]. However, when overexpressed [16], or when endocytosis is inhibited [17], OA1 accumulates at the cell surface. The observation that OA1 protein is present on the apical surface of RPE *in situ* led us to explore the issue further.

Effects of Tyrosine on OA1 Expression and Distribution

[0070] Endosomal localization of GPCRs occurs normally after exposure to a ligand. Therefore, it was investigated whether a ligand for the receptor was present in the standard incubation medium that could drive internalization of OA1. Since the standard culture medium contains 500 μ M tyrosine, and tyrosine is the starting material for pigment synthesis, the effect of tyrosine on receptor distribution was evaluated. To test whether tyrosine affected OA1 distribution in cultured cells DMEM was formulated without tyrosine, and dialyzed fetal bovine serum was used. In the presence of tyrosine-free medium, OA1 was detected on the plasma membrane of cultured RPE cells both in the absence (not shown), and in medium containing low concentrations of tyrosine (1 μ M, Fig. 1 B). Averaged over five experiments, 4.5 +/- 1% of total OA1 protein was observed on the surface of cultured RPE maintained in 1 μ M tyrosine, similar to what was observed for RPE *in situ*. In all experiments actin was observed in the unbound protein fraction, demonstrating the absence of any cytoplasmic protein in the cell surface assay. Similarly, OA1-GFP expressed in COS illustrated a cell surface expression that was tyrosine sensitive (Fig. 1 C). Quantification of six such experiments indicated significant variability in the amount of OA1 found at the cell surface using transient transfections. The range of OA1 in the bound fraction of transfected cells maintained in 1 μ M tyrosine ranged between 5-40%, unlike the results with the endogenous OA1 protein

that were reproducibly ~5%.

[0071] Not only was the distribution of OA1 in transfected cells sensitive to tyrosine levels in the medium, total OA1-GFP expression was increased 5-fold in cells maintained in 1 μ M tyrosine. To verify that this difference related to OA1 expression rather than cell number, actin expression was evaluated from the paired samples. The data (Fig. 1 D) presented as optical density units indicate no difference in actin. The amount of cell surface OA1 between the normal and low tyrosine groups was also compared. Importantly, in the five RPE experiments and six OA1-GFP in COS experiments, OA1 in the plasma membrane fraction of cells in standard medium was not reproducibly detected, similar to that found by others.

[0072] The distribution of OA1 in RPE cells also was evaluated by confocal microscopy. OA1 has previously been characterized as an endosomal protein in cultured RPE cells as shown in (Fig. 1 E). In contrast, the distribution of OA1 in low tyrosine medium was diffuse on the plasma membrane of cultured RPE cells, with little endosomal accumulation (Fig. 1 F), an observation consistent with the results obtained using biochemical methods.

L-DOPA as a Natural Agonist for OA1.

[0073] Tyrosinase function in melanogenesis begins with its activity on tyrosine to create L-DOPA, followed by a second reaction to create dopaquinone that leads to pigment formation [25]. Of the intermediates between tyrosine and melanin, L-DOPA has the greatest half-life, and L-DOPA is released into the subretinal space apical to the RPE when melanin synthesis occurs [23,24]. L-DOPA is also the precursor to dopamine, a neurotransmitter produced by dopaneuritic neurons from tyrosine. The release of calcium from intracellular stores is a common downstream effect of GPCR activation by a ligand. Since the expression of OA1 on the cell surface appears to be sensitive to tyrosine, it was examined whether tyrosine, or its metabolites L-DOPA and dopamine, could stimulate influx of Ca^{2+} into the cytoplasm in an OA1-dependent manner. CHO cells were transfected with an OA1 expression vector then maintained in DMEM containing 1 μ M tyrosine for 48 hours followed by tyrosine-free DMEM for 24 hours to facilitate cell surface expression of OA1. Intracellular Ca^{2+} was evaluated using Fura-2, and $[Ca^{2+}]_i$ was determined by ratiometric imaging [26]. In the absence of any ligand, $[Ca^{2+}]_i$ was not significantly different between transfected and untransfected cells (Fig 2). Tyrosine and several tyrosine metabolites were tested at 1 μ M for an effect on $[Ca^{2+}]_i$. As a positive control each experiment was ended by treatment with 20 mM KCl to depolarize the cell and increase $[Ca^{2+}]_i$ via activation of voltage-gated channels. This maneuver served to verify the Fura-2 loading and responsiveness of the cells being tested (Fig. 2). Only L-DOPA elicited a significant increase in $[Ca^{2+}]_i$ (Fig. 2 A). Tyrosine and dopamine had no positive effect on intracellular at $[Ca^{2+}]_i$ concentrations up to 1 mM (not shown). The slight negative effect of 1 μ M dopamine was not statistically significant, but reproducible among the 11 experiments with dopamine (Fig. 2 B).

[0074] Over expression of GPCRs in non-native cell lines can lead to false signal transduction coupling. To verify that OA1 signaling in response to L-DOPA was indeed a natural response, OA1 was expressed in RPE cells (Fig. 2 C). Results using transfected RPE cells were similar to those achieved with transfected CHO cells. RPE cells transfected to express OA1 responded to 1.0 μ M L-DOPA with an increase in $[Ca^{2+}]_i$. It was next determined whether RPE cells expressing the endogenous OA1 receptor, at endogenous levels exhibited L-DOPA responsiveness. Like all of the transfected cell experiments, RPE expressing OA1 demonstrated an increase in $[Ca^{2+}]_i$ after treatment with 1.0 μ M L-DOPA (Fig. 2 C).

[0075] To further characterize OA1 signaling activity, pertussis toxin was used to distinguish between Gq coupled $[Ca^{2+}]_i$ signaling and Gi linked signaling (Fig. 2 C). In all cells studied, pertussis toxin lowered the basal level of $[Ca^{2+}]_i$, indicating its activity on inhibition of the background signaling through Gi subunit activity. Pertussis toxin was used in experiments conducted in cells transfected to express OA1 including both CHO and RPE, as well as RPE expressing the endogenous OA1 protein at natural levels. In all transfected cells tested the measured $[Ca^{2+}]_i$ response to L-DOPA was greater than in the absence of the toxin (Fig 2), owing largely to the lower initial $[Ca^{2+}]_i$. Thus, the signaling through OA1 in response to L-DOPA that results in increase $[Ca^{2+}]_i$ is not pertussis toxin sensitive and likely Gq subunit mediated. The second messenger cAMP was also measured in CHO cells transfected to express OA1 (Fig. 2 D). Using inactive cells or a submaximal forskolin treatment, the experiments were set up to measure either an increase or decrease in cAMP in response to L-DOPA. In six such experiments, no change in cAMP was observed suggesting neither Gs nor Gi subunits are involved in OA1 signaling.

[0076] Standard methods of radiolabeled ligand binding were used to characterize the interaction between OA1 and L-DOPA (Fig. 3 A). CHO cells were transfected to express OA1, then binding of L-DOPA was quantified in a concentration-dependent manner, and the results were further characterized by Scatchard Plot analysis (Fig. 3E). Results illustrate saturable binding of L-DOPA to OA1 expressing cells with a Kd of 9.35×10^{-6} M. No specific binding was observed in untransfected CHO cells, indicating that the cells do not have an endogenous L-DOPA receptor (not shown). All binding parameters, total, specific, and nonspecific are shown as supplemental data (Figure 6A). Tyrosine exhibited the potential to interact with OCA1, but neither tyrosine nor dopamine stimulated OA1 signaling (see Fig. 2). Competitive ligand binding was used to determine whether either tyrosine or dopamine competed with L-DOPA for OA1 binding. At high

concentrations (1 mM), both tyrosine and dopamine competed with L-DOPA for OA1 binding (Fig. 3B). To further characterize this the kinetics of the competition between L-DOPA and either dopamine (Figure 3 C) or tyrosine (Fig. 6B) was examined. Dopamine exhibited competitive binding to a single site with L-DOPA with a K_i of $2.33 \times 10^{-6} \pm 0.2 \times 10^{-6}$ M. Similar experiments with tyrosine demonstrated inhibition of L-DOPA binding only at high concentrations (Fig. 6B).

5 Saturation kinetics were not possible with tyrosine because of its low affinity and insolubility at the high concentrations.

[0077] Given the relatively low affinity of OA1 for L-DOPA it was determined whether its signaling activity was dose-dependent in the range of this binding affinity. The concentrations in which binding data suggested the steepest rise in association between L-DOPA and OA1, 1.0 - 10 μ M were tested, and results illustrate a concentration dependent GPCR response as measured by $[Ca^{2+}]_i$ (Fig. 3 C). Thus, the activation kinetics of L-DOPA and OA1 matched the concentration

10 range observed in radiolabeled ligand binding experiments.

[0078] In response to ligand binding, GPCRs recruit β -arrestin to the plasma membrane which is followed by internalization of the ligand-receptor complex [27-33]. The effect of L-DOPA on β -arrestin localization was then tested (Fig. 4). Cells were transfected to express OA1 then cultured in 1 μ M tyrosine DMEM for 48 hours prior to analysis to allow cell surface expression of the protein. Cells were then treated with 1 μ M L-DOPA followed by rapid fixation on ice in cold 15 methanol. Initially, under resting conditions in the absence of an agonist, OA1-GFP was found at the cell surface and β -arrestin was diffuse in the cytoplasm (Fig. 4 A-C), with no co-localization between the proteins. After stimulation with L-DOPA, OA1 and β -arrestin were co-localized at the plasma membrane (Fig. 4 D-F). Untransfected cells showed no response to L-DOPA treatment (Fig. 4 G,H), illustrating that the L-DOPA effect on β -arrestin distribution was OA1 dependent, similar to results obtained for $[Ca^{2+}]_i$.

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Effects of 1-DOPA on PEDF Secretion

[0079] Mutations in OA1 cause defects in the development of the neurosensory retina. In previous work it has been shown that pigmented RPE secrete significantly more PEDF than nonpigmented RPE [34], and PEDF is a neurotrophic 25 factor with the potential of altering neurosensory retina development [35-41]. Mutations in OA1 cause a loss of pigmentation in the RPE, suggesting that OA1 activity governs RPE pigmentation. Thus, it was determined whether L-DOPA stimulation of pigmented RPE cells caused increased secretion of PEDF (Fig. 5). This assay is made somewhat more difficult because pigmenting RPE cells produce L-DOPA, which is the agonist for OA1, and OA1 is not readily detectable 30 in nonpigmented cultures of RPE. Thus, pigmented RPE were used to determine whether L-DOPA stimulation increases PEDF expression/secretion. RPE cells were placed in tyrosine-free medium for 24 hours then treated with 1 μ M L-DOPA for one hour. After treatment, the cells were returned to standard medium without exogenous L-DOPA for three days. Control cells were not treated with L-DOPA, but the medium was changed at the same time the experimental cells were 35 returned to normal medium. Conditioned medium was collected after three days and PEDF was measured. Results illustrate a significant increase in the secretion of PEDF in pigmented cells treated with L-DOPA when compared to paired, control monolayers of pigmented RPE (Fig. 5 A). Importantly, this significant increase occurred in cells which were pigmenting and therefore expressed OA1 and had a basal level of PEDF expression.

[0080] To determine whether pigmented RPE cells secrete PEDF through an autocrine loop involving tyrosinase 40 activity and OA1 signaling, a specific tyrosinase inhibitor phenylthiourea (PTU) was used to inhibit pigmentation and L-DOPA production (Fig. 5B). In these experiments, pigmented RPE cells were either maintained in DMEM, or DMEM containing 200 μ M PTU for three days, then PEDF secretion was measured. Pigmented RPE secreted substantial PEDF, but PTU caused a significant decrease in PEDF secretion indicating that tyrosinase activity is necessary for the high 45 level of PEDF secretion observed in pigmented RPE cells. To verify that it was the lack of L-DOPA in the PTU treated cells that caused the decreased PEDF secretion, 3 different cultures of pigmented RPE were used, and exposed to PTU for 48 hours, then treated with 1.0 μ M L-DOPA in the continued presence of PTU; PEDF was measured after 72 hours (Fig. 5 C). The data are presented as percent of control for this experiment because the cultures used varied in both pigmentation and PEDF expression before the experiment began. PTU treated RPE responded to the added L-DOPA by increasing PEDF secretion, indicating that the effect of PTU on PEDF secretion is caused by the lack of L-DOPA production when tyrosinase is inhibited.

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Discussion:

[0081] There is a complex inter-tissue relationship between the RPE and the neurosensory retina. One aspect of this 55 relationship is centered on RPE pigmentation, and defects in melanin synthesis which result in significant neurosensory retina alterations [8,23,42]. The data suggest that OA1 and tyrosinase participate in an autocrine loop through L-DOPA that regulates the secretion of at least one potent neurotrophic factor, PEDF. The data also suggest that the pathologic changes in retinal development that occur in albinism may result from changes in the activity of the OA1 signaling pathway. Reduced OA1 signaling activity can be caused either directly through OA1 mutations or indirectly through changes in L-DOPA production by tyrosinase activity. Thus, it is hypothesized that the similar retinal phenotypes that

accompany the diverse forms of albinism can be reconciled to a single common pathway, OA1 signaling.

[0082] In the study, OA1 on the apical surface of human RPE *in situ* was observed. Previous reports have suggested that OA1 in mice is localized to the melanosome [22], and in cultured cells to the endosomal compartment [15-18,20-22,43]. The results from *in situ* RPE preparations indicate that OA1 is distributed to the apical surface of the RPE. The limited quantities of OA1 on the surface of the RPE (~3.5% of total OA1) may account for the lack of observation of the protein in previous studies where immunogold electron microscopy was used. Like many cell surface GPCRs, OA1 is not an abundant protein.

[0083] The endosomal localization of OA1 reported in previous studies using cultured cells was reproduced in this study for both the endogenous protein and the transgenic protein. When tested in normal culture medium little detectable OA1 protein on the cell surface was found, in agreement with all previous work. However, reduction of tyrosine in the medium caused a modest increase in cell surface receptor accumulation of both the endogenous and recombinant OA1 proteins. This suggests that the distribution of OA1 to the cell surface in cultured cells is sensitive to tyrosine. A previous study has demonstrated OA1 could be localized to the cell surface when endocytosis is inhibited [17] and OA1 on the apical surface of human RPE was observed *in situ*. The data suggest OA1 is a cell surface GPCR, but is a target for endocytosis that may be stimulated by tyrosine or tyrosine metabolites. In this regard, the results differ from past reports of OA1 localization that have classified OA1 as a unique type of intracellular GPCR. Most GPCRs are cell surface proteins that are internalized by a variety of signals, and the data suggest OA1 is similar to most other GPCRs.

[0084] OA1 signaling activity was stimulated by L-DOPA, but not by either its precursor, tyrosine, or its neuronal metabolite dopamine. This result suggests an exquisitely sensitive receptor activity able to distinguish between closely related molecules, after all L-DOPA and tyrosine differ by a sole hydroxyl group. OA1 is sensitive to tyrosine, as tyrosine causes an intracellular localization of OA1 in cultured cells. However, no signaling response to tyrosine was noted, and competition binding studies suggest that tyrosine has a low affinity for OA1. The data suggest that the continuous exposure of cells to high concentrations of tyrosine present in normal medium is sufficient to result in internalization of OA1, but it is unlikely to result in measurable OA1 activation. Strong evidence of a single site competitive interaction between L-DOPA and dopamine was found. The K_i observed for dopamine was similar to the K_d observed for L-DOPA, suggesting that the affinity for the two tyrosine metabolites is similar. The results illustrated a slight, but reproducible, decrease in OA1 signaling from dopamine, suggesting that dopamine may be an effective antagonist or inverse agonist for OA1.

[0085] As an orphan GPCR, its signaling pathway has not previously been identified. In this study it was illustrated that OA1 signaling in response to L-DOPA causes an increase in $[Ca^{2+}]_i$. The data illustrate that the increased $[Ca^{2+}]_i$ observed in response to L-DOPA was insensitive to pertussis toxin and no effects on cAMP were found, indicating that OA1 is likely signaling through a Gq subunit. Previous work has suggested that OA1 can associate with multiple subunits in transfected cells including members of the G_o, G_i, and Gq subunit families. Innamorati *et al.* has shown that spontaneous activity of overexpressed OA1 is likely signaled through a Gq subunit [16]. The data indicate that ligand-dependent signaling from endogenous OA1 in RPE most likely occurs through a Gq mediated pathway, and no promiscuous coupling activities were observed when comparing OA1 over expression in CHO and RPE to natural OA1 expressed in RPE. Interestingly, two overactive mutant forms of Gq subunits cause hyperpigmentation in skin and hair [44], but whether they have an effect in RPE is unknown. RPE and cutaneous melanocytes use the same enzymes to produce pigmentation but differ in their control of melanogenesis. A recent report suggests that OA1 may signal through G α i3, because the retinal phenotype of OA1^{-/-} and G α i3^{-/-} are similar [45]. That study provided no data regarding interaction or signaling between G α i3 and OA1, and the results do not support OA1 signaling through G α i3. However, both OA1 and G α i3 could have activity in convergent pathways that govern some part of the complex system of retinal development.

[0086] The response of OA1 to L-DOPA was measured in three ways, increased $[Ca^{2+}]_i$, recruitment of β -arrestin to plasma membrane OA1, and the increased secretion of PEDF. In addition, inhibiting the activity of tyrosinase in pigmented RPE inhibits L-DOPA production, and results in a decreased secretion of PEDF. Taken together, these studies present a strong argument for a productive ligand:receptor relationship between L-DOPA and OA1. Further, the data suggest selectivity among tyrosine and its metabolites, with only L-DOPA being a productive ligand for OA1. We have determined the binding kinetics between OA1 and L-DOPA, and observed a typical one site receptor:ligand relationship between the two. The binding affinity between OA1 and L-DOPA, with a K_d in the μM range, is not uncommon for an endogenous ligand:receptor relationship. Future identification of a specific, high affinity antagonist for OA1 will aid in further biochemical characterization of the interaction between OA1 and L-DOPA, and be useful in determining whether dopamine is an inverse agonist.

[0087] This study illustrated the selective activation of OA1, an orphan GPCR, by L-DOPA, an intermediate product of melanin synthesis. This study has also illustrated that OA1 activity stimulates PEDF secretion by RPE, a molecule that has the potential to support normal retinal development [40,41]. In humans, this suggests that pharmacologic intervention through OA1 activation could be useful for albinism caused by defects in the melanogenic machinery (OCA 1-4). Unfortunately, the data also suggest that OA1 is necessary for such pharmacologic intervention, and mutations in Oa1 are the most common cause of albinism.

Methods:**Cell Culture**

5 [0088] **RPE-** Cells were isolated as described [46] and maintained in Dulbecco's modified essential medium (DMEM) supplemented with 5% fetal bovine serum (FBS). For experiments in which tyrosine concentrations were lowered, custom manufactured DMEM produced without tyrosine by JRH Biosciences (Lenexa, KS) was used. Dialyzed FBS was purchased from Invitrogen, (San Diego, CA).

10 [0089] **COS-7** and **CHO-** Cells were obtained from ATCC and cultured in DMEM supplemented with 5% FBS. For analysis of OA1 distribution, cells were cultured in tyrosine-free DMEM supplemented with 1 μ M tyrosine, 5% dialyzed FBS for 2-4 days, then tyrosine-free media as described for the experiment.

Cell Surface Biotinylation

15 [0090] **Human RPE *in situ*-** Human eyecups were produced by dissection ~2mm anterior to the equator and removals of the anterior segment. The vitreous and retina were removed without impairing the underlying RPE monolayer, and the retina was cut at the optic nerve head. The resulting eyecups with RPE exposed were rinsed three times with reaction buffer (100 mM NaCl, 50 mM NaHCO₃, pH 8.0) then filled with Sulfo-NHS-LC-Biotin (1 mg/ml) two times for thirty minutes. The reaction was stopped with TG buffer (25 mM Tris, 192 mM Glycine, pH 8.3) then the cells were harvested in lysis buffer (2 mM EDTA, 1% Triton X and 1% Tween 20 in Tris Base Saline Buffer) containing Halt Protease Inhibitor Cocktail. Intact cells and pigment granules were removed by centrifugation at 14,000 rpm for 20 minutes. Biotinylated proteins were captured overnight with immobilized streptavidin beads and then mixed with 4X reducing buffer (250 mM Ttis, pH 6.8, 8% SDS, 40% Glycerol, 20% Beta-mercaptoethanol, 0.08% bromophenol blue). The OA1 protein was separated on a 10% SDS-PAGE gel and identified by a using a polyclonal rabbit OA1 antibody for western blot analysis. Paired western blots were probed with a monoclonal antibody directed against actin.

20 [0091] **Cultured Cells-** RPE and transfected cells were maintained in DMEM containing tyrosine concentrations described for the experiments. Cultures were rinsed three times in reaction buffer, then biotinylated as described above for the *in situ* preparation.

Cloning of Oa1

25 [0092] A cDNA library was constructed from pooled tissue from 6 human donor eyes. Total RNA was harvested using Trizol reagent, then cDNA was synthesized using Poly-T primers for the first strand synthesis, and random hexamers for the second strand. Following cDNA synthesis, RNA was removed using RNase A. The coding sequence for OA1 was obtained by PCR using terminal primers that added restriction sites to the 5' and 3' ends and removed the native stop codon. The PCR product was ligated in frame with GFP in the pEGFP N-1 vector (Clontech). The sequence was verified by automated sequencing in both directions over the entire sequence.

Immunocytochemistry

30 [0093] Cells on slides were fixed with 3% paraformaldehyde at RT, rinsed with 0.1% Triton X-100 in 10% milk in TBST then blocked with 10% milk in TBST. β -arrestin was visualized using a polyclonal antibody directed against β -arrestin, and incubated overnight at 4°C. Cover slips were mounted using 50% glycerol and immunostaining was analyzed by optical sectioning using a Nikon Eclipse E800 laser scanning confocal microscope powered by Compix Confocal Imaging Systems software (Simple PCI Version 4.0.6.1605). Three-dimensional analysis of OA1-GFP and β -arrestin distribution was performed in Image J 1.32.

Measurement of [Ca²⁺]i

35 [0094] OA1-GFP expressing CHO cells plated on glass cover slips were rinsed in Ca²⁺ containing HEPES buffered Hanks Balanced Salt Solution (HBSS) (pH 7.45), then incubated with 2.5 μ M Fura-2 (solubilized in anhydrous dimethylsulfoxide and 0.002% pluronic acid) for 20 minutes at 37°C, 5% CO₂. The Fura-2 loaded cells were rinsed with HBSS for 15 minutes at 37°C, 5% CO₂ to allow for full cleavage of the dye to its active form. Each cover slip was incubated in 1 ml of HBSS in a chamber held at 37°C on the stage of an inverted Olympus IX70 microscope equipped with a 40 x 55 1.35 NA UV-fluor objective.

40 [0095] Using a filter wheel, excitation light from a 200 W Xe bulb was passed alternately through 340 and 380 nm filters. A 10 nm bandpass filter, centered at 510 nm, selected for the emitted fluorescence which was passed to a CCD camera (Photometrics CH-250). For each experiment, image pairs were taken every minute for the first three minutes,

which established a stable baseline. Then L-DOPA (1 μ M final concentration) was added and image sets were taken every 30 seconds for the next three minutes. Finally, KCl (20 mM final concentration) was added one minute before completion of each experiment as a positive control to establish that the cells were loaded with Fura-2. The same was repeated independently for tyrosine and dopamine (both at 1 μ M final concentration). Using a Silicon Graphics Personal 5 IRIS computer, the 340/380 nm ratio was computed for each pixel within a cell, and then analyzed using Microsoft Excel version 4.0 (Microsoft, Redmond, WA). Once the 340/380 nm ratio was determined, each ratio was normalized to 1 (ratio at time zero divided by itself), then the free ion concentration was calculated using the following equation:

$$[Ca_i] = Kd * (R - R_{min}) / (R_{max} - R)$$

in which R , R_{min} , and R_{max} are the measured, minimum, and maximum ratios, respectively. R_{max} represents the ratio of fluorescence intensity of ion-sensitive wavelengths under fully deprotonated conditions, whereas R_{min} is the ratio for the dye when it is fully protonated. In the case of Fura-2, R increases with increasing Ca^{2+} ; hence R_{min} represents Fura-2 in the absence of Ca^{2+} ($Ca^{2+} < 1$ nM) whereas R_{max} represents the Ca^{2+} -Fura-2 chelate as previously described [26]. 15 R_{min} , R_{max} and Kd were determined in independent experiments in Fura-2 loaded cells, and subsequently utilized for calculation of free Ca^{2+} for the experimental procedures.

Radiolabeled Ligand Binding

20 [0096] CHO cells were transfected to express OA1-GFP were plated into 24-well plates. Cells were chilled to -2C, then rinsed in cold binding buffer, 25 mM Tris, 150 mM NaCl, 5 mM EDTA, 5 μ M digitonin (pH 7.45). Cells were incubated for two hours in binding buffer containing [3 H]-L-DOPA (Moravek Biochemicals, Brea, CA) at concentrations between 10⁻⁴M to 10⁻⁹M. The temperature was not allowed to exceed -2°C at any step of the assay. Controls included assays 25 conducted on nontransfected CHO and specific binding was determined by competition with excess unlabelled L-DOPA at 10⁻³M. Bound L-DOPA was quantified by scintillation spectroscopy.

Measurement of cAMP

30 [0097] Cells were pretreated with forskolin (15 minutes) then challenged with L-DOPA using an assay setup as previously described [47]. After 1 minute of ligand exposure, cells are scraped into ice-cold buffer, boiled then centrifuged. Equivalent volumes, 50 μ l, of supernate and 3 H-cAMP (New England Nuclear) then combined with 100 μ l cold PKA. After 2 hours, the solution is passed over activated charcoal, and supernates are counted in a scintillation counter. Results are compared to those achieved using a standard curve, instead of cytosol, produced using 50 μ l of cAMP 35 0.25-32.0 pmole/50 μ l.

Example 2: The OA1 loop functions in vivo

40 [0098] PEDF secretion in OA deficient mice was compared to wild type mice, and showed that wild-type mice secreted significantly more PEDF than OA1-/y mice. The culture medium (C.M.) used contains PEDF, and it is likely that PEDF in the CM from OA1-/y is from the medium used, not the RPE. Results (Figure 7) are quantified and summarized in the graph. The difference, even with the background PEDF in the CM for both groups is significant. T-test analysis results are presented

45 [0099] Tyrosinase deficient pregnant mice were maintained under normal conditions (No L-DOPA), or supplemented with 1.0mg/ml L-DOPA in their drinking water, beginning on embryonic day 7 for their pups. Animals were maintained on supplemental until postnatal day 14, when ocular development is over and the eyes are open.

50 [0100] Two cell types are reduced in number in albinism: retinal ganglion cells and photoreceptors. Figure 8A demonstrates that L-DOPA supplementation increases retinal ganglion cell numbers compared to what is expected in a normal wild-type mouse. Figure 8B shows the same result for photoreceptors. Photoreceptors are not counted directly as they are too dense. Rather, the area occupied by photoreceptor nuclei is measured as a measure of photoreceptor numbers. L-DOPA supplementation increased the photoreceptor nuclear area, so the number of photoreceptors were increased. Again, this appeared to restore the albino animal to normal levels.

55 [0101] As shown in Figure 8C, Four paired littermate animals, 2 wild-type and 2 OA1 -/y (female OA1 deficient) were euthanized and the retinas from each animal were loaded independently in a lane, then proteins were western blotted to detect PEDF, which was readily observed in the retina from wild-type mice. In contrast, PEDF is not readily detected in the retinas from the OA1 -/y mice.

[0102] In summary this data illustrate that OA1 -/y mice make less PEDF than wild type mice. L-DOPA stimulation in

tyrosinase defective mice rescues the two most prominent neurosensory retina defects of albinism: a loss of photoreceptor cells and retinal ganglion cells. Finally, PEDF levels are reduced in the retinas of mice lacking OA 1. Thus, it is concluded that the OA1 autocrine loop functions *in vivo*, and can be stimulated with oral L-DOPA.

[0103] The data together illustrate that the linkage between RPE pigmentation and AMD are likely through the signaling activity of OA1. The data illustrate that the ligand for OA1 is L-DOPA, and that OA1 signaling from L-DOPA controls the expression of PEDF. PEDF is the most potent neurotrophic factor made by RPE. Thus, the identification of L-DOPA as the ligand for OA1, which controls PEDF expression, ties together L-DOPA and neurotrophic activity in the RPE. Because L-DOPA is produced as a by-product of pigment production, this established for the first time a linkage between RPE pigmentation and neurotrophic activity. This system is defined as the OA1 autocrine loop. Tyrosinase makes pigment and releases L-DOPA. Released L-DOPA binds to and initiates signaling through OA1. OA1 signaling controls the expression of both tyrosinase and PEDF.

[0104] To date the data illustrate this model biochemically, in cultured cells, and *in vivo*. The fact that retinal development in an albino animal can be rescued using dietary L-DOPA indicates that dietary L-DOPA can be used to stimulate RPE trophic factor expression *in vivo*. AMD is clearly tied to an RPE defect somehow related to its pigmentation. Blue-eyed individuals get AMD at a much greater frequency than dark-eyed individuals, so the level of RPE pigmentation controls the AMD process. The level of RPE pigmentation is controlled by OA1 signaling and is part of the same OA1 autocrine loop described above. Thus, AMD is related to OA1 signaling in RPE. Therefore, those with lower RPE pigmentation will have lower tyrosinase, lower L-DOPA, lower OA1 signaling, and lower PEDF production. We can use dietary L-DOPA as ligands for OA1 and stimulate that activity. The final determinant of the health of the neurosensory retina is PEDF, but we can use OA1 signaling to increase the OA1 loop activity, and increase the neurotrophic activity of the RPE. The effect of OA1 signaling will be to foster neuron survival.

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5 Arg Thr Gln Pro Met Ala Ser Pro Arg Leu Gly Thr Phe Cys Cys Pro
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10 Thr Arg Asp Ala Ala Thr Gln Leu Val Leu Ser Phe Gln Pro Arg Ala
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15 Phe His Ala Leu Cys Leu Gly Ser Gly Gly Leu Arg Leu Ala Leu Gly
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Leu Leu Gln Leu Leu Pro Gly Arg Arg Pro Ala Gly Pro Gly Ser Pro
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20 Ala Thr Ser Pro Pro Ala Ser Val Arg Ile Leu Arg Ala Ala Ala Ala
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25 Cys Asp Leu Leu Gly Cys Leu Gly Met Val Ile Arg Ser Thr Val Trp
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30 Leu Gly Phe Pro Asn Phe Val Asp Ser Val Ser Asp Met Asn His Thr
115 120 125

Glu Ile Trp Pro Ala Ala Phe Cys Val Gly Ser Ala Met Trp Ile Gln
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35 Leu Leu Tyr Ser Ala Cys Phe Trp Trp Leu Phe Cys Tyr Ala Val Asp
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40 Ala Tyr Leu Val Ile Arg Arg Ser Ala Gly Leu Ser Thr Ile Leu Leu
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	Tyr His Ile Met Ala Trp Gly Leu Ala Thr Leu Leu Cys Val Glu Gly			
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5	Ala Ala Met Leu Tyr Tyr Pro Ser Val Ser Arg Cys Glu Arg Gly Leu			
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10	Asp His Ala Ile Pro His Tyr Val Thr Met Tyr Leu Pro Leu Leu Leu			
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	Val Leu Val Ala Asn Pro Ile Leu Phe Gln Lys Thr Val Thr Ala Val			
	225	230	235	240
15	Ala Ser Leu Leu Lys Gly Arg Gln Gly Ile Tyr Thr Glu Asn Glu Arg			
	245	250	255	
20	Arg Met Gly Ala Val Ile Lys Ile Arg Phe Phe Lys Ile Met Leu Val			
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25	Leu Ile Ile Cys Trp Leu Ser Asn Ile Ile Asn Glu Ser Leu Leu Phe			
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	Tyr Leu Glu Met Gln Thr Asp Ile Asn Gly Gly Ser Leu Lys Pro Val			
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Ala Thr Gln Leu Val Leu Ser Phe Gln Pro Arg Val Phe His Ala Leu
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Cys Leu Gly Ser Gly Thr Leu Arg Leu Val Leu Gly Leu Leu Gln Leu
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15

Leu Ser Gly Arg Arg Ser Val Gly His Arg Ala Pro Ala Thr Ser Pro
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Ala Ala Ser Val His Ile Leu Arg Ala Ala Thr Ala Cys Asp Leu Leu
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Gly Cys Leu Gly Ile Val Ile Arg Ser Thr Val Trp Ile Ala Tyr Pro
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Glu Phe Ile Glu Asn Ile Ser Asn Val Asn Ala Thr Asp Ile Trp Pro
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Ala Thr Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Leu Tyr Ser
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Ala Cys Phe Trp Trp Leu Phe Cys Tyr Ala Val Asp Val Tyr Leu Val
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Ile Arg Arg Ser Ala Gly Arg Ser Thr Ile Leu Leu Tyr His Ile Met
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Ala Trp Gly Leu Ala Val Leu Leu Cys Val Glu Gly Ala Val Met Leu
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Tyr Tyr Pro Ser Val Ser Arg Cys Glu Arg Gly Leu Asp His Ala Ile
 180 185 190

Pro His Tyr Val Thr Thr Tyr Leu Pro Leu Leu Val Leu Val Ala
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Asn Pro Ile Leu Phe His Lys Thr Val Thr Ser Val Ala Ser Leu Leu

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210

215

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5 Lys Gly Arg Lys Gly Val Tyr Thr Glu Asn Glu Arg Leu Met Gly Ala
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10 Val Ile Lys Thr Arg Phe Phe Lys Ile Met Leu Val Leu Ile Ala Cys
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15 Trp Leu Ser Asn Ile Ile Asn Glu Ser Leu Leu Phe Tyr Leu Glu Met
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20 Gln Pro Asp Ile His Gly Gly Ser Leu Lys Arg Ile Gln Asn Ala Ala
 275 280 285

25 Arg Thr Thr Trp Phe Ile Met Gly Ile Leu Asn Pro Ala Gln Gly Leu
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30 Leu Leu Ser Leu Ala Phe Tyr Gly Trp Thr Gly Cys Ser Leu Asp Val
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35 His Pro Pro Lys Met Val Ile Gln Trp Glu Thr Met Thr Ala Ser Ala
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40 Ala Glu Gly Thr Tyr Gln Thr Pro Val Arg Ser Cys Val Pro His Gln
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45 Asn Pro Arg Lys Val Val Cys Val Gly Gly His Thr Ser Asp Glu Val
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50 Leu Ser Ile Leu Ser Glu Asp Ser Asp Ala Ser Thr Val Glu Ile His
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<213> Xenopus tropicalis

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Ala Thr Gln Leu Val Leu Asp Phe Gln Pro Gln Ile Tyr Gly Ser Leu
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5 Cys Ile Gly Ser Gly Leu Val Ser Leu Leu Leu Thr Ile Val Gln Leu
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10 Leu Pro Lys Thr Lys Gln Gly Tyr Arg Arg Leu Gly Arg Ala Met Leu
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15 Pro Lys Pro Ser Ser Arg Ile Leu Phe Leu Val Ile Ile Cys Asp
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20 Leu Leu Gly Cys Leu Gly Ile Leu Ile Arg Ser Ser Val Trp Ile Ser
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25 Ser Pro Gly Phe Ile Ser Asn Met Ser Leu Met Asn Thr Ser Asp Ile
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30 Trp Pro Ser Thr Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Phe
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35 Tyr Ser Ala Ser Phe Trp Trp Leu Phe Cys Tyr Ala Ile Asp Ala Tyr
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40 Leu Val Val Arg Arg Ser Ala Gly Ile Ser Thr Ile Val Leu Tyr His
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45 Met Met Thr Trp Gly Leu Ala Leu Met Leu Cys Ile Glu Gly Val Ala
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50 Met Leu Tyr Tyr Pro Ser Val Ser Asn Cys Glu Asn Gly Leu Glu His
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Ala Ile Pro His Tyr Val Thr Thr Tyr Ala Pro Leu Leu Ile Val Met
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Phe Ala Asn Pro Ile Leu Phe Arg Arg Thr Val Ala Ala Val Ala Ser
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55 Leu Leu Lys Gly Arg Gln Gly Ile Tyr Thr Glu Asn Glu Arg Arg Leu
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Gly Thr Glu Ile Gln Leu Arg Phe Phe Lys Ile Met Leu Val Phe Met
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Ile Cys Trp Thr Ala Asn Ile Ile Asn Glu Thr Leu Leu Phe Tyr Leu
 260 265 270

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5 Ala Ala Leu Ile Thr Trp Phe Ile Met Gly Ile Leu Asn Pro Met Gln
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10 Gly Phe Leu Phe Thr Ile Ala Phe Tyr Gly Trp Thr Gly Trp Asn Val
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Asp Phe Asn Phe Arg Gln Lys Glu Thr Ala Trp Glu Arg Val Ser Thr
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15 Ser Thr Ile Thr Glu Thr Ala His Asn Gly Thr Asn Gly Ser Phe Leu
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20 Asp Tyr Pro Gly Tyr Ile Gln Asn Gln Asn Lys Thr Glu Ile Gly Asn
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25 Ser Gln Gln Thr Asp Glu Ala Leu Ser Ile Leu Ser Glu Gly Asn Gly
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35 <400> 7

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55	<212> PRT	
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5 Ala Thr Gln Leu Ala Leu Gly Phe Gln Pro Arg Ala Phe His Ala Leu
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10 Cys Leu Gly Ser Gly Ala Leu Arg Leu Ala Leu Gly Leu Leu Gln Leu
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15 Arg Pro Gly Arg Arg Pro Ala Gly Pro Gly Ile Ala Ser Ala Ser Pro
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Ala Thr Ser Ala Arg Val Pro Ala Ser Val Arg Ile Val Arg Ala Ala
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20 Thr Ala Cys Asp Leu Leu Gly Cys Leu Gly Ile Ala Val Arg Ser Ala
85 90 95

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	Val Asp Ala Tyr Leu Val Ile Gln Arg Ser Ala Gly Gln Ser Thr Ile	
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15	Leu Leu Tyr His Leu Met Thr Trp Gly Leu Ala Ala Leu Leu Ser Val	
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20	Glu Gly Ala Leu Met Leu Tyr Tyr Pro Ser Met Ala Arg Cys Glu Arg	
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25	Leu Leu Val Leu Val Gly Asn Pro Ile Leu Phe Arg Lys Thr Val Thr	
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	235	240
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45	Gln Val Arg Asn Ala Ala Lys Thr Thr Trp Phe Met Met Gly Ile Leu	
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50	Asn Pro Ala Gln Gly Phe Leu Leu Ser Leu Ala Phe Tyr Gly Trp Thr	
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	315	320
	Gly Cys Arg Leu Thr Leu Pro Gly Pro Ser Lys Glu Ile Gln Trp Asp	
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55	Ser Met Thr Thr Ser Ala Thr Glu Gly Ala Pro Pro Ser Pro Gly Gly	
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Pro Gln Glu Pro Gly Glu Gly Pro Ala Pro Lys Lys Glu Leu Pro Gly
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5 Gly Thr His Thr Ser Asp Glu Ala Leu Ser Leu Leu Ser Glu Gly Ser
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5 Ala Thr Gln Leu Val Leu Thr Phe Gln Pro Arg Val Phe His Ala Leu
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15 Leu Thr Gly Arg Arg Ser Val Gly His Arg Ala Pro Ala Thr Thr Pro
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Ala Ala Ser Val His Ile Leu Arg Ala Ala Thr Ala Cys Asp Leu Leu
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20 Glu Cys Leu Gly Ile Val Ile Arg Ser Thr Val Trp Ile Ala Tyr Pro
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25 Glu Phe Ile Glu Asn Ile Ser Asn Met Asn Gly Thr Asp Ile Trp Pro
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30 Thr Ala Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Leu Tyr Ser
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40 Ala Trp Gly Leu Pro Val Leu Leu Cys Val Glu Gly Ala Val Met Leu

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5 Tyr Tyr Pro Ser Val Ser Arg Cys Glu Arg Gly Leu Asp His Ala Ile
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10 Pro His Tyr Val Thr Thr Tyr Leu Pro Leu Met Leu Val Leu Val Ala
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Asn Pro Ile Leu Phe His Lys Thr Val Ile Ser Val Ala Ser Leu Leu
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15 Lys Gly Arg Lys Gly Val Tyr Thr Glu Asn Glu Arg Leu Met Gly Ala
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20 Val Ile Lys Thr Arg Phe Phe Lys Ile Met Leu Val Leu Ile Ala Cys
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25 Gln Pro Asp Thr His Gly Gly Ser Leu Lys Arg Ile Gln Asn Ala Ala
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30 Arg Thr Thr Trp Phe Ile Met Gly Ile Leu Asn Pro Ser Gln Gly Leu
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40 Ala Glu Gly Thr Tyr Gln Thr Pro Glu Gly Ser Cys Val Pro His Gln
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45 Asn Pro Arg Lys Val Val Cys Val Gly Gly His Thr Ser Asp Glu Val
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15 Ala Ser Ile Leu Leu Phe Ile Ser Ala Cys Asp Leu Leu Gly Cys Leu
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20 Gly Val Ile Phe Arg Ser Thr Val Trp Leu Gly Phe Pro Asp Phe Val
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25 Gly Asn Ile Ser Val Val Asn Gly Thr Asp Gly Trp Pro Ser Ala Phe
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20 Leu Pro Lys Thr Lys His Gly Tyr Arg Arg His Gly Arg Ser Met Leu
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25 Pro Lys Pro Ser Ser Ser Arg Ile Leu Phe Leu Val Ile Val Cys Asp
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Leu Leu Gly Cys Leu Gly Ile Leu Ile Arg Ser Ser Val Trp Ile Ser
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30 Ser Pro Gly Phe Ile Ser Asn Met Ser Leu Met Asn Thr Ser Asp Ile
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35 Trp Pro Ser Ser Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Phe
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Tyr Ser Ala Ser Phe Trp Trp Leu Phe Cys Tyr Ala Ile Asp Ala Tyr
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40 Leu Val Val Arg Arg Ser Ala Gly Ile Ser Thr Ile Val Leu Tyr His
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45 Met Met Thr Trp Gly Leu Ala Leu Met Leu Cys Val Glu Gly Val Ala
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5 Phe Ala Asn Pro Ile Leu Phe Arg Arg Thr Val Ala Ala Val Ala Ser
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10 Leu Leu Lys Gly Arg Gln Gly Ile Tyr Thr Glu Asn Glu Arg Arg Leu
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15 Ile Cys Trp Thr Ala Asn Ile Ile Asn Glu Thr Leu Leu Phe Tyr Leu
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20 Glu Met Gln Pro Asp Ile Lys Thr Asp Gln Leu Lys Asn Val Arg Asn
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Ala Ala Leu Ile Thr Trp Phe Ile Met Gly Ile Leu Asn Pro Met Gln
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30 Asp Phe Asn Phe Arg Gln Lys Glu Thr Ala Trp Glu Arg Val Ser Thr
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35 Ser Ser Leu Thr Glu Ala Ala His Asn Gly Thr Asn Gly Ser Phe Leu
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Asp Tyr Pro Gly Tyr Ile Gln Asn Gln Asn Lys Thr Glu Ile Gly Asn
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5 Cys Ile Gly Ser Ala Ser Ala Ser Leu Leu Leu Thr Ile Leu Gln Leu
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10 Leu Pro Lys Lys Gly Gln Ser Leu Arg Lys Met Pro Lys Ala Ser Ser
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Ser Ser Thr Ile Leu Leu Ile Ser Val Cys Asp Ile Leu Gly Gly
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15 Ser Gly Val Ile Phe Arg Ser Ser Val Trp Leu Gly Phe Pro Ser Phe
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20 Ile Ala Asn Ile Ser Val Ala Asn Gly Thr Asp Ile Trp Pro Ser Ala
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Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Leu Tyr Ser Ala Gly
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25 Phe Trp Trp Leu Phe Cys Tyr Ala Val Asp Ser Tyr Leu Val Val Arg
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30 Arg Ser Ala Gly Arg Ser Thr Ile Val Leu Tyr His Met Met Ala Trp
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35 Gly Leu Ala Val Leu Leu Cys Met Glu Gly Val Met Leu Leu Tyr Tyr
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40 Tyr Ile Thr Thr Tyr Ala Pro Leu Leu Leu Val Leu Val Val Asn Pro
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45 Val Leu Phe Arg Arg Thr Val Thr Ala Val Ala Ser Leu Leu Lys Gly
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50 Arg Gln Gly Ile Tyr Thr Glu Asn Glu Arg Arg Leu Gly Thr Glu Ile
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Gln Met Arg Phe Phe Lys Ile Met Leu Val Phe Thr Val Cys Trp Ser
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55 Ser Asn Ile Ile Asn Glu Ser Leu Leu Phe Tyr Leu Glu Met Gln Pro
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Asp Ile Asn Glu Thr Pro Leu Lys Asn Ile Arg Ser Ala Ala Leu Ile
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5 Thr Trp Ile Ile Met Gly Val Leu Asn Pro Met Gln Gly Phe Leu Phe
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10 Thr Leu Ala Phe Tyr Gly Trp Thr Gly Trp Lys Val Asp Leu Lys Trp
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Gln Lys Arg Glu Ile Pro Trp Glu Ser Met Ser Ser Ser Thr Val Gly
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15 Asp Asn Asp Tyr Pro Ser Pro Val Asn Tyr Gln Ser Asn Val His Asp
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<213> Danio rerio

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20 Leu Pro Lys Arg Arg Ser Phe Arg Pro Gln Ala His Ser Ser Arg Ala
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25 Ala Ser Ser Ser Arg Ile Leu Thr Ile Ile Ser Val Cys Asp Ile Leu
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30 Gly Cys Thr Gly Ile Ile Arg Ser Ser Leu Trp Ile Gly Leu Pro
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	315	320
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<400> 22

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Cys Leu Gly Ser Gly Ala Leu Arg Leu Ala Leu Gly Leu Leu Gln Leu
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Leu Pro Gly Arg Arg Pro Ala Gly Pro Gly Ser Pro Ala Thr Ser Pro
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Pro Ala Ser Val Arg Ile Leu Arg Ala Ala Thr Ala Cys Asp Leu Leu
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Gly Cys Leu Gly Val Val Ile Arg Ser Thr Val Trp Leu Gly Phe Pro
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Asn Phe Val Asp Ser Ile Ser Asp Val Asn Arg Thr Glu Ile Trp Pro
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Ala Val Phe Cys Val Gly Ser Ala Met Trp Ile Gln Leu Leu Tyr Ser
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Ala Cys Phe Trp Trp Leu Phe Cys Tyr Ala Val Asp Ala Tyr Leu Val
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50	Asn Pro Ala Ser Lys Lys Val Ser Arg Val Gly Gly Gln Thr Ser Asp	
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10 <213> Rhesus macaque

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40 <213> Homo sapiens

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10 Pro Asp Ser Thr Gly Ala Leu Val Glu Glu Glu Asp Pro Phe Phe Lys
35 40 45

15 Val Pro Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp
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Leu Tyr Arg Val Arg Ser Ser Thr Ser Pro Thr Thr Asn Val Leu Leu
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20 Ser Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala
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Ser Asp Leu Ser Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met
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40 Ser Ile Ile Phe Phe Leu Pro Leu Lys Val Thr Gln Asn Leu Thr Leu
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45 Ile Glu Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu
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Tyr Glu Gly Glu Val Thr Lys Ser Leu Gln Glu Met Lys Leu Gln Ser
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55 Leu Phe Asp Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys
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Leu Thr Gln Val Glu His Arg Ala Gly Phe Glu Trp Asn Glu Asp Gly
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5 Ala Gly Thr Thr Pro Ser Pro Gly Leu Gln Pro Ala His Leu Thr Phe
370 375 380

10 Pro Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp
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<212> DNA

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25	Glu Leu Trp Phe Ser Asp Asp Pro Asn Val Thr Lys Thr Leu Gln Phe			
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	210	215	220	
35	Ala Val Leu Thr Gly Lys Lys Val Val His Leu Asp Val Arg Gly Asn			
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45	Gly Ala Glu Val Lys Ser Arg Thr Thr Leu Phe Arg Lys Ile Gly Asp			
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325

330

335

5 Gly Asn Met Gly Lys Ile Leu Pro Glu Tyr Leu Ser Asn Trp Thr Met
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Glu Lys Val Lys Arg Glu Gly Val Lys Val Met Pro Asn Ala Ile Val
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Gln Ser Val Gly Val Ser Gly Gly Lys Leu Leu Ile Lys Leu Lys Asp
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Gly Arg Lys Val Glu Thr Asp His Ile Val Thr Ala Val Gly Leu Glu
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Pro Asn Val Glu Leu Ala Lys Thr Gly Gly Leu Glu Ile Asp Ser Asp
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Phe Gly Gly Phe Arg Val Asn Ala Glu Leu Gln Ala Arg Ser Asn Ile
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Trp Val Ala Gly Asp Ala Ala Cys Phe Tyr Asp Ile Lys Leu Gly Arg
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Arg Arg Val Glu His His Asp His Ala Val Val Ser Gly Arg Leu Ala
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Gly Glu Asn Met Thr Gly Ala Ala Lys Pro Tyr Trp His Gln Ser Met
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Phe Trp Ser Asp Leu Gly Pro Asp Val Gly Tyr Glu Ala Ile Gly Leu
 485 490 495

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Val Asp Ser Ser Leu Pro Thr Val Gly Val Phe Ala Lys Ala Thr Ala
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Gln Asp Asn Pro Lys Ser Ala Thr Glu Gln Ser Gly Thr Gly Ile Arg
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Lys Gly Val Ile Phe Tyr Leu Arg Asp Lys Val Val Val Gly Ile Val
 565 570 575

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<210> 29

<211> 1486

<212> DNA

15 <213> Taeniopygia guttata

<400> 29

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 20 25 30

10 Tyr Arg Gln Gln Ser Ile Arg Thr Ala Thr Ala Asn Val Leu Leu Ser
 35 40 45

15 Pro Phe Ser Leu Ala Thr Ala Leu Ser Gly Leu Ser Leu Gly Ala Gly
 50 55 60

Glu Arg Thr Glu Asp Val Ile Ser Arg Ala Leu Phe Tyr Asp Leu Leu
 65 70 75 80

20 Asn Lys Ala Glu Val His Asp Thr Tyr Lys Glu Leu Leu Ser Ser Val
 85 90 95

25 Thr Gly Pro Glu Lys Ser Met Lys Ser Ala Ser Arg Ile Ile Leu Glu
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Lys Arg Leu Arg Ala Arg Pro Gly Phe His Ser Gln Leu Glu Lys Ser
 115 120 125

30 Tyr Lys Met Arg Pro Arg Ala Leu Ser Gly Asn Thr Gln Leu Asp Leu
 130 135 140

35 Gln Glu Ile Asn Thr Trp Val Arg Gln Gln Thr Lys Gly Arg Ile Met
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40 Arg Phe Met Lys Asp Met Pro Thr Asp Val Ser Ile Leu Leu Ala Gly
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45 Ala Ala Phe Phe Lys Gly Thr Trp Lys Thr Lys Phe Asp Thr Lys Arg
 180 185 190

Thr Ala Leu Gln Asp Phe His Leu Asp Glu Asp Arg Thr Val Lys Val
 195 200 205

50

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Ser Met Met Ser Asp Pro Lys Ala Ile Leu Arg Tyr Gly Phe Asp Ser
 210 215 220

5 Glu Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Glu Gly Ile Ser
 225 230 235 240

10 Ala Met Phe Phe Leu Pro Thr Lys Val Thr Gln Asn Met Thr Leu Ile
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Glu Glu Ser Leu Thr Ser Glu Phe Val His Asp Val Asp Lys Glu Leu
 260 265 270

15 Lys Thr Val His Ala Val Leu Ser Leu Pro Lys Leu Lys Leu Asn His
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20 Glu Glu Ala Leu Gly Ser Thr Leu Lys Glu Thr Arg Leu Gln Ser Leu
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25 Phe Thr Ser Pro Asp Phe Ser Lys Ile Ser Ala Lys Pro Leu Arg Leu
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30 Ser His Val Gln His Lys Ala Met Leu Glu Leu Gly Glu Asp Gly Glu
 325 330 335

35 Ile Glu Tyr His Val Asp Arg Pro Phe Leu Leu Val Leu Arg Asp Asp
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40 Thr Thr Gly Thr Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly
 370 375 380

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45 <210> 31
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 <213> Equus caballus

50 <400> 31

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	<211> 417	
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	<213> Equus caballus	
45	<400> 32	

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55	Pro Asp Ile Thr Gly Ala Pro Val Glu Glu Glu Asp Pro Phe Leu Lys	
	35 40 45	

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	Ser Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala	
10	85	90
	Glu Gln Arg Thr Glu Ser Ser Ile His Leu Ala Leu Tyr Tyr Asp Leu	
	100	105
15	Ile Lys Asn Pro Asp Ile His Gly Thr Tyr Lys Glu Leu Leu Ala Ser	
	115	120
20	125	
	Val Thr Ala Pro Asn Lys Asn Phe Lys Ser Ala Ser Arg Ile Ile Phe	
20	130	135
	140	
	Glu Lys Lys Leu Arg Ile Lys Ser Ser Phe Val Thr Pro Leu Glu Lys	
25	145	150
	155	160
	Ser Tyr Gly Thr Arg Pro Lys Ile Leu Thr Gly Asn Ser Arg Thr Asp	
	165	170
	175	
30	Leu Gln Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Ile	
	180	185
	190	
35	Ala Arg Ser Thr Arg Glu Val Pro Ser Glu Ile Ser Ile Leu Leu	
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	205	
	Gly Val Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg	
	210	215
	220	
40	Lys Thr Ser Leu Gln Asp Phe His Leu Asp Glu Glu Arg Thr Val Thr	
	225	230
	235	240
45	Val Pro Thr Met Ser Asp Pro Lys Ala Ile Leu Arg Tyr Gly Leu Asp	
	245	250
	255	
50	Ser Asp Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met	
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	270	
	Ser Ile Val Phe Phe Leu Pro Gln Lys Val Thr Gln Asn Leu Thr Met	
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55	Ile Glu Glu Ser Leu Thr Ser Glu Phe Leu His Asp Ile Asp Arg Glu	
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	300	

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Leu Lys Thr Val Gln Ala Val Leu Thr Ile Pro Lys Leu Lys Leu Ser
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5 Tyr Glu Gly Glu Val Thr Lys Ser Leu Gln Glu Ile Lys Leu Gln Ser
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10 Leu Phe Asp Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Leu Lys
 340 345 350

15 Leu Thr Gln Val Glu His Arg Ala Gly Phe Glu Trp Asn Glu Asp Gly
 355 360 365

Ala Thr Asn Pro Ser Gln Gly Pro Gln Pro Ala His Leu Thr Phe Pro
 370 375 380

20 Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr
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25 Asp Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly
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30 <210> 33

<211> 1503

<212> DNA

<213> Xenopus (Silurana) tropicalis

35 <400> 33

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55 gcagacacaa gggaaagtgg tgaagttctt caaagagatt ccaactagtg tgagcattct 660

gctgctcgga actacttact taaaaggcca gtggcgatc aaatctaattc ctcggaaaac 720

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<210> 34

<211> 409

<212> PRT

<213> Xenopus (Silurana) tropicalis

<400> 34

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10 Glu Asp Pro Phe Tyr Lys Ser Pro Ile Asn Arg Leu Ala Ser Ser Ala
35 40 45

15 Ser Asn Phe Gly Tyr Asp Leu Tyr Arg Met Gln Ala Asn Lys Asn Pro
50 55 60

Asn Ser Asn Ile Ile Ile Ser Pro Leu Ser Ile Ala Thr Ser Leu Ser
65 70 75 80

20 Ser Leu Ser Leu Gly Gly Gln Arg Thr Glu Ser Leu Ile Gln Arg
85 90 95

25 Ser Leu Tyr Tyr Asp Leu Leu Asn Asp Pro Glu Val His Ala Thr Tyr
100 105 110

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	Lys Asp Leu Leu Ala Ser Phe Thr Ser Gln Ala Ser Gly Leu Lys Ser			
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	130	135	140	
10	Val Thr Gln Val Glu Lys Phe Tyr Gly Asn Lys Pro Lys Val Leu Thr			
	145	150	155	160
Gly Ser Thr Arg Leu Asp Leu Gln Glu Ala Asn Asp Phe Ile Gln Lys				
	165	170	175	
15	Gln Thr Gln Gly Lys Val Val Lys Phe Phe Lys Glu Ile Pro Thr Ser			
	180	185	190	
20	Val Ser Ile Leu Leu Leu Gly Thr Thr Tyr Leu Lys Gly Gln Trp Ala			
	195	200	205	
Tyr Lys Phe Asn Pro Arg Glu Thr Val Gln Arg Glu Phe His Leu Asp				
	210	215	220	
25	Glu Gln Thr Ser Val Thr Val Pro Met Met Ser Ser Lys Asn Ile Pro			
	225	230	235	240
30	Val Arg Tyr Gly Leu Asp Ser Asp Phe Asn Cys Lys Ile Val Gln Leu			
	245	250	255	
35	Pro Leu Thr Gly Gly Val Ser Ile Met Phe Phe Leu Pro Asn Thr Val			
	260	265	270	
40	Thr Gln Asn Leu Thr Met Ile Glu Glu Gly Leu Thr Ser Glu Phe Val			
	275	280	285	
His Asp Ile Asp Gln Ala Leu Gln Pro Ile Asn Leu Val Leu Ser Val				
	290	295	300	
45	Pro Lys Leu Lys Leu Asn Tyr Glu Ala Glu Leu Lys Glu Ala Leu Gln			
	305	310	315	320
Glu Ser Lys Leu Gln Ser Leu Phe Ala Thr Pro Asp Phe Ser Lys Ile				
	325	330	335	
50	Ser Ser Lys Pro Leu Lys Leu Ser Tyr Val Val His Lys Ala Thr Leu			
	340	345	350	
55	Glu Leu Asn Glu Glu Gly Ala Glu Thr Ala Pro Lys Pro Glu Asp Ser			
	355	360	365	

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His Arg Asn Tyr Phe Pro Leu Glu Tyr His Leu Asp His Pro Phe Leu
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5 Phe Val Leu Arg Ala Asn Asp Asn Gly Ala Leu Leu Phe Ile Gly Lys
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10 Val Met Asp Pro Lys Gly Phe Ser Phe
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<210> 35

<211> 1497

<212> DNA

15 <213> Mus musculus

<400> 35

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	<213> Mus musculus	
	<400> 36	

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10 Asp Ser Thr Gly Glu Pro Val Glu Glu Glu Asp Pro Phe Phe Lys Val
35 40 45

Pro Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu
50 55 60

15 Tyr Arg Leu Arg Ser Ser Ala Ser Pro Thr Gly Asn Val Leu Leu Ser
65 70 75 80

20 Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu
85 90 95

25 His Arg Thr Glu Ser Val Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile
100 105 110

30 Thr Asn Pro Asp Ile His Ser Thr Tyr Lys Glu Leu Leu Ala Ser Val
115 120 125

Thr Ala Pro Glu Lys Asn Leu Lys Ser Ala Ser Arg Ile Val Phe Glu
130 135 140

35 Arg Lys Leu Arg Val Lys Ser Ser Phe Val Ala Pro Leu Glu Lys Ser
145 150 155 160

40 Tyr Gly Thr Arg Pro Arg Ile Leu Thr Gly Asn Pro Arg Val Asp Leu
165 170 175

Gln Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Ile Ala
180 185 190

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	Arg Ser Thr Arg Glu Met Pro Ser Ala Leu Ser Ile Leu Leu Leu Gly			
	195	200	205	
5	Val Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys			
	210	215	220	
10	Thr Thr Leu Gln Asp Phe His Leu Asp Glu Asp Arg Thr Val Arg Val			
	225	230	235	240
15	Pro Met Met Ser Asp Pro Lys Ala Ile Leu Arg Tyr Gly Leu Asp Ser			
	245	250	255	
20	Asp Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met Ser			
	260	265	270	
25	Ile Ile Phe Phe Leu Pro Leu Thr Val Thr Gln Asn Leu Thr Met Ile			
	275	280	285	
30	Glu Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu			
	290	295	300	
35	Lys Thr Ile Gln Ala Val Leu Thr Val Pro Lys Leu Lys Leu Ser Phe			
	305	310	315	320
40	Glu Gly Glu Leu Thr Lys Ser Leu Gln Asp Met Lys Leu Gln Ser Leu			
	325	330	335	
45	Phe Glu Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Val Lys Leu			
	340	345	350	
50	Thr Gln Val Glu His Arg Ala Ala Phe Glu Trp Asn Glu Glu Gly Ala			
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55	Gly Ser Ser Pro Ser Pro Gly Leu Gln Pro Val Arg Leu Thr Phe Pro			
	370	375	380	
60	Leu Asp Tyr His Leu Asn Gln Pro Phe Leu Phe Val Leu Arg Asp Thr			
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65	Asp Thr Gly Ala Leu Leu Phe Ile Gly Arg Ile Leu Asp Pro Ser Ser			
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<210> 37

<211> 1810

<212> DNA

<213> Salmo salar

<400> 37

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40	ccacacccctt ttcgtactgc gtggaccatc cttcctgtt cctgggtgagg gatgaggcct	1200
	cgggagcact gctctttatt ggcaaggtgg tcaacccacg caatctgagg atataaacac	1260
	agacacacac tgccttctaa gcaggtccta ggaggggatc agccatcggtt aagcttaagc	1320
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	atttgtgatt gaaaagtaca gctctcataa tttttaataa gaggcacatt cttaacccc	1560
	aaaaataactc atcataatat tgtcaattgc gatgcaagaa ataaacattt aagttaagtc	1620
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55	aaaaaaaaaga	1810

<211> 405
<212> PRT
<213> Salmo salar

5 <400> 38

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Met Leu Arg Thr Thr Leu Leu Leu Cys Leu Gly Ala Leu Leu Ser Leu
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5 Ser Tyr Ala Gln Leu Leu Glu Thr Glu Ala Ala Gly Gly Glu Glu Glu
 20 25 30

10 Ala Val Glu Leu Phe Thr Thr Pro Arg Ala Lys Met Ala Ala Ala Thr
 35 40 45

15 Ser Asp Phe Gly Tyr Asn Leu Phe Arg Ala Leu Ala Gly Arg Asn Pro
 50 55 60

Asn Thr Asn Val Phe Leu Ala Pro Ile Ser Ile Ser Ala Val Leu Thr
 65 70 75 80

20 Gln Leu Ser Met Gly Ala Ser Pro Asp Arg Ser Glu Arg Trp Leu Tyr
 85 90 95

25 Arg Ala Leu Arg Tyr His Thr Leu Gln Asp Pro Gln Leu His Asp Thr
 100 105 110

30 Leu Arg Asp Leu Leu Ala Ser Leu Arg Ala Pro Gly Lys Gly Leu Ser
 115 120 125

Ile Ala Ala Arg Val Tyr Leu Ala Arg Arg Leu Arg Leu Lys Gln Glu
 130 135 140

35 Tyr Phe Gly Val Val Glu Lys Gln Tyr Gly Val Arg Pro Lys Ala Leu
 145 150 155 160

40 Met Gly Gly Ala Lys Asp Val Asn Glu Ile Asn Asp Trp Val Lys Gln
 165 170 175

Gln Thr Gly Gly Lys Val Asp Arg Phe Met Ser Lys Pro Leu Gly Arg
 180 185 190

45 Asn Ser Gly Val Val Pro Leu Gly Ala Ala Tyr Phe Lys Val Lys Trp
 195 200 205

50 Met Thr Arg Phe Ser Gln Ser Gly Val Met Glu Asp Phe Gln Leu Val
 210 215 220

Gly Glu Ala Pro Ala Arg Ile Ser Met Met Gln Gln Asp Asn Tyr Pro
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5 Val Lys Met Gly Val Asp Pro Asp Leu Gly Cys Thr Ile Ala Gln Ile
245 250 255

10 Gln Met Gln Asp Asp Val Ser Met Phe Val Phe Leu Pro Asp Asp Val
260 265 270

Thr Gln Asn Met Thr Leu Val Glu Glu Ser Leu Thr Ala Glu Phe Val
275 280 285

15 Gln Asp Leu Ser Met Thr Leu His Pro Val Gln Thr Ala Leu Thr Leu
290 295 300

20 Pro Val Leu Lys Phe Ser Tyr Ser Thr Asp Leu Leu Pro Leu Leu Thr
305 310 315 320

Asp Leu Gly Leu Asp Glu Phe Leu Ala Asp Thr Asp Leu Thr Lys Ile
325 330 335

25 Thr Ser Gln Ala Ala Lys Leu Gly Ser Leu Asn His Lys Val Val Met
340 345 350

30 Glu Met Ala Pro Glu Gly Thr Gln Tyr Ala Ser Ser Leu Pro Ala Ser
355 360 365

35 Thr Pro Leu Ser Tyr Cys Val Asp His Pro Phe Leu Phe Leu Val Arg
370 375 380

Asp Glu Ala Ser Gly Ala Leu Leu Phe Ile Gly Lys Val Val Asn Pro
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40 Arg Asn Leu Arg Ile
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45 <210> 39

<211> 1422

<212> DNA

<213> Ovis aries

50 <400> 39

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	cccgtaaca agctggcggc agccgtctcc aacttcggct acgacctgta ccgcgtgaga	240
	tctggcgaga gccccaccac caacgtgctg ctgtctccgc tcagcgtggc cacggcgctc	300
10	tctgcccgt cgctgggtgc ggaacagcgg acagaatcca gcattcaccg ggctctgtac	360
	tacgacctga tcagtaaccc agacatccac ggcacctaaca aggacctcct tgcctccgtc	420
15	actgcccccc agaagaacct taaaagtgtc tcccggatta tcttgagag gaagctgcgg	480
	ataaaagcca gttcgtccc acccctcgag aagtcatatg ggaccaggcc cagaatcctg	540
	accggcaact ctcgaataga cttcaggag attaacaact gggtgcaggc ccagatgaaa	600
20	gggaaaattg ctagatccac acggaaata cccagtggaa tcagcattct cttcttgggt	660
	gtggcttact tcaaggggca gtggtaaca aagtttgaact ccaggaagac ttccctggag	720
	gatttccact tggatgaggg gaggaccgtg aaagttccca ttagtgcaga ccctaaggcc	780
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30	gaagagagcc tcacctctga gttcattcat gacatagacc gagaactgaa gactgttcag	960
	gcagtcctga ccattcccaa gctgaagctg agttatgaag gcgaactcac gaagtctgtg	1020
	cagagactga agctacaatc cctgtttgat gcaccagact ttagcaagat cacaggcaaa	1080
35	cctatcaaac ttactcaagt ggaacatcgc atcggattcg agtggaatga ggatggggcg	1140
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	cttaaccaac ctttcatctt tgtactgagg gacacagaca cagggccct tctttcata	1260
40	ggcaaaattc tggacccag aggacttaa tactcaactt aatgttcaaa taccggcaaa	1320
	gaaaaaaaaaca ctagcgggat ggcagattat atattatatg aaggctgccc ctacgttca	1380
45	atgtatactt tgcaataaaa gtgtttctc cttaaaaaaaaaa aa	1422
	<210> 40	
	<211> 416	
	<212> PRT	
50	<213> Ovis aries	
	<400> 40	

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Met Gln Ala Leu Val Leu Leu Leu Trp Thr Gly Ala Leu Leu Gly Phe
1 5 10 15

5 Gly His Cys Gln Asn Ala Gly Pro Glu Ala Gly Ser Leu Ala Pro Glu
20 25 30

10 Ser Thr Gly Ala Pro Val Glu Glu Glu Asp Pro Phe Phe Lys Val Pro
35 40 45

15 Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu Tyr
50 55 60

Arg Val Arg Ser Gly Glu Ser Pro Thr Thr Asn Val Leu Leu Ser Pro

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65	70	75	80
Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln 5 85 90 95			
Arg Thr Glu Ser Ser Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile Ser 100 105 110			
10	115	120	125
Asn Pro Asp Ile His Gly Thr Tyr Lys Asp Leu Leu Ala Ser Val Thr 130 135 140			
Ala Pro Gln Lys Asn Leu Lys Ser Ala Ser Arg Ile Ile Phe Glu Arg 145 150 155 160			
Lys Leu Arg Ile Lys Ala Ser Phe Val Pro Pro Leu Glu Lys Ser Tyr 165 170 175			
25	180	185	190
Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Ile Ala Arg 195 200 205			
Ser Thr Arg Glu Ile Pro Ser Gly Ile Ser Ile Leu Leu Leu Gly Val 210 215 220			
30	225	230	235
Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr 240			
35	245	250	255
Ser Leu Glu Asp Phe His Leu Asp Glu Gly Arg Thr Val Lys Val Pro 260 265 270			
Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Thr Ser Ile 275 280 285			
Ile Phe Phe Leu Pro Gln Lys Val Thr Gln Asn Leu Thr Leu Ile Glu 290 295 300			
40	305	310	315
Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu Lys 320			
Thr Val Gln Ala Val Leu Thr Ile Pro Lys Leu Lys Leu Ser Tyr Glu			

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Gly Glu Leu Thr Lys Ser Val Gln Glu Leu Lys Leu Gln Ser Leu Phe
325 330 335

5 Asp Ala Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys Leu Thr
340 345 350

10 Gln Val Glu His Arg Ile Gly Phe Glu Trp Asn Glu Asp Gly Ala Gly
355 360 365

15 Thr Asn Ser Ser Pro Gly Val Gln Pro Ala Arg Leu Thr Phe Pro Leu
370 375 380

Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr Asp
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20 Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly Thr
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<210> 41

<211> 1465

25 <212> DNA

<213> Cavia porcellus

<400> 41

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	agctgccagg acatgcCAGG caacCCGGAG gactccccgt cccctgaaAG cacaggggAG	180
	ccagtggagg aggaggacCC cttcttcaag gtccctgtga acaagctggc tgcagccatc	240
10	tccaaCTTTG gctacgacCT ataccgggtg agatccatcg agagCCCCAC caccatgtg	300
	ctgctgtccc ccctcagcgt ggccaccGCC ctctctgccc tttcgctggg ggcggAACAG	360
	cgaacagaAG ccaccattCA tcgggctctc tactatgaca tgatcagcaa ccctgacatc	420
15	cacagcacCT acaaggAGCT cctggccACT gtcaccGCC CGCAGAAGAA CCTGAAGAGT	480
	gcttcgagga ttgtcttGA gaggAAAGCTG CGCATAAAAT ccagcATTGT CGCACTACTG	540
	gaaaAGTCAT attcgaccAG gcccagaATC ctgactggca accctcgcat tgaccttcaa	600
20	gagattAGCA actgggtgca ggcccAGATG aaaggAAAAAA tcaccaggTC tacgaggAA	660
	gtgcccAGTG gcatcagcat tctcTTCTC ggtgtggCTT acttcaAGGG gcagtgggtc	720
25	acaaaATTG actccAGAAA gacttCTCTC caggATTCC acttggatGA ggagaggACT	780
	gtaaaAGTTC ccatgatgTC agacCCCAAG gccatcatac gctatggcCT ggataCTgtat	840
	ctcaactgca agattGCCA gctGCCCTG actggaAGCA tgagtatCAT cttcttCTG	900
30	cccattgaggg caacCCAGAA ctGACCATG atagaAGAGA gcctCACCTC CGAGTTGTT	960
	catgacataa accgagaACT gaaggCTgTC caagCGGTTc tcagcatCCC caggCTGAAG	1020
	ctgagTTTCG aaggCGAAct taccaAGTCC ctgcaggAGA tgaAGCTgCA ttccTTGTT	1080
35	gagtccccCG actttagCAA gatCACAGGC aaacCTATCA agCTgACTCA agtggAACAC	1140
	cgggCTggTT tcgagtggAA tgaggAGGG gGCCAGGAA ccagcacCAA ctcagacCTC	1200
40	cagcCTactG gttcacATT ctctCTGGAC tatCACCTGA accAGCCGTT catctTCgTC	1260
	ctgagAGACA cggacACGGG ggCCCTCTC ttcatAGGC AAATTCTGGA ccccAGAAgT	1320
	acttaatGCT ccagTTAAT gttctactAC tctAGAAAGA AACCCAGAA ggatGGCAGT	1380
45	ttatacatta cagggggcA gccccacAG ttcaGtGTA tactttGCAA taaaAGAGCT	1440
	ttatcCTTAA aaaaaaaaaa aaaaa	1465
50	<210> 42 <211> 418 <212> PRT <213> Cavia porcellus	
55	<400> 42	

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Met Gln Val Leu Val Leu Leu Leu Trp Thr Gly Ala Leu Leu Gly Arg
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5 Gly Ser Cys Gln Asp Ile Ala Ser Asn Pro Glu Asp Ser Pro Ser Pro
20 25 30

10 Glu Ser Thr Gly Glu Pro Val Glu Glu Glu Asp Pro Phe Phe Lys Val
35 40 45

15 Pro Val Asn Lys Leu Ala Ala Ala Ile Ser Asn Phe Gly Tyr Asp Leu
50 55 60

Tyr Arg Val Arg Ser Ile Glu Ser Pro Thr Thr Asn Val Leu Leu Ser
65 70 75 80

20 Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu
85 90 95

25 Gln Arg Thr Glu Ala Thr Ile His Arg Ala Leu Tyr Tyr Asp Met Ile
100 105 110

Ser Asn Pro Asp Ile His Ser Thr Tyr Lys Glu Leu Leu Ala Thr Val
115 120 125

30 Thr Ala Pro Gln Lys Asn Leu Lys Ser Ala Ser Arg Ile Val Phe Glu
130 135 140

35 Arg Lys Leu Arg Ile Lys Ser Ser Leu Val Ala Leu Leu Glu Lys Ser
145 150 155 160

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	Tyr Ser Thr Arg Pro Arg Ile Leu Thr Gly Asn Pro Arg Ile Asp Leu			
	165	170	175	
5	Gln Glu Ile Ser Asn Trp Val Gln Ala Gln Met Lys Gly Lys Ile Thr			
	180	185	190	
10	Arg Ser Thr Arg Glu Val Pro Ser Gly Ile Ser Ile Leu Leu Leu Gly			
	195	200	205	
	Val Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys			
	210	215	220	
15	Thr Ser Leu Gln Asp Phe His Leu Asp Glu Glu Arg Thr Val Lys Val			
	225	230	235	240
20	Pro Met Met Ser Asp Pro Lys Ala Ile Ile Arg Tyr Gly Leu Asp Thr			
	245	250	255	
25	Asp Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met Ser			
	260	265	270	
	Ile Ile Phe Phe Leu Pro Met Arg Ala Thr Gln Asn Leu Thr Met Ile			
	275	280	285	
30	Glu Glu Ser Leu Thr Ser Glu Phe Val His Asp Ile Asn Arg Glu Leu			
	290	295	300	
35	Lys Ala Val Gln Ala Val Leu Ser Ile Pro Arg Leu Lys Leu Ser Phe			
	305	310	315	320
	Glu Gly Glu Leu Thr Lys Ser Leu Gln Glu Met Lys Leu His Ser Leu			
	325	330	335	
40	Phe Glu Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys Leu			
	340	345	350	
45	Thr Gln Val Glu His Arg Ala Gly Phe Glu Trp Asn Glu Glu Gly Ala			
	355	360	365	
50	Pro Gly Thr Ser Thr Asn Ser Asp Leu Gln Pro Thr Gly Phe Thr Phe			
	370	375	380	
	Ser Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp			
	385	390	395	400
55	Thr Asp Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg			

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Ser Thr

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25	gcacgggggc accagtggag gaagaggatc cttcttcaa ggtccctgtg aacaagctgg	180
30	cggcagcggt ctccaacttc ggctacgacc tgtaccgcgt gagatccggt gagagccccca	240
35	ccgccaatgt gctgctgtct ccgctcagcg tggccacggc gctctctgcc ctgtcgctgg	300
40	gtgcggaaaca gcggacagaaa tccaaacattc accgggtct gtactacgac ctgatcagta	360
45	acccagacat ccacggcacc tacaaggacc tccttgcctc cgtcaccgccc ccccaagaaga	420
50	accttaagag tgcttccgg attatcttg agaggaagct gcggataaaaa gccagcttca	480
55	tcccacccct ggagaagtca tatggacca ggcccagaat cctgaccggc aactctcgag	540
60	tagaccttca ggagattaac aactgggtgc aggcccagat gaaagggaaa gtcgcttaggt	600
65	ccacgaggga gatgcccaagt gagatcagca ttttcctcct gggcgtggct tacttcaagg	660
70	ggcagtgggt aacaaagttt gactccagaa aaactccct ggaggatttc tacttggatg	720
75	aggagaggac cgtgaaagtc cccatgatgt cagaccctca ggccgtttta cggtacggct	780
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85	tcttcctcct gcctcagaaa gtgaccaga acttgacctt gatagaagag agcctcacct	900
90	ctgagttcat tcatgacata gaccgagaac tgaagactgt tcaggcggtc ctgaccattc	960
95	ccaagctgaa gctgagttat gaaggcgaac tcacgaagtc cgtgcaggag ctgaagctgc	1020
100	aatccctgtt tgatgcacca gactttagca agatcacagg caaacctatc aaacttactc	1080
105	aagtggaaaca tcgcgtcgga tttgagtggatgg atgaggatgg ggccggtaact aactccagcc	1140
110	caggggtcca gcctgcccgc ctcaccttcc ctctggacta tcacctaacc caaccttca	1200
115	tctttgtact gagggacaca gacacagggg cccttctctt cataggcaaa attctggacc	1260
120	ccaggggcac ttagtactcc aactaaatgt tcaaataccc cagaagaaaa aaacactaga	1320
125	gggatggcag attatatatt atacgaaggc tgcccctaca tttcaatgta tactttgcaa	1380
130	taaaaagtgtt ttatccttaa aaaaaaaaaa	1408

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<210> 44
<211> 416
<212> PRT
<213> Bos taurus

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<400> 44

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Met Gln Ala Leu Val Leu Leu Leu Trp Thr Gly Ala Leu Leu Gly Phe
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5 Gly Arg Cys Gln Asn Ala Gly Gln Glu Ala Gly Ser Leu Thr Pro Glu
 20 25 30

10 Ser Thr Gly Ala Pro Val Glu Glu Asp Pro Phe Phe Lys Val Pro
 35 40 45

15 Val Asn Lys Leu Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu Tyr
 50 55 60

Arg Val Arg Ser Gly Glu Ser Pro Thr Ala Asn Val Leu Leu Ser Pro
 65 70 75 80

20 Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln
 85 90 95

25 Arg Thr Glu Ser Asn Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile Ser
 100 105 110

Asn Pro Asp Ile His Gly Thr Tyr Lys Asp Leu Leu Ala Ser Val Thr
 115 120 125

30 Ala Pro Gln Lys Asn Leu Lys Ser Ala Ser Arg Ile Ile Phe Glu Arg
 130 135 140

35 Lys Leu Arg Ile Lys Ala Ser Phe Ile Pro Pro Leu Glu Lys Ser Tyr
 145 150 155 160

40 Gly Thr Arg Pro Arg Ile Leu Thr Gly Asn Ser Arg Val Asp Leu Gln
 165 170 175

Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Val Ala Arg
 180 185 190

45 Ser Thr Arg Glu Met Pro Ser Glu Ile Ser Ile Phe Leu Leu Gly Val
 195 200 205

50 Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr
 210 215 220

Ser Leu Glu Asp Phe Tyr Leu Asp Glu Glu Arg Thr Val Lys Val Pro
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235

240

5 Met Met Ser Asp Pro Gln Ala Val Leu Arg Tyr Gly Leu Asp Ser Asp
 245 250 255

10 Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Thr Ser Ile
 260 265 270

Ile Phe Phe Leu Pro Gln Lys Val Thr Gln Asn Leu Thr Leu Ile Glu
 275 280 285

15 Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu Lys
 290 295 300

20 Thr Val Gln Ala Val Leu Thr Ile Pro Lys Leu Lys Leu Ser Tyr Glu
 305 310 315 320

25 Gly Glu Leu Thr Lys Ser Val Gln Glu Leu Lys Leu Gln Ser Leu Phe
 325 330 335

Asp Ala Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys Leu Thr
 340 345 350

30 Gln Val Glu His Arg Val Gly Phe Glu Trp Asn Glu Asp Gly Ala Gly
 355 360 365

35 Thr Asn Ser Ser Pro Gly Val Gln Pro Ala Arg Leu Thr Phe Pro Leu
 370 375 380

40 Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr Asp
 385 390 395 400

Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly Thr
 405 410 415

45 <210> 45
 <211> 1418
 <212> DNA
 <213> Sus scrofa

50 <400> 45

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5	gcccggagga gggctccccg gcccctgaca cggtgtgggc gccagtggag gaggaggatc	180
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	accgggtct ctactatgac ctgatcagca acccggacct ccacggcacc tacaaggagc	420
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25	agacgtcgct ggaggatttc cacttggatg aggagagaac cgtgaaggtg cccatgatgt	780
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	tcacgaagtc tgtcaggaa ctgaagctgc aatccttgc tgattcacca gacttttagca	1080
35	agatcacggg caaacctatc aaacttactc aagtggaaaca tcgcattggc tttgagtggaa	1140
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<211> 413

<212> PRT

50 <213> Sus scrofa

<400> 46

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20 25 30

10 Thr Val Gly Ala Pro Val Glu Glu Glu Asp Pro Phe Phe Lys Val Pro
35 40 45

15 Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu Tyr
50 55 60

Arg Val Arg Ser Ser Glu Ser Pro Thr Ala Asn Val Leu Leu Ser Pro

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65	70	75	80
5	Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln 85 90 95		
10	Arg Thr Glu Ser Ser Leu His Arg Ala Leu Tyr Tyr Asp Leu Ile Ser 100 105 110		
15	Asn Pro Asp Leu His Gly Thr Tyr Lys Glu Leu Leu Ala Ala Val Thr 115 120 125		
20	Ala Pro Gln Lys Asn Leu Lys Ser Ala Ser Arg Ile Ile Phe Glu Lys 130 135 140		
25	Lys Leu Arg Ile Lys Ala Ser Phe Val Ala Pro Leu Glu Lys Ser Tyr 145 150 155 160		
30	Gly Thr Arg Pro Arg Ile Leu Thr Gly Asn Ser Arg Leu Asp Leu Gln 165 170 175		
35	Glu Val Asn Asn Trp Val Gln Ala Gln Thr Lys Gly Lys Val Ala Arg 180 185 190		
40	Ser Thr Arg Glu Leu Pro Gly Glu Ile Ser Ile Leu Leu Gly Val 195 200 205		
45	Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr 210 215 220		
50	Ser Leu Glu Asp Phe His Leu Asp Glu Glu Arg Thr Val Lys Val Pro 225 230 235 240		
55	Met Met Ser Asp Pro Lys Ala Val Leu Arg Tyr Gly Leu Asp Ser Asp 245 250 255		
60	Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met Ser Ile 260 265 270		
65	Ile Phe Phe Leu Pro Leu Lys Val Thr Gln Asn Leu Thr Met Ile Glu 275 280 285		
70	Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu Lys 290 295 300		
75	Thr Val Gln Ala Val Leu Thr Val Pro Lys Leu Lys Leu Ser Tyr Glu 305 310 315 320		

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Gly Glu Leu Thr Lys Ser Val Gln Glu Leu Lys Leu Gln Ser Leu Phe
325 330 335

5 Asp Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys Leu Thr
340 345 350

10 Gln Val Glu His Arg Ile Gly Phe Glu Trp Asn Glu Asp Gly Gly Ser
355 360 365

15 Ala Thr Ser Ser Pro Gly Pro Arg Leu Thr Phe Pro Leu Asp Tyr His
370 375 380

Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr Asp Thr Gly Ala
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20 Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Ser Thr
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<211> 1317

25 <212> DNA

<213> Ornithorhynchus anatinus

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	gaggaggagg accctttctt caaggtccct gtgaacaagc tggcagccgc cgtctccaac	240
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	<211> 417	
	<212> PRT	
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50	<400> 48	

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20 25 30

10 Ala Thr Gly Thr Ala Val Val Glu Glu Glu Asp Pro Phe Phe Lys Val
35 40 45

15 Pro Val Asn Lys Leu Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu
50 55 60

Tyr Arg Gln Lys Ser Ser Ser Ser Pro Thr Thr Asn Val Leu Leu Ser
65 70 75 80

20 Pro Leu Ser Val Ala Thr Ala Leu Ser Ser Leu Ser Leu Gly Ala Gly
85 90 95

25 Pro Arg Thr Glu Ser Leu Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile
100 105 110

30 His Asn Pro Asp Ile His Gly Thr Tyr Lys Glu Leu Leu Ala Thr Val
115 120 125

35 Thr Ala Pro Gln Lys Asn Leu Lys Thr Ala Ser Arg Leu Val Leu Glu
130 135 140

Arg Lys Leu Arg Ile Lys Ala Gly Phe Val Gly Leu Leu Glu Lys Ser
145 150 155 160

40 Tyr Gly Ser Arg Pro Lys Ile Leu Thr Gly Asn Thr Arg Thr Asp Leu
165 170 175

45

50

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His Glu Met Asn Asn Trp Met Gln Thr Gln Thr Lys Gly Lys Met Gly
 180 185 190

5 Arg Thr Leu Lys Glu Leu Pro Ser Gly Ile Ser Val Leu Leu Leu Gly
 195 200 205

10 Ile Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Pro Lys Lys
 210 215 220

Thr Ser Leu Gln Asp Phe His Leu Asp Glu Asp Arg Thr Val Lys Val
 225 230 235 240

15 Pro Met Met Ser Asp Pro Lys Ala Ile Ile Arg Tyr Gly Leu Asp Ser
 245 250 255

20 Asp Leu Asn Cys Lys Ile Ala Gln Leu Pro Leu Glu Gly Ser Met Ser
 260 265 270

25 Val Ile Phe Phe Leu Pro Leu Lys Ala Thr Gln Asn Leu Thr Leu Ile
 275 280 285

Glu Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu
 290 295 300

30 Lys Thr Ile Gln Ala Val Leu Thr Val Pro Lys Leu Gln Leu Ser Phe
 305 310 315 320

35 Glu Gly Glu Val Ser Lys Thr Phe Gln Glu Ile Lys Leu Gln Ser Leu
 325 330 335

Phe Asn Ser Pro Asp Leu Ser Lys Ile Thr Pro Arg Pro Ile Lys Leu
 340 345 350

40 Thr His Val Val His Arg Ser Ser Leu Glu Trp Ser Glu Asp Gly Val
 355 360 365

45 Gly Asp Ala Pro Ser Pro Ala Leu Leu Pro Ala Arg Leu Thr Phe Pro
 370 375 380

50 Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr
 385 390 395 400

Asp Thr Gly Thr Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly
 405 410 415

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Asn

<210> 49
 <211> 1484
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 <213> Canis lupus familiaris

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<400> 49

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<210> 50
 <211> 396
 <212> PRT
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<400> 50

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Met Arg Ala Ala Pro Lys Asp Ser Pro Ala Pro Asp Ala Thr Gly Val
 1 5 10 15

5 Pro Val Glu Glu Glu Asp Pro Phe Phe Arg Val Pro Val Asn Lys Leu
 20 25 30

10 Ala Ala Ala Ile Ser Asn Phe Gly Tyr Asp Leu Tyr Arg Val Arg Ser
 35 40 45

Ser Phe Ser Pro Ala Ala Asn Val Leu Leu Ser Pro Leu Ser Val Ala
 50 55 60

15 Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln Arg Thr Glu Ser
 65 70 75 80

20 Thr Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile Ser Asn Pro Asp Ile
 85 90 95

25 His Ser Thr Tyr Lys Glu Leu Leu Ala Ser Val Thr Ala Pro Glu Lys
 100 105 110

Asn Phe Lys Ser Ala Ser Arg Ile Val Phe Glu Arg Lys Leu Arg Ile
 115 120 125

30 Lys Ser Ser Phe Val Ala Pro Leu Glu Lys Ser Tyr Ser Thr Arg Pro
 130 135 140

35 Arg Ile Leu Thr Gly Asn Pro Arg Leu Asp Leu Gln Glu Val Asn Asn
 145 150 155 160

40 Trp Val Gln Ala Gln Met Lys Gly Lys Ile Ala Arg Ser Thr Arg Glu
 165 170 175

Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr Ser Leu Glu Asp
 195 200 205

45 Phe His Leu Asp Glu Glu Arg Thr Val Lys Val Pro Met Met Ser Asp
 210 215 220

Pro Lys Ala Ile Leu Arg Tyr Gly Leu Asp Ser Asp Leu Ser Cys Lys
 225 230 235 240

55 Ile Ala Gln Leu Pro Leu Thr Gly Ser Met Ser Ile Ile Phe Phe Leu

245

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255

5 Pro Leu Lys Val Thr Gln Asn Leu Thr Met Ile Glu Glu Ser Leu Thr
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10 Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu Lys Thr Ile Gln Ala
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15 Val Leu Thr Ile Pro Lys Leu Lys Leu Ser Tyr Glu Gly Glu Val Thr
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20 Lys Ser Leu Gln Glu Met Lys Leu Gln Ser Leu Phe Asp Ser Pro Asp
 305 310 315 320

25 Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys Leu Thr Gln Val Glu His
 325 330 335

30 Arg Ala Gly Phe Glu Trp Asn Glu Asp Gly Ala Gly Thr Thr Pro Ser
 340 345 350

35 Pro Gly Leu Gln Pro Thr Arg Leu Thr Phe Pro Leu Asp Tyr His Leu
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40 Asn Arg Pro Phe Ile Phe Val Leu Arg Asp Thr Asp Thr Gly Ala Leu
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<210> 51

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<212> DNA

40 <213> Macaca fascicularis

<400> 51

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	<213> Macaca fascicularis	
	<400> 52	

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10 Pro Asp Ser Thr Gly Ala Leu Val Glu Glu Glu Asp Pro Phe Phe Lys
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15 Val Pro Val Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp
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20 Leu Tyr Arg Val Arg Ser Ser Met Ser Pro Thr Thr Asn Val Leu Leu
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25

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Ser Pro Leu Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala
85 90 95

5 Glu Gln Arg Thr Glu Ser Val Ile His Arg Ala Leu Tyr Tyr Asp Leu
100 105 110

10 Ile Ser Ser Pro Asp Ile His Gly Thr Tyr Lys Glu Leu Leu Gly Thr
115 120 125

Val Thr Ala Pro Gln Lys Asn Leu Lys Ser Ala Ser Arg Ile Val Phe
130 135 140

15 Glu Lys Lys Leu Arg Ile Lys Ser Ser Phe Val Ala Pro Leu Glu Lys
145 150 155 160

20 Ser Tyr Gly Thr Arg Pro Arg Val Leu Thr Gly Asn Pro Arg Leu Asp
165 170 175

Leu Gln Glu Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Leu
180 185 190

25 Ala Arg Ser Thr Lys Glu Leu Pro Asp Glu Ile Ser Ile Leu Leu Leu
195 200 205

30 Gly Val Ala Tyr Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Pro Arg
210 215 220

35 Lys Thr Ser Leu Glu Asp Phe His Leu Asp Glu Glu Arg Thr Val Arg
225 230 235 240

Val Pro Met Met Ser Asp Pro Lys Ala Ile Leu Arg Tyr Gly Leu Asp
245 250 255

40 Ser Asp Leu Ser Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Met
260 265 270

45 Ser Ile Ile Phe Phe Leu Pro Leu Lys Val Thr Gln Asn Leu Thr Leu
275 280 285

Ile Glu Glu Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu
290 295 300

50 Leu Lys Thr Val Gln Ala Val Leu Thr Leu Pro Lys Leu Lys Leu Ser
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55 Tyr Glu Gly Glu Val Thr Lys Ser Leu Gln Glu Thr Lys Leu Gln Ser
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Leu Phe Asp Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys
 340 345 350

5 Leu Thr Gln Val Glu His Arg Ala Gly Phe Glu Trp Asn Glu Asp Gly
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10 Ala Gly Ala Thr Pro Ser Pro Gly Leu Gln Pro Ala His Leu Thr Phe
 370 375 380

15 Leu Leu Asp Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp
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Thr Asp Thr Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg
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<210> 53

<211> 1935

<212> DNA

25 <213> Pan troglodytes

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<210> 54

<211> 415

<212> PRT

<213> Pan troglodytes

<400> 54

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45	Thr Gly Ala Leu Val Glu Glu Glu Asp Pro Phe Phe Lys Val Pro Val	
	35 40 45	
50	Asn Lys Leu Ala Ala Ala Val Ser Asn Phe Gly Tyr Asp Leu Tyr Arg	
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55	Val Arg Ser Ser Met Ser Pro Thr Thr Asn Val Leu Leu Ser Pro Leu	
	65 70 75 80	

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Ser Val Ala Thr Ala Leu Ser Ala Leu Ser Leu Gly Ala Glu Gln Arg
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5 Thr Glu Ser Ile Ile His Arg Ala Leu Tyr Tyr Asp Leu Ile Ser Ser
100 105 110

10 Pro Asp Ile His Gly Thr Tyr Lys Glu Leu Leu Asp Thr Val Thr Ala
115 120 125

Pro Gln Lys Asn Leu Lys Ser Ala Ser Arg Ile Val Phe Glu Lys Lys
130 135 140

15 Leu Arg Ile Lys Ser Ser Phe Val Ala Pro Leu Glu Lys Ser Tyr Gly
145 150 155 160

20 Thr Arg Pro Arg Val Leu Thr Gly Asn Pro Arg Leu Asp Leu Gln Glu
165 170 175

Ile Asn Asn Trp Val Gln Ala Gln Met Lys Gly Lys Leu Ala Arg Ser
180 185 190

25 Thr Lys Glu Ile Pro Asp Glu Ile Ser Ile Leu Leu Leu Gly Val Ala
195 200 205

His Phe Lys Gly Gln Trp Val Thr Lys Phe Asp Ser Arg Lys Thr Ser
210 215 220

30 Leu Glu Asp Phe His Leu Asp Glu Glu Arg Thr Val Arg Val Pro Met
225 230 235 240

35 Met Ser Asp Pro Lys Ala Val Leu Arg Tyr Gly Leu Asp Ser Asp Leu
245 250 255

40 Ser Cys Lys Ile Ala Gln Leu Pro Leu Thr Gly Ser Thr Ser Ile Ile
260 265 270

Phe Phe Leu Pro Leu Lys Val Thr Gln Asn Leu Thr Leu Ile Glu Glu
45 275 280 285

50 Ser Leu Thr Ser Glu Phe Ile His Asp Ile Asp Arg Glu Leu Lys Thr
290 295 300

Val Gln Ala Val Leu Thr Val Pro Lys Leu Lys Leu Ser Tyr Glu Gly
305 310 315 320

55 Glu Val Thr Lys Ser Leu Gln Glu Met Lys Leu Gln Ser Leu Phe Asp
325 330 335

Ser Pro Asp Phe Ser Lys Ile Thr Gly Lys Pro Ile Lys Leu Thr Gln
 340 345 350

5 Val Glu His Arg Ala Gly Phe Glu Trp Asn Glu Asp Gly Ala Gly Thr
 355 360 365

10 Thr Pro Ser Pro Gly Leu Gln Pro Ala His Leu Thr Phe Pro Leu Asp
 370 375 380

Tyr His Leu Asn Gln Pro Phe Ile Phe Val Leu Arg Asp Thr Asp Thr
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15 Gly Ala Leu Leu Phe Ile Gly Lys Ile Leu Asp Pro Arg Gly Thr
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20 <210> 55

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<212> DNA

<213> Macaca mulatta

25 <400> 55

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Claims

- 20 1. An agonist of the OA1 receptor for use in treating or preventing age-related macular degeneration (AMD), wherein the agonist of the OA1 receptor is L-DOPA.

25 **Patentansprüche**

- 26 1. Agonist des OA1-Rezeptors zum Gebrauch beim Behandeln oder Verhindern von altersbedingter Makuladegeneration (AMD), wobei der Agonist des OA1-Rezeptors L-DOPA ist.

30 **Revendications**

- 31 1. Agoniste du récepteur OA1 pour une utilisation dans le traitement ou la prévention de la dégénérescence maculaire liée à l'âge (DMLA), où l'agoniste du récepteur OA1 est L-DOPA.

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Figure 1

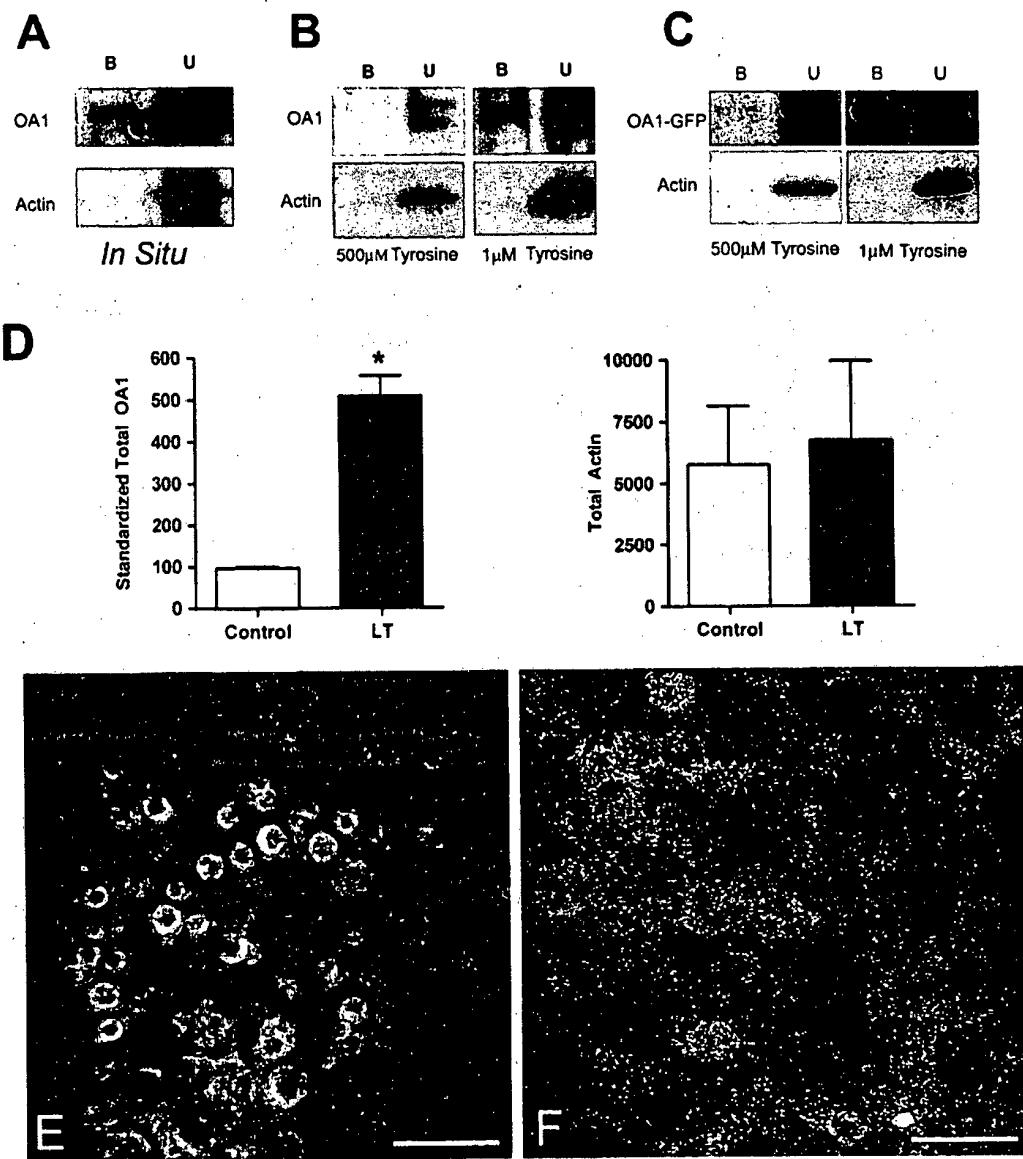
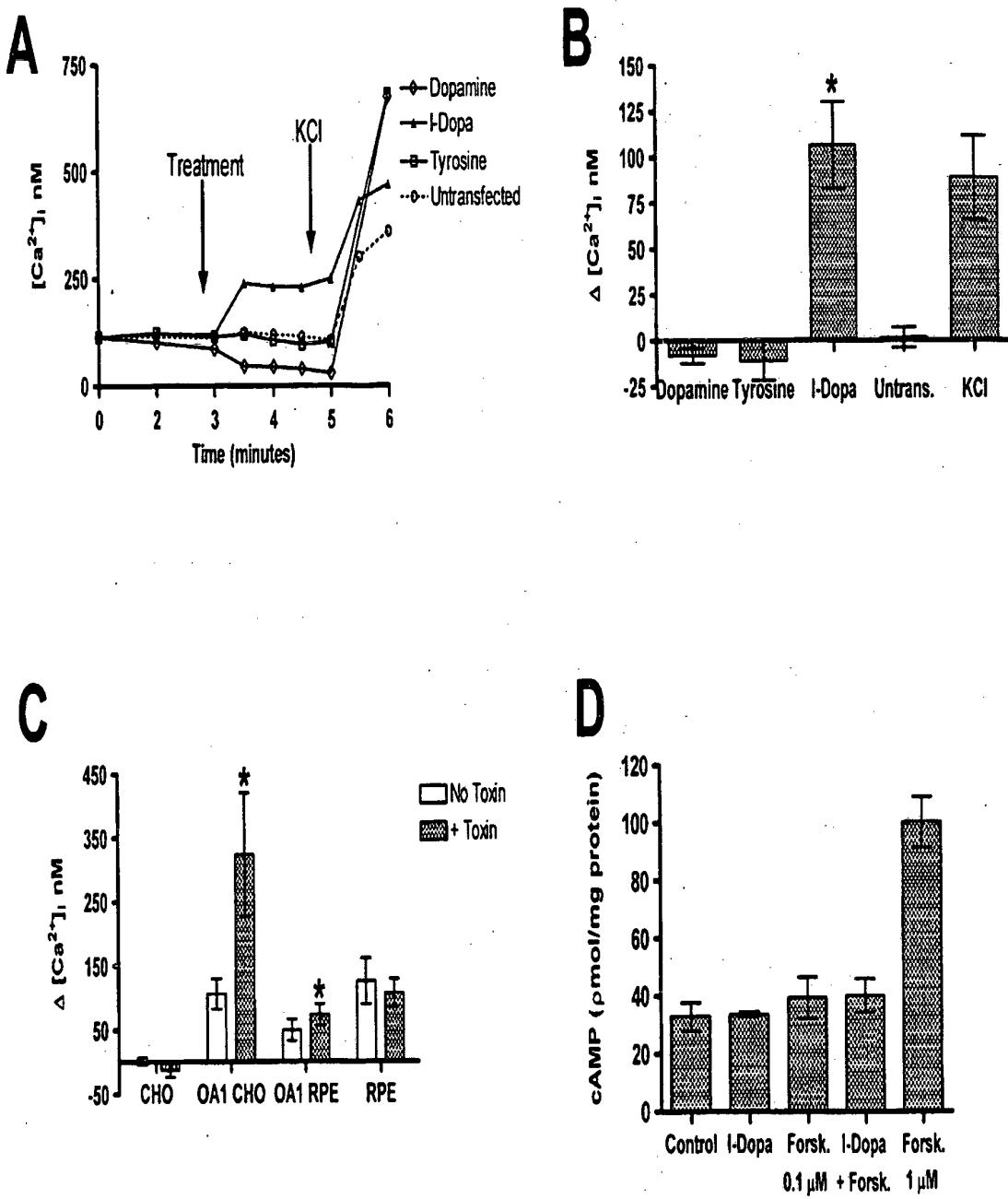
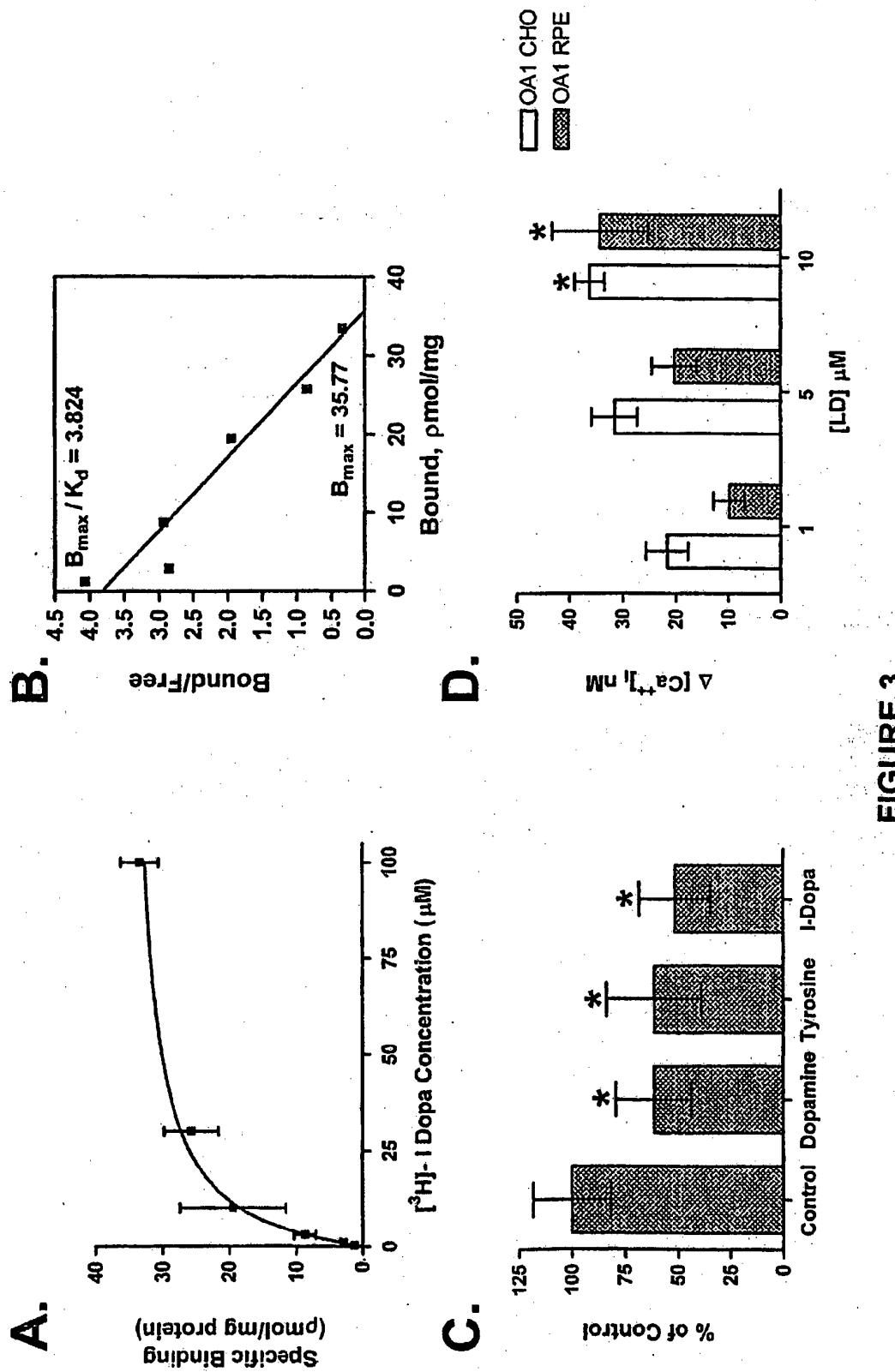


Figure 2



**FIGURE 3**

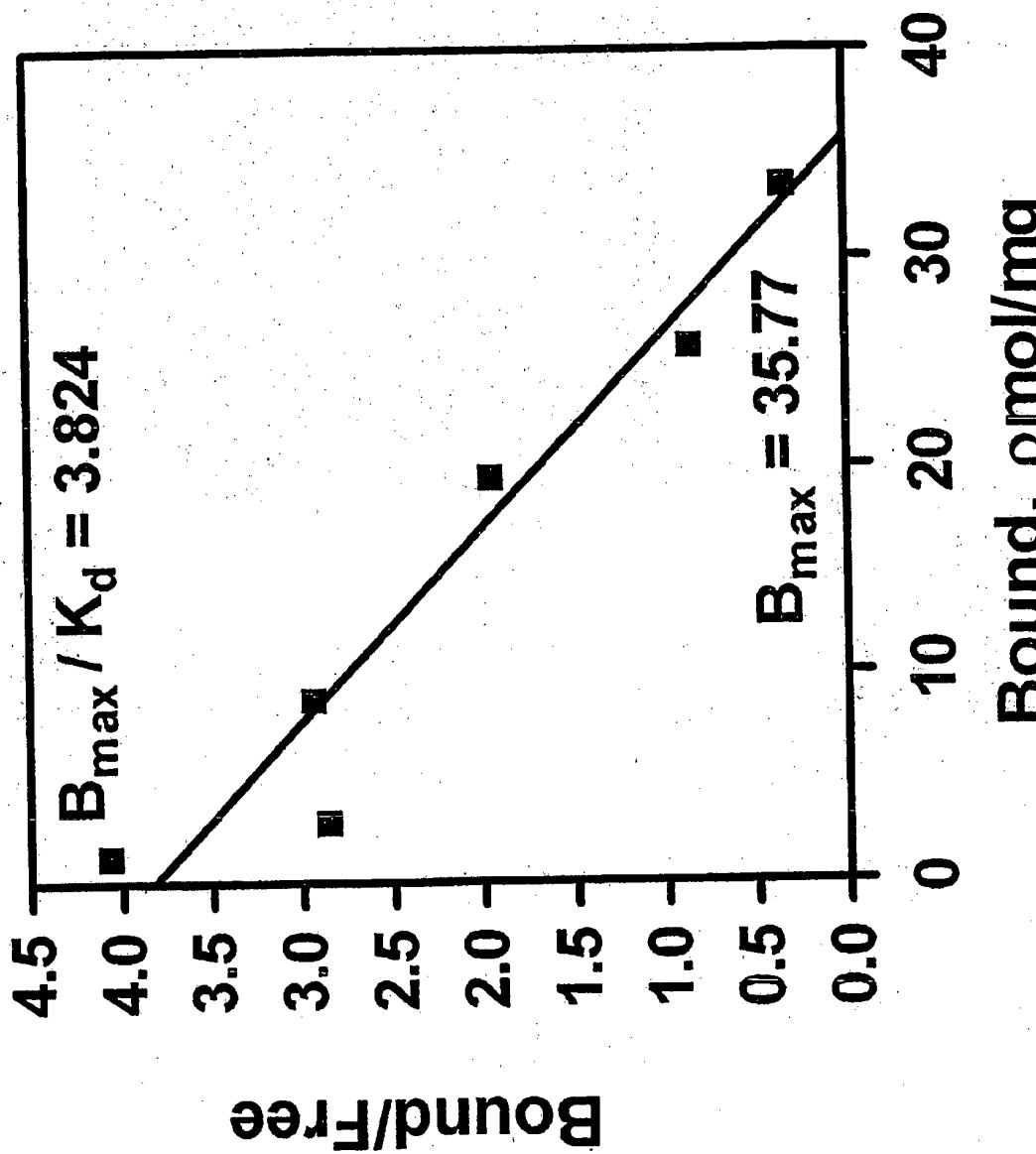


FIGURE 3E

Figure 4

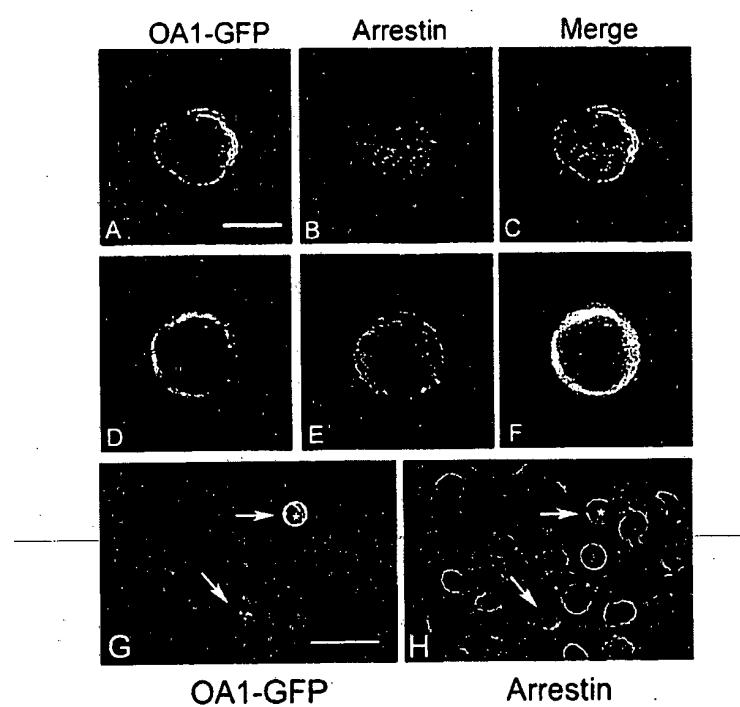


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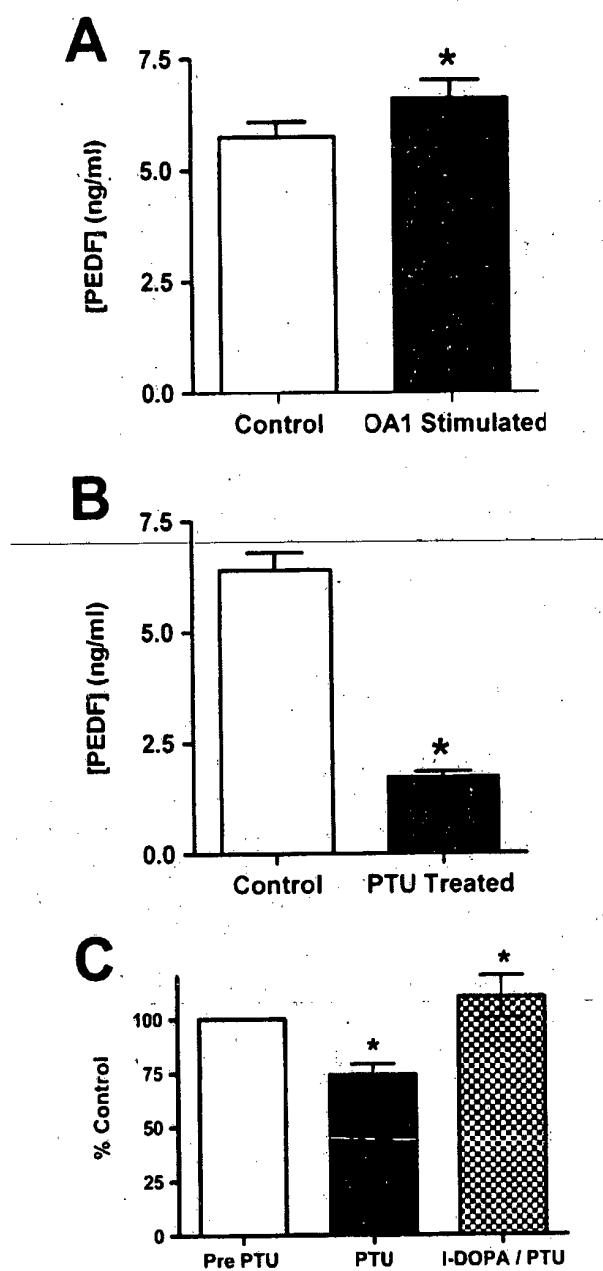


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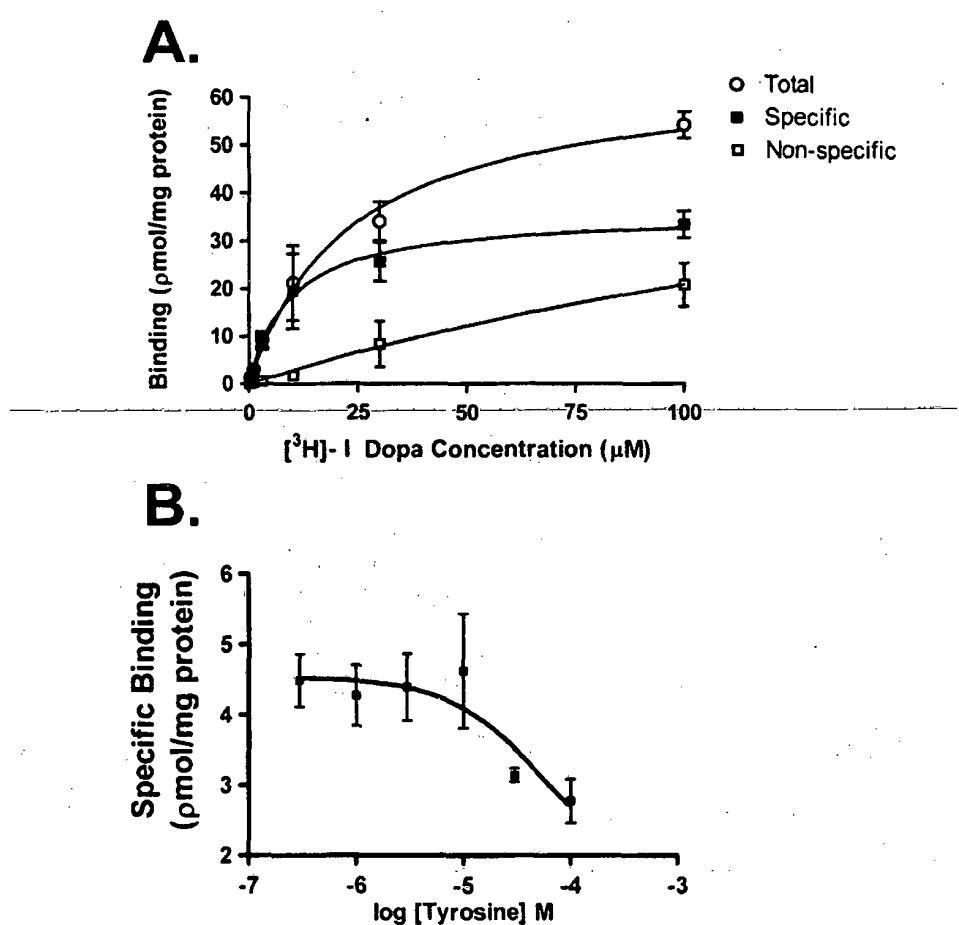


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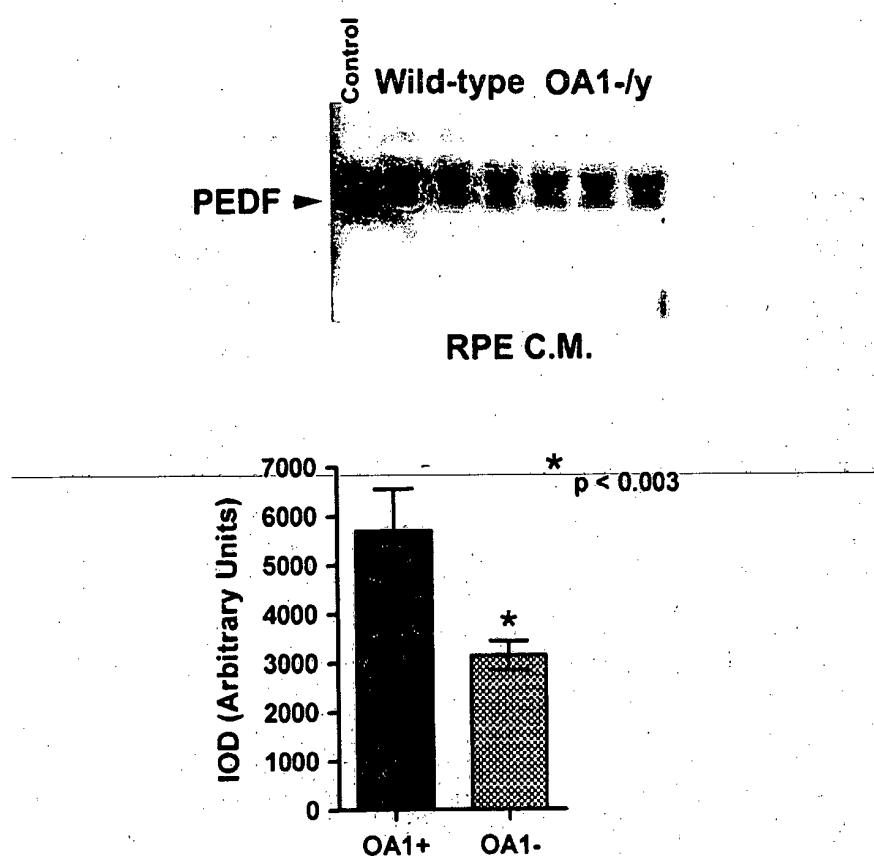
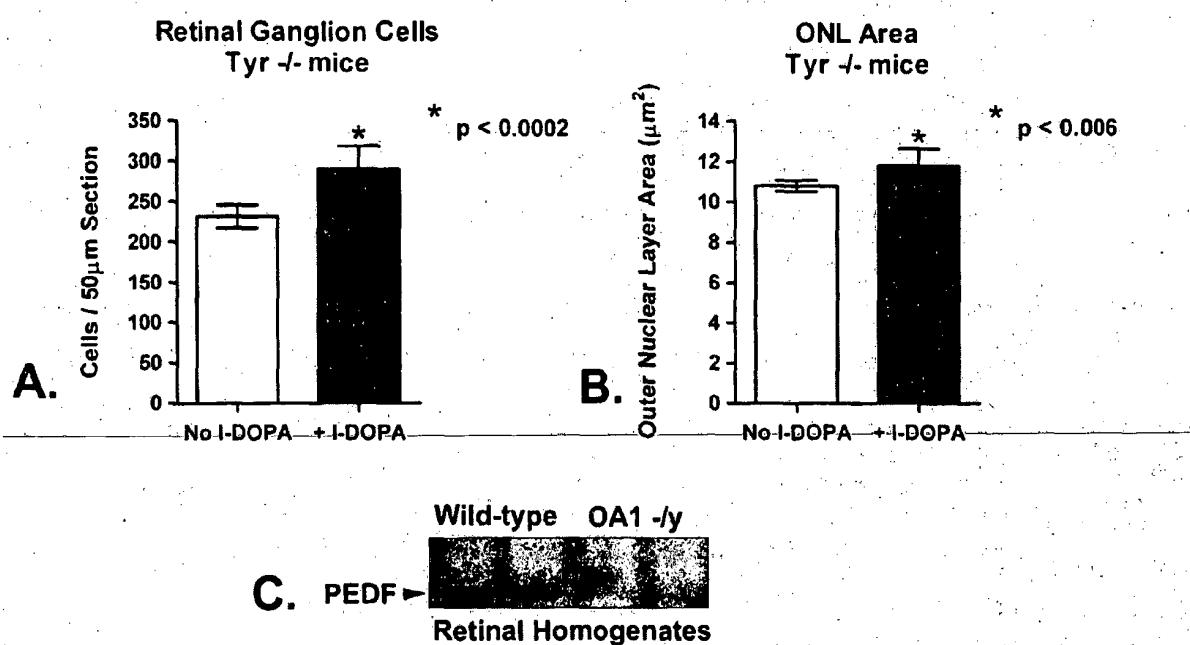


Figure 8



REFERENCES CITED IN THE DESCRIPTION

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